

## **Screening of Drugs in Saliva**

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## **Editorial**

Saliva is an interesting medium for analytical research due to the ease of collection and sample preparation prior to analysis. On the other hand, many drugs diffuse through the salivary tissues and can be detected in the salivary fluidat concentrations similar to blood plasma [1,2]. Therapeutic drug monitoring and clinical pharmacokinetics studies are intensively conducted on blood samples withdrawn whether from healthy or patient subjects who is invasive mean of sampling and might be inappropriate for uncooperative subjects like children and elderly. Moreover, the collection of blood samples requires special facilities and highly trained personnel. Saliva provides fast and noninvasive sampling besides being the best choice for efficient screening of illicit drugs that are either ingested (ex. Alcohols) or smoked (i.e., cocaine, PCP, methamphetamine, and marijuana) following recent use [3]. However, the quantitative aspects for drug analysis in saliva may still be under question as the salivary drug levels are affected by different factors like salivary pH. protein binding and the pKa values which may vary from one subject to another according to the nutritional habits, time of collection regarding ingestion of food, ingestion of other drugs or the bacterial content [4]. Screening of drugs in saliva is to be very useful in case of investigation of illegal drug use with emergency room admission, as serum and urine screening methods cannot be convenient from potentially uncooperative patients. Thus, saliva samples as a rapid and minimally invasive analytical technique can introduce the optimum alternative. The composition of salivary fluid, containing 99.5% water, facilitates the analytical processes and sample preparation [5]. Saliva exhibits only the free fraction of a drug as the protein bound fraction cannot diffuse to the salivary tissues as well as both the parent compound and metabolites can be detected which may give a clear demonstration of the drug profile after administration. Saliva can be collected easily using adsorbent pads placed in the oral cavity to collect pooled saliva for 1-2 minutes then compressed against the collecting container [4].

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A study conducted on Cocaine has shown that the saliva levels of cocaine and its metabolite are significantly correlated with plasma concentrations. Moreover, the ratio of the metabolite/parent in salivary fluid can indicate the time of last use [6,7]. A study have shown that Cocaine detected in saliva reached concentrations up to 4.9 times higher than in urine and plasma. Cocaine was detected in all saliva samples while urine and serum samples from same subject had no detectable levels [8].

Methamphetamine and amphetamine can be detected rapidly in plasma and oral fluidafter administration. It was reported that the concentration levels of amphetamine methamphetamine to in saliva werehigher than plasma concentrations bv approximately four times [9]. On the other hand, Amphetamine and p-hydroxymethamphetamine are metabolites of methamphetamine and their presence in oral fluid can indicate the substantiate use [10]. MDMA or 3,4-Methylenedioxymethamphetamine reaches the peak saliva and plasma concentration following oral administration to humans after 1.5 h of drug administration [11]. The concentrations of MDMA in salivary fluid were higher than in plasma. However, salivary concentrations of MDMA were found highly

correlated with plasma. The detection of MDMA at high concentrations in human saliva after intake was related to thelow plasma-protein binding of MDMA [5,11].

Delta-9-tetrahydrocannabinol (THC) the major constituent of cannabis was detected in oral fluid after recent smoked and orally ingested cannabis [12-14]. On the other hand, a rapid decline was shown in THC oral fluid concentration similarly to plasma concentrations [15]. The high THC salivary concentrations have been attributed to the high deposition in the oral cavity following the intake, rather than from transfer from blood.

Techniques such as HPLC-MS/MS and GC-MS/MS are very useful in the detection of trace levels of drugs in saliva with broad application to different types of samples (biological or environmental). Minor sample preparation procedures are required in case of saliva samples. Micro-extraction by packed sorbent (MEPS) is a miniaturized technique that can provide highly efficientextraction of target analytes using a programmable automatic pipette eVol® which facilitate handling and maintain the reproducibility of the method [4,16].

In conclusion, Saliva can be a real alternative to serum and urine analyses regarding investigation of recent illicit drug use and in a non-invasive manner that applies suitably for all kinds of subjects as well as drugrelated emergency admission.

## References

- Drummer OH (2005) Review: Pharmacokinetics of illicit drugs in oral fluid, Forensic Sci Int 150: 133-142.
- 2. Dana K, Shende C, Huang H, Farquharson S (2015) Rapid Analysis of Cocaine in Saliva by Surface-Enhanced Raman Spectroscopy, Journal of analytical & bioanalytical techniques 6: 1-5.
- 3. Crouch DJ, Day J, Baudys J, Fatah AA (2005) Evaluation of Saliva/Oral Fluid as an Alternate Drug Testing Specimen, U.S. Department of Justice.
- 4. Elmongy H, Abdel-Rehim M (2016) Saliva as an alternative specimen to plasma for drug bioanalysis. A review, TrAC Trends in Analytical Chemistry 83: 70-79.
- Cone EJ, Huestis MA (2007) Interpretation of Oral Fluid Tests for Drugs of Abuse, Ann N Y Acad Sci 1098: 51-103.

- 6. Scheidweiler KB, Spargo EA, Kelly TL, Cone EJ, Barnes AJ, et al. (2010) Pharmacokinetics of cocaine and metabolites in human oral fluid and correlation with plasma concentrations after controlled administration, Ther Drug Monit 32(5): 628-637.
- Dams R, Choo RE, Lambert WE, Jones H, Huestis MA (2007) Oral fluid as an alternative matrix to monitor opiate and cocaine use in substance-abuse treatment patients. Drug Alcohol Depend 87: 258-267.
- 8. Schramm W, Craig PA, Smith RH, Berger GE (1993) Cocaine and benzoylecgonine in saliva, serum, and urine. Clin Chem 39: 481-487.
- 9. Schepers RJ, Oyler JM, Joseph RE, Cone EJ, Moolchan ET, et al. (2003) Methamphetamine and amphetamine pharmacokinetics in oral fluid and plasma after controlled oral methamphetamine administration to human volunteers, Clin Chem 49: 121-132.
- 10. Caldwell J, Dring LG, Williams RT (1972) Metabolism of (14 C)methamphetamine in man, the guinea pig and the rat, Biochem J 129: 11-22.
- 11. Navarro M, Pichini S, Farre M, Ortuno J, Roset PN, et al. (2001) Usefulness of saliva for measurement of 3,4-methylenedioxymethamphetamine and its metabolites: correlation with plasma drug concentrations and effect of salivary pH. Clin Chem 47: 1788-1795.
- 12. Niedbala RS, Kardos KW, Fritch DF, Kunsman KP, Blum KA, et al. (2005) Passive cannabis smoke exposure and oral fluid testing. II. Two studies of extreme cannabis smoke exposure in a motor vehicle. J Anal Toxicol 29: 607-615.
- 13. Niedbala S, Kardos K, Salamone S, Fritch D, Bronsgeest M, et al. (2004) Passive cannabis smoke exposure and oral fluid testing. J Anal Toxicol 28: 546-552.
- 14. Niedbala RS, Kardos KW, Fritch DF, Kardos S, Fries T, et al. (2001) Detection of marijuana use by oral fluid and urine analysis following single-dose administration of smoked and oral marijuana. J Anal Toxicol 25: 289-303.
- 15. Huestis MA, Cone EJ (2004) Relationship of Delta 9tetrahydrocannabinol concentrations in oral fluid and plasma after controlled administration of smoked cannabis. J Anal Toxicol 28: 394-399.

16. Elmongy H, Ahmed H, Wahbi AA, Amini A, Colmsjo A, et al. (2016) Determination of metoprolol enantiomers in human plasma and saliva samples utilizing microextraction by packed sorbent and liquid chromatography-tandem mass spectrometry, Biomedical Chromatography 30: 1309-1317.