



Amyloid B-Protein Aggregation at Physiologically Relevant Concentrations. A Critical Role of Membranes

Lyubchenko YL*

Department of Pharmaceutical Sciences, University of Nebraska Medical Center, USA

*Corresponding author: Yuri L Lyubchenko, Department of Pharmaceutical Sciences, University of Nebraska Medical Center, USA, 986025 Nebraska Medical Center, Omaha, NE 68198, USA, Tel: 1-402-559-1971; Email: ylyubchenko@unmc.edu

Hypothesis Paper

Volume 3 Issue 1

Received Date: September 28, 2020

Published Date: October 28, 2020

Abstract

Background: The aggregation of amyloid beta (Ab) is a self-assembly process that results in the production of fibrillar structures along with neurotoxic aggregates. However, in the vast majority studies in vitro the required Ab concentrations is several orders higher of the physiological relevant concentrations of A β ; no aggregation is observed at physiological low nanomolar range of A β . This suggests that the assembly of A β in aggregates in vivo utilizes pathways different from those used in experiments in vitro.

Results: The spontaneous assembly of A β oligomers within the physiologically relevant concentration range can occur, but it is the on-surface aggregation mechanism, in which the surface plays a role of the catalyst of the aggregation process. The model for the on-surface aggregation process suggests that the self-assembly of A β oligomers is initiated by the interaction of amyloid proteins with the cellular membrane. The membrane catalyzes amyloid aggregation by stabilizing an aggregation-prone conformation of amyloids. The lipid composition contributes to the membrane-mediated misfolding and aggregation of A β monomers.

Conclusion: Membrane-mediated aggregation catalysis explains a number of observations associated with the development of AD. The affinity of A β monomers to the membrane surface is the major factor defining the aggregation process rather than A β concentration. According to the model, the development of potential preventions for the interaction of monomeric amyloids with membrane can help control the aggregation process. This is a paradigm change for the development of efficient treatments, early diagnostics, and preventions for Alzheimer's disease.

Keywords: Amyloid beta; Membrane composition; Oligomers; Alzheimer's disease

Abbreviations: AD: Alzheimer's disease; A β : amyloid b proteon; PD: Parkinson's disease; ACH: amyloid cascade hypothesis.

Introduction

Amyloid Cascade Hypothesis

The involvement of protein aggregates in the

development of protein misfolding diseases, including *Alzheimer's disease (AD)* and *Parkinson's disease (PD)*, among others, is supported by numerous data and the formation of deposits (plaques) in the brain [1]. Numerous physical, chemical, and structural data reveal a spontaneous assembly of amyloidogenic proteins into aggregates, and the amyloid cascade hypothesis (ACH), proposed more than a quarter-century years ago, and is the major model used to describe the pathology of AD and other neurodegenerative diseases

[2-6]. ACH posits that the onset of diseases involves the spontaneous assembly of an amyloidogenic polypeptide. In turn, accumulation of aggregates defines the disease state. Translational studies in the framework of ACH are focused on decreasing the concentration of amyloid proteins to decelerate the aggregation process [2,3,7]. However, drug development based on decreased A β concentration, as well as disaggregating the plaques, has failed [1,8], which challenges the validity of ACH. Indeed, in the monomeric state, all amyloidogenic proteins are functionally important and the findings in Hillen H, et al. [9-11] point to neuroprotective features of monomeric A β . Therefore, approaches focused on the decreased concentration of amyloids can impair positive functional roles of amyloid proteins. In fact, there are a number of problems with ACH, and the most challenging is the concentration of A β [3]. Specifically, *in vitro* aggregation experiments require A β concentrations in the micromolar range, whereas A β levels in brain and cerebral spinal fluid (CSF) are frequently in the low nanomolar range [7,12-15]; this value remains in the same range regardless of the disease state [16,17]. No aggregation of A β and other amyloidogenic proteins occurs *in vitro* at such low concentrations. At the same time, plaques are formed *in vivo*, suggesting that A β aggregation, regardless of a low concentration, does occur, but the mechanism allowing for the amyloid protein to aggregate is unknown.

Ab Aggregation at Physiologically Relevant Concentrations

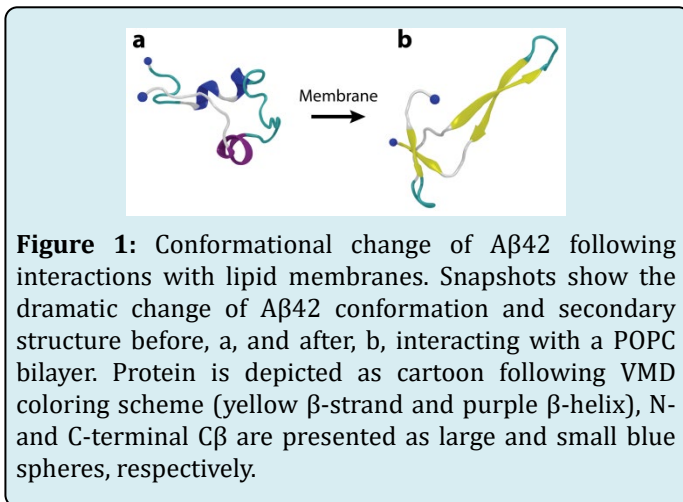
It was found recently by the direct observations of the aggregation process with atomic force microscopy (AFM) that a spontaneous assembly of A β monomers into aggregates can be observed at physiologically low concentrations if aggregation takes place at surfaces [18]. The process occurs at ambient conditions, physiological pH values, and without agitation, which is often used to stimulate the aggregation. These findings suggest that the surface plays a role of a catalyst and a model for the surface mediated catalysis has been proposed and tested in Pan Y, et al. [19]. According to this model, aggregation starts with protein monomers transiently attaching to the surface due to molecular interactions. This process increases the local concentration of proteins, which in turn increases the probability of oligomerization reactions to occur on the surface. These results are in line with Lindberg DJ, et al. [20] that reported catalytic properties of the zwitterionic lipid vesicles during the formation of A β 42 fibrils. Note previous publications in which catalytic property of surfaces in amyloid fibrils formation has been discussed [21-24]. The catalytic effect of surfaces in amyloid aggregation explains the experiments on aggregation of A β 40 at low nanomolar concentrations in

cell culture [25]. Local intracellular compartmentalization including the formation of proteinaceous membrane-less organelles is considered as a potential mechanism alleviating the problem with the overall low concentration of amyloids; however, it is considered as a complement to effects of membranes and membrane-encapsulated organelles [26].

Ab Misfolding and Interaction with Membranes

Recent time-lapse AFM experiments provided direct evidence for the A β catalysis by phospholipid bilayers, which are models for cell membranes [27-30]. Importantly, the computer modeling revealed that at the bilayer surface A β dramatically changes conformation. As is illustrated in Figure 1, overall unstructured A β 42 monomer (Figure 1a) adopts conformations with extended β -sheet segments (yellow arrows in Figure 1b). The β -sheets are characteristic features for A β 42 fibrils, suggesting that the membrane-bound A β 42 monomer adopts the aggregation prone, misfolded conformation. Indeed, the simulations in Banerjee S, et al. [28] showed that dimerization on the bilayer surface was rapid, although the dimer interacts with the lipid surfaces transiently. This finding is in line with the experiments according to which the aggregates assembled on the surface can dissociate into the bulk solution [27-30]. These features are not limited to A β interaction with the bilayers. Similar results were obtained with β -syn [27-30]. Combined experimental and computer modeling studies led to the hypothesis that the on-surface amyloid misfolding is the mechanism by which the disease-related aggregation nuclei are formed, providing the seeds for aggregation [27,28]. Inside the brain, the surface-assembled oligomers can induce neurotoxic effects induced by binding to specific receptors and phosphorylation of the tau protein to initiate its misfolding and aggregation, among other problems [15]. According to the AFM observations, the vast majority of cases, the aggregates formed on the surface are oligomers, which are considered to be the most neurotoxic amyloid aggregates [3,4,15,31]. Thus, these observations suggest that interaction of amyloid proteins with cellular membrane can be the mechanism by which amyloid aggregation can be initiated *in vivo* at the physiological concentration range. According to the model proposed in Banerjee S, et al. [28], the aggregation of Ab is a stepwise process, which starts with the conformation change of the monomer upon interaction with the membrane. Therefore, another monomer, after the induced conformational change, assembles with the first into a dimer followed by the growth of the oligomer. The aggregate can dissociate from the surface and initiate neurotoxic effects via different pathways [3,32-40]. This model is supported by a very recent publication [41] in which accelerated aggregation of A β 42 on membranes of

neuroblastoma cell was visualized.



Membrane Composition and the Aβ Self-Assembly Process

Cell membranes consist of a large number of lipids suggesting that the lipid composition can be a factor contributing to the on-membrane aggregation of amyloids. Indeed, recent publications revealed the role of such lipids as cholesterol, sphingomyelins and gangliosides on the formation of Aβ fibrils on membrane surfaces [42-44]. Time lapse AFM studies demonstrated that even the ratio between phospholipids changes the aggregation propensity of supported bilayers for Aβ42 and more strongly for α-synuclein [27,29]. Very recent publication [45] demonstrated that cholesterol in the lipid bilayer significantly enhances the aggregation process of Aβ42 at nanomolar monomer concentration. Importantly, computer modeling showed that Aβ42 has an elevated affinity to cholesterol-containing membranes adopting a set of aggregation-prone conformations.

Membrane Catalysis of Amyloid Aggregation and Early Stages of the Disease Development

Recent studies with the physiologically relevant concentrations of Aβ proteins lead to a concept on a critical role of membranes in triggering the aggregation process and hence, the disease state. Within this concept, the membrane composition is a factor controlling the aggregation process, so the change in membrane composition can shift the ratio between monomeric and aggregated states of Aβ towards aggregated ones, which define the disease state. The findings on contribution of cholesterol, sphingomyelins and gangliosides to neurotoxic effects of Aβ aggregates [15,46,47] makes these lipids as prime candidates, suggesting that their content in cell membranes can be the disease defining

parameter.

A number of observations can be explained in the framework of the membrane composition concept. Note the role of cholesterol in AD pathogenesis and specifically hypercholesterolemia as a risk factor in AD [48-50]. Catalysis of the aggregation process by cholesterol explains these observations. Additionally, the role of diet in controlling the AD development is widely discussed [51-54]. Note a recent article [54] in which the link between the AD biomarkers and a protective dietary pattern (Mediterranean-style low fat diet) is reviewed. Contributions of the low-lipid diet to the lipid composition of membranes can explain this effect.

In conclusion, a novel concept explains the self-assembly of amyloid proteins in the disease-prone aggregates at physiologically relevant concentrations. It also explains a number of observations associated with the development of AD. The affinity of Aβ monomers to the membrane surface is the major factor defining the aggregation process rather than Aβ concentration. Therefore, development of therapeutics need not focus on the change of concentration of Aβ monomers. This is another important feature of the membrane aggregation concept as the change of the monomers concentrations can impair their functional roles. It is very likely that the membrane aggregation concept can be extended to other neurodegenerative diseases.

Acknowledgements

This research was funded by National Institutes of Health, grants GM096039 and GM118006 to YLL.

Authors Contribution

YLL designed and supervised the research and wrote the manuscript.

Competing Interests

The author has no competing interests.

References

1. Hardy J, De Strooper B (2017) Alzheimer's disease: where next for anti-amyloid therapies? *Brain* 140(4): 853-855.
2. Mohamed T, Shakeri A, Rao PPN (2016) Amyloid cascade in Alzheimer's disease: Recent advances in medicinal chemistry. *European Journal of Medicinal Chemistry* 113: 258-272.
3. Armstrong RA (2014) A critical analysis of the amyloid

- cascade hypothesis. *Folia Neuropathologica* 52(3): 211-225.
4. Hardy J (2006) Has the amyloid cascade hypothesis for Alzheimer's disease been proved? *Curr Alzheimer Res* 3(1): 71-73.
 5. Hardy J (2006) Alzheimer's disease: the amyloid cascade hypothesis: an update and reappraisal. *J Alzheimers Dis* 9: 151-153.
 6. Hardy JA, Higgins GA (1992) Alzheimer's disease: the amyloid cascade hypothesis. *Science* 256(5054): 184-185.
 7. McGeer PL, McGeer EG (2013) The amyloid cascade-inflammatory hypothesis of Alzheimer disease: implications for therapy. *Acta Neuropathol* 126(4): 479-497.
 8. Abbott A, Dolgin E (2016) Leading Alzheimer's theory survives drug failure. *Nature* 540: 15-16.
 9. Hillen H (2019) The Beta Amyloid Dysfunction (BAD) Hypothesis for Alzheimer's Disease. *Front Neurosci* 13: 1154.
 10. Bate C, Williams A (2018) Monomeric amyloid- β reduced amyloid- β oligomer-induced synapse damage in neuronal cultures. *Neurobiology of Dis* 111: 48-58.
 11. Giuffrida ML, Caraci F, Pignataro B, Cataldo S, De Bona P, et al. (2009) Beta-amyloid monomers are neuroprotective. *J Neurosci* 29(34): 10582-10587.
 12. An SSA, Lee BS, Yu JS, Lim K, Kim GJ, et al. (2017) Dynamic changes of oligomeric amyloid beta levels in plasma induced by spiked synthetic A β 42. *Alzheimers Res Ther* 9(1): 86.
 13. Bjerke M, Portelius E, Minthon L, Wallin A, Anckarsater H, et al. (2010) Confounding factors influencing amyloid Beta concentration in cerebrospinal fluid. *Int J Alzheimers Dis*, pp: 986310.
 14. Copani A (2017) The underexplored question of β -amyloid monomers. *European Journal of Pharmacology* 817: 71-75.
 15. Cline EN, Bicca MA, Viola KL, Klein WL (2018) The Amyloid-beta Oligomer Hypothesis: Beginning of the Third Decade. *J Alzheimers Dis* 64(1): S567-S610.
 16. Potter R, Patterson BW, Elbert DL, Ovod V, Kasten T, et al. (2013) Increased in vivo amyloid-beta42 production, exchange, and loss in presenilin mutation carriers. *Sci Transl Med* 5(189).
 17. Palmqvist S, Scholl M, Strandberg O, Mattsson N, Stomrud E, et al. (2017) Earliest accumulation of beta-amyloid occurs within the default-mode network and concurrently affects brain connectivity. *Nat Commun* 8(1): 1214.
 18. Banerjee S, Hashemi M, Lv Z, Maity S, Rochet JC, et al. (2017) A novel pathway for amyloids self-assembly in aggregates at nanomolar concentration mediated by the interaction with surfaces. *Sci Rep* 7: 45592.
 19. Pan Y, Banerjee S, Zagorski K, Shlyakhtenko LS, Kolomeisky AB, et al. (2020) Molecular Model for the Surface-Catalyzed Protein Self-Assembly. *J Phys Chem B* 124(2): 366-372.
 20. Lindberg DJ, Wesén E, Björkeröth J, Rocha S, Esbjörner EK (2017) Lipid membranes catalyse the fibril formation of the amyloid- β (1-42) peptide through lipid-fibril interactions that reinforce secondary pathways. *Biochimica et Biophysica Acta (BBA) – Biomembranes* 1859(10): 1921-1929.
 21. Cebecauer M, Hof M, Amaro M (2017) Impact of GM1 on Membrane-Mediated Aggregation/Oligomerization of beta-Amyloid: Unifying View. *Biophys J* 113(6): 1194-1199.
 22. Korshavn KJ, Satriano C, Lin Y, Zhang R, Dulchavsky M, Bhunia A, et al. (2017) Reduced Lipid Bilayer Thickness Regulates the Aggregation and Cytotoxicity of Amyloid-beta. *J Biol Chem* 292(11): 4638-4650.
 23. Khondker A, Alsop RJ, Rheinstadter MC (2017) Membrane-Accelerated Amyloid-beta Aggregation and Formation of Cross-beta Sheets 7(3): 49.
 24. Gorbenko GP, Kinnunen PK (2006) The role of lipid-protein interactions in amyloid-type protein fibril formation. *Chem Phys Lipids* 141: 72-82.
 25. Podlisny MB, Walsh DM, Amarante P, Ostaszewski BL, Stimson ER, et al. (1998) Oligomerization of endogenous and synthetic amyloid beta-protein at nanomolar levels in cell culture and stabilization of monomer by Congo red. *Biochemistry* 37(11): 3602-3611.
 26. Uversky VN (2019) Supramolecular Fuzziness of Intracellular Liquid Droplets: Liquid-Liquid Phase Transitions, Membrane-Less Organelles, and Intrinsic

- Disorder. *Molecules* 24(18): 3265.
27. Lv Z, Hashemi M, Banerjee S, Zagorski K, Rochet JC, et al. (2019) Assembly of alpha-synuclein aggregates on phospholipid bilayers. *Biochim Biophys Acta Proteins Proteom* 1867: 802-812.
 28. Banerjee S, Hashemi M, Zagorski K, Lyubchenko YL (2020) Interaction of Abeta42 with Membranes Triggers the Self-Assembly into Oligomers. *Int J Mol Sci* 21(3): 1129.
 29. Lv Z, Banerjee S, Zagorski K, Lyubchenko YL (2018) Supported Lipid Bilayers for Atomic Force Microscopy Studies. *Methods Mol Biol* 1814: 129-143.
 30. Banerjee S, Lyubchenko YL (2019) Interaction of Amyloidogenic Proteins with Membranes and Molecular Mechanism for the Development of Alzheimer's disease. *Alzheimers Res Ther* 2(1): 000106.
 31. Korczyn AD (2008) The amyloid cascade hypothesis. *Alzheimer's & Dementia* 4(3): 176-178.
 32. Wang J, Gu BJ, Masters CL, Wang YJ (2017) A systemic view of Alzheimer disease - insights from amyloid-beta metabolism beyond the brain. *Nat Rev Neurol* 13(10): 612-623.
 33. Rasmussen J, Mahler J, Beschorner N, Kaeser SA, Häsler LM, et al. (2017) Amyloid polymorphisms constitute distinct clouds of conformational variants in different etiological subtypes of Alzheimer's disease. *Proceedings of the National Academy of Sciences* 114(49): 13018-13023.
 34. Kumar S, Henning-Knechtel A, Chehade I, Magzoub M, Hamilton AD (2017) Foldamer-Mediated Structural Rearrangement Attenuates A β Oligomerization and Cytotoxicity. *Journal of the American Chemical Society* 139(47): 17098-17108.
 35. Eskici G, Axelsen PH (2017) Amyloid Beta Peptide Folding in Reverse Micelles. *Journal of the American Chemical Society* 139(28): 9566-9575.
 36. Bode DC, Baker MD, Viles JH (2017) Ion Channel Formation by Amyloid-beta42 Oligomers but Not Amyloid-beta40 in Cellular Membranes. *J Biol Chem* 292(4): 1404-1413.
 37. Aleksis R, Oleskovs F, Jaudzems K, Pahnke J, Biverstål H (2017) Structural studies of amyloid- β peptides: Unlocking the mechanism of aggregation and the associated toxicity. *Biochimie* 140: 176-192.
 38. Zhao Y, Wu X, Li X, Jiang LL, Gui X, et al. (2018) TREM2 Is a Receptor for beta-Amyloid that Mediates Microglial Function. *Neuron* 97(5): 1023-1031.
 39. Cenini G, Rub C, Bruderek M, Voos W (2016) Amyloid beta-peptides interfere with mitochondrial preprotein import competence by a coaggregation process. *Mol Biol Cell* 27(21): 3257-3272.
 40. Sorrentino V, Romani M, Mouchiroud L, Beck JS, Zhang H, et al. (2017) Enhancing mitochondrial proteostasis reduces amyloid-beta proteotoxicity. *Nature* 552: 187-193.
 41. Ruiz-Arias A, Paredes JM, Di Biase C, Cuerva JM, Giron MD, et al. (2020) Seeding and Growth of beta-Amyloid Aggregates upon Interaction with Neuronal Cell Membranes. *Int J Mol Sci* 21(14): 5035.
 42. Matsubara T, Yasumori H, Ito K, Shimoaka T, Hasegawa T, et al. (2018) Amyloid-beta fibrils assembled on ganglioside-enriched membranes contain both parallel beta-sheets and turns. *J Biol Chem* 293(36): 14146-14154.
 43. Ewald M, Henry S, Lambert E, Feuillie C, Bobo C, et al. (2019) High speed atomic force microscopy to investigate the interactions between toxic Abeta1-42 peptides and model membranes in real time: impact of the membrane composition. *Nanoscale* 11(15): 7229-7238.
 44. Hammond K, Ryadnov MG, Hoogenboom BW (2020) Atomic force microscopy to elucidate how peptides disrupt membranes. *Biochim Biophys Acta Biomembr* 1863(1): 183447.
 45. Banerjee S, Hashemi M, Zagorski K, Lyubchenko YL (2020) Cholesterol in membranes facilitates aggregation of amyloid β protein at physiologically low concentrations. *bioRxiv*.
 46. Bucciantini M, Rigacci S, Stefani M (2014) Amyloid Aggregation: Role of Biological Membranes and the Aggregate-Membrane System. *J Phys Chem Lett* 5(3): 517-527.
 47. Roher AE, Kokjohn TA, Clarke SG, Sierks MR, Maarouf CL, et al. (2017) APP/A β structural diversity and Alzheimer's disease pathogenesis. *Neurochemistry International* 110: 1-13.
 48. Wingo TS, Cutler DJ, Wingo AP, Le NA, Rabinovici GD, et

- al. (2019) Association of Early-Onset Alzheimer Disease With Elevated Low-Density Lipoprotein Cholesterol Levels and Rare Genetic Coding Variants of APOB. *JAMA Neurol* 76(7): 809-817.
49. Xue-Shan Z, Juan P, Qi W, Zhong R, Li Hong P, et al. (2016) Imbalanced cholesterol metabolism in Alzheimer's disease. *Clin Chim Acta* 456: 107-114.
50. Umeda T, Tomiyama T, Kitajima E, Idomoto T, Nomura S, et al. (2012) Hypercholesterolemia accelerates intraneuronal accumulation of Abeta oligomers resulting in memory impairment in Alzheimer's disease model mice. *Life Sci* 91(23-24): 1169-1176.
51. Loef M, von Stillfried N, Walach H (2012) Zinc diet and Alzheimer's disease: a systematic review. *Nutr Neurosci* 15(5): 2-12.
52. Singh B, Parsaik AK, Mielke MM, Erwin PJ, Knopman DS, et al. (2014) Association of mediterranean diet with mild cognitive impairment and Alzheimer's disease: a systematic review and meta-analysis. *J Alzheimers Dis* 39: 271-282.
53. Yusuf M, Weyandt LL, Piryatinsky I (2017) Alzheimer's disease and diet: a systematic review. *Int J Neurosci* 127(2): 161-175.
54. Hill E, Goodwill AM, Gorelik A, Szoek C (2019) Diet and biomarkers of Alzheimer's disease: a systematic review and meta-analysis. *Neurobiol Aging* 76: 45-52.

