



Comparative Evaluation of Proximate, Mineral, Vitamins, Phytochemical and Antioxidant Properties of Pulp and Seeds of Doum Palm (*Hyphaene Thebaica*) in India

John AO*

Department of Animal Nutrition and Biochemistry, Sumitra Research Institute, India

*Corresponding author: Alagbe Olujimi John, Department of Animal Nutrition and Biochemistry, Sumitra Research Institute, Gujarat, India, Email: dralagbe@outlook.com

Research Article

Volume 8 Issue 1

Received Date: March 01, 2024

Published Date: April 05, 2024

DOI: 10.23880/beba-16000229

Abstract

Medicinal plants are natural gift that contains phytochemicals that are ecofriendly, effective and non-toxic. These chemicals can also prevent antimicrobial resistance. However, the concentration of phytochemicals and nutritional properties underlines certain variation due to geographical location, age of plant, storage condition, extraction methods, amongst others. Therefore, this study was carried out to compare the proximate, mineral, vitamins, phytochemical and antioxidant properties of pulp (DPML) and seeds of Doum palm (DPSM) (*Hyphaene Thebaica*) in India. Proximate evaluation of DPML revealed that it had higher ($p < 0.05$) levels of moisture (8.77%), crude fibre (13.06%), ether extracts (1.21%), ash (8.21%), carbohydrate (69.33%) and lower protein (6.21%) relative to DPSM which contained moisture, crude protein, crude fibre, ether extract, ash and carbohydrates at 6.93 %, 8.07 %, 11.69 %, 0.09 %, 6.44 % and 54.08 % respectively. DPML had higher ($p < 0.05$) concentration of calcium (387.21 mg/100g), phosphorus (2956.1mg/100g), potassium (175.1mg/100g), manganese (28.03mg/100g), magnesium (186.1mg/100g), zinc (10.33mg/100g), copper (8.61mg/100g), sodium (228.1mg/100g) and iron (9.11 mg/100g) compared to DPSM which contained calcium, phosphorus, potassium, manganese, magnesium, zinc, copper, sodium and iron at 206.8 mg/100g, 1600.2 mg/100g, 113.5 mg/100g, 15.14 mg/100g, 102.4 mg/100g, 10.05 mg/100g, 5.78 mg/100g, 190.2 mg/100g and 6.08 mg/100g respectively. Doum palm pulp meal contained greater concentrations ($p < 0.05$) of vitamin A (9.38iu/100g), vitamin B1 (26.01 μ g/100g), vitamin B1 (11.73 μ g/100g), vitamin B6 (57.10 μ g/100g), vitamin B12 (15.61 μ g/100g), vitamin C (122.6 μ g/100g) and vitamin E (31.62 μ g/100g) relative to 5.06iu/100g, 21.48 μ g/100g, 9.45 μ g/100g, 42.08 μ g/100g, 10.20 μ g/100g, 95.18 μ g/100g and 23.49 μ g/100g reported for vitamin A, vitamin B1, vitamin B6, vitamin B12, vitamin C and vitamin E respectively. Doum palm pulp meal produced higher concentrations of phenols (82.10mg/g), flavonoids (39.15mg/g) and saponins (2.16mg/g) and lower concentrations of tannins (11.72mg/g) and alkaloids (6.13mg/g) relative to doum palm seed meal ($p < 0.05$) which contained phenols (65.72mg/g), flavonoids (30.86 mg/g), tannins (18.83mg/g), alkaloids (9.17mg/g) and saponins (1.91mg/g). DPPH was greater ($p < 0.05$) in doum palm pulp meal with 31.82% compared to doum palm seed meal (26.07%). It was concluded that DPML and DPSM is loaded with nutrients and other medicinal properties making them useful in livestock production.

Keywords: Phytochemicals; Medicinal Plants; Nutrients; Vitamins; Minerals; Antioxidant

Abbreviations: DPSM: Doum Palm Seed Meal; SAS: Statistical Analysis System; DPML: Doum Palm Pulp Meal.

Introduction

Plants are an essential source of nutrition and medicinal substances, and they have always been prized natural riches [1]. They are capable of producing a large range of different compounds, which can be further classified into primary and secondary metabolites. All plants have primary metabolites like sugar and lipids, but only a limited number of plants produce secondary metabolites, which are created by those plants and have particular uses [2]. Secondary metabolites are highly intriguing and potent natural chemicals that are acknowledged for their medical usefulness [3].

A very broad range of physiologically active substances, including alkaloids, glucosinolates, cyanogenic glucosides, flavonoids, tannins, coumarins, lignans, terpenoids, saponins, organic acids, and many more, are part of the defensive chemistry of plants [4-7]. These substances function as signaling substances to draw in pollinators or predators in addition to acting as deterrents against herbivory and frequently against microbial infection [3]. Biologically active chemicals are typically found in complex combinations found in medicinal plants. They have an impact on several targets. Plants used in phytotherapy often have low toxicity levels [8]. Treating chronic diseases may benefit from the use of certain of the active secondary metabolites. The doum palm is one of the world's potentially beneficial plants [9].

Doum palm (*Hyphaene thebaica*) is a dichotomous, tall, multi-stemmed desert plant belonging to the family Arecaceae and order Arecales. It can grow up to 15 m in height and is endemic to Africa and some parts of Asia especially west India [10,11]. The leaves contain strong fibres and are used to make mats, bind parcels and writing paper. The trunk of the palm is used for construction, as well as for manufacture of various domestic utensils [8,12]. The oblong, yellow orange apple sized fruit has a red outer skin, a thick, spongy and rather sweet, fibrous fruit pulp that tastes like gingerbread and a large kernel [13]. The covering of the fruit is edible and can either be pounded to form a powder or cut off in slices; the powder is often dried then added to food as a flavoring agent [14,15].

According to Datti, et al. [16], doum palm fruit pulp contains minerals: calcium, phosphorus, manganese, potassium, magnesium, sodium, zinc, iron, copper and cobalt needed for enzymatic reactions in the body of animals. It is also rich in protein at 2.86 %, ash (6.20%), ether extract (0.92%), crude fibre (13%) and carbohydrate at 68.5%. Phytochemical analysis of doum palm fruit also reveals

the presence of tannins, flavonoids, phenols, alkaloids and saponins which are capable of performing multiple biological activities: antimicrobial, antioxidant, immune-stimulatory, hepato-protective, antifungal, antiviral, amongst others [17,18]. Doum palm pulp extracts also have the capacity to inhibit the activities of some pathogenic organisms such as: *Shigella flexneri*, *Salmonella typhi*, *Proteus mirabilis*, *Pseudomonas aeruginosa*, *Klebsiella pneumonia*, *Enterococcus faecalis*, *Listeria monocytogenes* and *Bacillus cereus* due to the presence of phytochemicals [19,20].

Previous studies have shown that chemical composition of Doum palm can be influenced by age of plants, geographical location, method of processing, climate, storage conditions, amongst others [21]. This research is timely, because it gives a clue on the nutritional properties of *Hyphaene thebaica* found in Gujarat as well as to examine whether they can be utilized as a potential feed additive for livestock.

Materials and Methods

Site of the Experiment

The investigation was carried out at the Department of Animal Nutrition and Biochemistry, Sumitra Research laboratory, Gujarat, India in the month of January, 2024. The Research Institute is located between 23°13'N and 72°41'E. All laboratory procedures were strictly adhered to according to guidelines laid down by Association of Analytical Chemists. Laboratory kits were adjusted following the manufacturer's instruction.

Collection of Fresh Doum Palm and Processing

Fresh, dried mature fruits of doum palm were harvested within the premises of Sumitra Research Institute, Gujarat India from different trees and taken to the department of Biological Sciences of the same institute where samples were identified by a certified plant taxonomist and assigned a voucher specimen number AA/SM/24. The collected fruits were crushed using a mechanical splitter to separate doum pulp from the kernel. Each of the component (pulp and seed) were shade dried for five days before it was pulverized into powder and stored separately in a labeled air tight polythene bag before extraction. Doum palm pulp meal was abbreviated as DPML while doum palm seed meal (DPSM).

200 g of the sample were soaked separately into 1litre of water and agitated in a blender for 5 minutes before it was transferred into a container, stirred intermittently and kept for 48 hours before it was filtered through a filter. The extracts (filtrate) were used for the determination of antioxidant properties and vitamin composition in the samples.

Reagents for Phytochemical Evaluation

Sodium hydroxide, sodium bicarbonate, folin-ciocalteu's reagent, aluminum chloride, sodium nitrate, sulphuric acid, bromocresol solution, ferric ammonium sulphate, amyl alcohol and ammonium thiocyanate solution.

Laboratory Equipment Used for the Experiment

Test tubes, beakers, conical flask, water bath, digital thermometer and conical flask.

Proximate Analysis of DPML and DPSM

Proximate composition of samples was carried out using Perkin Elmer near infra-red kit (Model: FT 9700, China). 200 grams of each sample was passed through the sample collecting vat connected to a monitor (for display of results; scanning grating transmittance). For proper efficiency the equipment was set to a wavelength of 570 to 1100 nm to display results (moisture, crude protein, crude fibre, ether extract, ash and carbohydrates) at an analysis time of 25 seconds.

Mineral Evaluation of DPML and DPSM

100 g of each sample was used for mineral analysis using Atomic Absorption Spectrometer (Model: SP-AA 4500, China) equipped with integrated computer controlled longitudinal heated graphite furnace. For efficiency, the machine is set at a programmable temperature up to 3000°C in 1°C increment, wavelength repeatability and accuracy of ± 0.1 nm and ± 0.3 nm respectively. Slits are automatically selected at 0.1; 0.2; 0.4; 0.7; 1.4 and 2.0 nm, wavelength range (185–900 nm), injection volumes from 1 to 50 μ L, heating rate of 2000°C/s under software control (SPWinAA Software Package) to display results.

Vitamin Analysis of DPML and DPSM

Quantifying fat and water soluble vitamins was carried out using Agilent LC/MS kit equipped with jet steam electrospray ionization coupled with diode array detector. For precision, all the manufacturers instruction is strictly adhered to. For the MS chamber: drying gas temperature and sheath gas flow is adjusted at 250°C and 350°C, sheath gas flow (12 L/min), capillary (4,000 V) and nozzle voltage (1000 V) while LC chamber is configured at an injection volume of 5 μ L, auto sampler temperature (15°C) and mobile phase: H₂O with 0.1% FA, 4.5 mM ammonium formate and 0.5 mM ammonium fluoride as well as MeOH with 0.1% FA, 4.5 mM ammonium formate and 0.5 mM ammonium fluoride respectively in each column.

Gas Chromatograph Analyzer for DPML and DPSM

Different optical densities for phytochemical samples was carried out with YL6500 GC gas chromatograph made up of 3 components viz: inlet, column oven and detector (data acquisition chamber). Inlet chamber is adjusted to a maximum temperature of 450°C, total flow setting range for samples: 0.01 to 100 mL/minute, pressure range (0.001 to 100 psi) and flow stability ($< \pm 0.05$ mL/min) while the column oven is maintained at a heating rate 120°C/min, cooling down option (80°C to 450°C with LN₂ cryogenic cooling), maximum run time of 9,999 min while the data acquisition unit is coupled with flame ionization detector, thermal conductivity detector, electron capture detector, nitrogen phosphorus detector, flame photometric detector and pulsed discharge detector at temperatures of 450°C, 400°C, 400°C, 400°C, 300°C and 400°C respectively.

Quantification of Total Tannins in DPML and DPSM

5g of DPML and DPSM was added to 1.5mL of ammonium sulphate in a conical flask, the mixture was covered for 5 minutes and kept under room temperature followed by the addition of 1.0 mL sodium bicarbonate, 10 mL distilled water and stirred before it was introduced to YL6500 GC gas analyzer and adjusted to an optical density of 560 nm.

Estimation of Total Phenolic Compounds in DPML and DPSM

5g of DPML and DPSM was added to 1.5 mL of Folin-Ciocalteu reagent in a beaker, the mixture was covered for 10 minutes and kept under room temperature followed by the addition of 1.0 mL sodium carbonate, 10 mL distilled water and stirred before it was introduced to YL6500 GC gas analyzer and adjusted to an optical density of 700nm.

Estimation of Flavonoids in DPML and DPSM

5 g of DPML and DPSM was added was added to 0.8 mL nitric oxide mixed together in a test tube, the mixture was incubated for 10 minutes followed by the addition of 0.5 mL of sodium hydroxide. It was introduced to YL6500 GC gas analyzer and adjusted to an optical density of 850 nm according to standard laboratory procedures outlined by Mahmoudi, et al.

Determination of Alkaloid Concentration

5g of DPML and DPSM was diluted with 2 mL of phosphate buffer and 1.5 mL of bromocresol green solution were mixed

together in a test tube covered and allowed to stay for 30 minutes before it was injected into spectrophotometer and set at an optical density of 510 nm. Other procedures were carried out according to the methods outline by Njoku and Chidi.

Determination of Saponins

5 g of DPML and DPSM was diluted with 1.0 mL of distilled water followed by the addition of 0.5 mL ferric ammonium sulphate. The mixture was thoroughly mixed in a test tube, covered and cooled at room temperature for 10 minutes. YL6500 GC gas analyzer and adjusted to an optical density of 650 nm according to standard laboratory procedures outlined by Bayero, et al.

Antioxidant Properties of DPML and DPSM

Free radical scavenging activity of the extract was measured by 1,1-diphenyl-2-picryl hydrazyl (DPPH). Here, 2 mL of DPML and DPSM extract was added to 2 mL of 0.4 nM methanolic solution of DPPH. The mixture was then shaken vigorously and allowed to stand at room temperature for 30 min. The absorbance was then measured at 520 nm using YL6500 GC gas analyzer.

Statistical Analysis

Data collected were subjected to analysis adopting Student's T-test and Statistical Analysis System (SAS, version 9.4) PROC ANOVA GLM Analysis of Variance.

Results and Discussion

As presented in Table 1, proximate composition of doum palm pulp meal (DPML) and doum palm seed meal (DPSM). DPML had higher ($p < 0.05$) levels of moisture (8.77%), crude fibre (13.06%), ether extracts (1.21%), ash (8.21%), carbohydrate (69.33%) and lower protein (6.21%) relative to DPSM which contained moisture, crude protein, crude fibre, ether extract, ash and carbohydrates at 6.93%, 8.07%, 11.69%, 0.09%, 6.44% and 54.08% respectively. Moisture content in a sample is used to determine its level of deterioration, thus a lower moisture level suggests a longer shelf life of a sample [22,23]. The moisture content reported in this experiment were higher lower than those reported for *Kigelia africana* pulp (9.70%). Though the protein content of doum palm seed meal was higher compared to doum palm pulp meal, both result indicates that they are low in protein and cannot be used as protein supplement in the diets of monogastric animals. Results obtained for protein in this experiment, disagrees with the findings of Waleed, et al. and Datti, et al. [24] who recorded 2.86 % and 2.32 % for doum palm seed meal and doum pulp meal respectively.

These discrepancies can be attributed to variation in climate, geographical location and age of plants [2]. Ash content in a sample is used to determine the concentration of minerals in a sample [23]. Ash content in DPML and DPSM was higher than those reported for *Phoenix dactylifera* seeds (1.30%) by Eimad, et al. [25]. Crude fibre in diet of animals helps to improve digestion and reduce serum cholesterol level, thus preventing the incidence of coronary diseases [26,27]. The result in this experiment suggests that DPML and DPSM are good sources of dietary fibre. The values recorded for crude fibre and ether extract is in consonance with the report of Aboshora, et al. [28] when fresh, epicarp and pitted sample of doum fruit was examined. Result on carbohydrate composition of DPML revealed that it is a good source of energy compared to DPSM. Carbohydrate are needed for the maintenance of basal metabolism of various organs and tissues of animals and also needed for proper body motion [21]. Earlier studies have reported 62.70%, 75.20% and 43.0% carbohydrate composition for seeds and meal of cashew nut as well as those of tiger nut residues [29,30].

Variables	*DPML	**DPSM	p value
Moisture	8.77 ± 0.17b	6.93 ± 0.15a	< 0.01
Crude protein	6.21 ± 0.04b	8.07 ± 0.06a	< 0.01
Crude fibre	13.06 ± 0.03a	11.69 ± 0.02b	0.02
Ether extracts	1.21 ± 0.04a	0.09 ± 0.06b	< 0.01
Ash	8.21 ± 0.02a	6.44 ± 0.01b	0.01
Carbohydrate	69.33 ± 0.27a	54.08 ± 0.12b	0.03

*Doum palm pulp meal: DPML; **Doum palm seed meal: DPSM

Table 1: Proximate composition of Doum palm pulp meal (DPML) and doum palm seed meal (DPSM).

As presented in Table 2, the mineral composition of doum palm pulp meal (DPML) and doum palm seed meal (DPSM). DPML had higher ($p < 0.05$) concentration of calcium (387.21mg/100g), phosphorus (2956.1mg/100g), potassium (175.1mg/100g), manganese (28.03mg/100g), magnesium (186.1mg/100g), zinc (10.33mg/100g), copper (8.61mg/100g), sodium (228.1mg/100g) and iron (9.11mg/100g) compared to DPSM which contained calcium, phosphorus, potassium, manganese, magnesium, zinc, copper, sodium and iron at 206.8mg/100g, 1600.2mg/100g, 113.5mg/100g, 15.14 mg/100g, 102.4mg/100g, 10.05mg/100g, 5.78mg/100g, 190.2mg/100g and 6.08mg/100g respectively. This result suggests that DPML is loaded with various minerals needed for the activation of enzymes and movement of nutrients and waste around the body cells of animals [1]. For instance, calcium is needed for heathy bones, effective blood pressure regulation and muscle contraction [31]. Phosphorus aids in the synthesis

of deoxyribonucleic acid and maintains acid base balance [32]. Magnesium are needed for muscle contraction and acts as co-factors for metabolic enzymes [33]. Zinc boost the body immunity and are needed for making protein and genetic material [34]. Copper is needed for iron metabolism and creation of haemoglobin regulating neurotransmitters [34]. Sodium enhances proper fluid and pH balance, nerve

transmission and muscle contraction [35]. Potassium regulates heart beats and effective nerve functioning [36]. Iron is a part of haemoglobin found in red blood cell that carries oxygen in the body and also needed for energy metabolism. The result obtained is in agreement with the findings of Bonde, et al. [37] when nutritional composition of doum palm fruits in west coast of India was examined.

Variables	*DPML (mg/100g)	**DPSM (mg/100g)	p value
Calcium	387.21 ± 0.12a	206.8 ± 0.09b	0.012
Phosphorus	2956.1 ± 0.02a	1600.2 ± 0.01b	0.001
Potassium	175.1 ± 0.06a	113.5 ± 0.10b	0.003
Manganese	28.03 ± 0.02a	15.14 ± 0.08b	0.01
Magnesium	186.1 ± 0.10a	102.4 ± 0.12b	0.001
Zinc	10.33 ± 0.03a	10.05 ± 0.01b	0.002
Copper	8.61 ± 0.01a	5.78 ± 0.01b	0.001
Sodium	228.1 ± 0.17a	190.2 ± 0.14b	0.001
Iron	9.11 ± 0.03a	6.08 ± 0.02b	0.001

*Doum palm pulp meal: DPML; **Doum palm seed meal: DPSM

Table 2: Mineral composition of doum palm pulp meal (DPML) and doum palm seed meal (DPSM).

As presented in Table 3, vitamin composition of doum palm pulp meal (DPML) and doum palm seed meal (DPSM). Doum palm pulp meal contained greater concentrations ($p < 0.05$) of vitamin A (9.38 iu/100g), vitamin B₁ (26.01µg/100g), vitamin B₁ (11.73µg/100g), vitamin B₆ (57.10µg/100g), vitamin B₁₂ (15.61µg/100g), vitamin C (122.6µg/100g) and vitamin E (31.62µg/100g) relative to 5.06iu/100g, 21.48µg/100g, 9.45µg/100g, 42.08µg/100g, 10.20µg/100g, 95.18µg/100g and 23.49µg/100g reported for vitamin A, vitamin B₁, vitamin B₆, vitamin B₁₂, vitamin C and vitamin E respectively. Vitamins are chemical compounds that are needed in small amounts for biological functions and growth maintenance [37]. Insufficient inclusion of vitamins in the diets of animals could lead to deficiency syndrome [38]. For instance, vitamin B₁ is needed in the production of cholesterol and different kinds of hormones, synthesis of fats

and carbohydrates as well as replication of deoxyribonucleic acid [39]. Vitamin B₆ helps in the creation of red blood cells and maintaining the immune system of animals [40]. Vitamin B₁₂ is required for the maintenance of the nervous system as well as synthesis of glucose [41]. Lack of vitamin B complexes in the diet of animals may result in muscle body weakness [42]. Vitamin C is an antioxidant that protects cells from free radicals and unstable molecules that can damage body cells [43]. Vitamin D is required to maintain normal levels of calcium and phosphorus in the blood [32]. Antioxidant that protects other nutrients like vitamin A and certain lipids from being damaged is the sole responsibility of vitamin E [44]. Insufficient quantities of vitamin C, D and E in the diets of animals could result in scurvy, rickets and neuropathy/anemia respectively [41].

Variables	Units	*DPML	**DPSM	p value
Vitamin A	iu/100g	9.38 ± 0.03a	5.06 ± 0.02b	< 0.01
Vitamin B1	µg/100g	26.01 ± 0.19a	21.48 ± 0.16b	0.02
Vitamin B2	µg/100g	11.73 ± 0.09a	9.45 ± 0.06b	0.01
Vitamin B6	µg/100g	57.10 ± 0.12a	42.08 ± 0.10b	0.01
Vitamin B12	µg/100g	15.61 ± 0.17a	10.20 ± 0.14b	< 0.01
Vitamin C	µg/100g	122.6 ± 0.44a	95.18 ± 0.21b	0.02
Vitamin E	µg/100g	31.62 ± 0.11a	23.49 ± 0.08b	0.01

*Doum palm pulp meal: DPML; **Doum palm seed meal: DPSM

Table 3: Vitamin composition of doum palm pulp meal (DPML) and doum palm seed meal (DPSM).

As presented in Table 4, the phytochemical analysis of doum palm pulp meal (DPML) and doum palm seed meal (DPSM). Doum palm pulp meal produced higher concentrations of phenols (82.10mg/g), flavonoids (39.15mg/g) and saponins (2.16mg/g) and lower concentrations of tannins (11.72mg/g) and alkaloids (6.13mg/g) relative to doum palm seed meal ($p < 0.05$) which contained phenols (65.72mg/g), flavonoids (30.86mg/g), tannins (18.83mg/g), alkaloids (9.17mg/g) and saponins (1.91mg/g). The results suggest that both samples contain chemical compounds with pharmacological properties viz: antioxidant, hypoglycemic, anti-bacterial, antifungal, hepatoprotective, anti-inflammatory, neuroprotective, antiseptic, antiviral, anti-androgenic, anti-proliferative and antipyretic properties [45,46]. However, results obtained is in agreement with the findings of Ghada, et al. [15]; Mohammed, et al. [47]. Flavonoids are known to have antimicrobial, anti-inflammatory, anti-allergic, anti-hyperglycemic and antioxidant properties. Phenols possess antioxidant properties and are capable of preventing the body against oxidative stress, thus preventing infections [47,48]. Recent findings demonstrate that tannins possess gastro-protective, anti-inflammatory and antimicrobial activities [49]. Alkaloids exhibit anti-cancer, anti-inflammatory, anticonvulsant, anti-dysentery and analgesics activities [50]. Pharmacologically, saponins have been shown to exhibit hormonal and anti-fungal, antioxidants and hypolipidemic and anti-androgenic activities [51].

Variables	*DPML (mg/g)	**DPSM (mg/g)	p value
Phenols	82.10 ± 0.08a	65.72 ± 0.03b	0.003
Flavonoids	39.15 ± 0.17a	30.86 ± 0.12b	0.001
Tannins	11.72 ± 0.25b	18.83 ± 0.21a	0.002
Alkaloids	6.33 ± 0.00b	9.17 ± 0.00a	0.001
Saponins	2.16 ± 0.01a	1.91 ± 0.02b	0.001

*Doum palm pulp meal: DPML; **Doum palm seed meal: DPSM

Table 4: Phytochemical analysis of doum palm pulp meal (DPML) and doum palm seed meal (DPSM).

As presented in Table 5, antioxidant properties of doum palm pulp meal (DPML) and doum palm seed meal (DPSM) to scavenge 1,1-diphenyl-2-picryl hydrazyl. DPPH was greater ($p < 0.05$) in doum palm pulp meal with 31.82 % compared to doum palm seed meal (26.07%). The results indicated that both samples possess antioxidant properties. However, DPML has more potential to scavenge reactive oxygen species and free radicals thus preventing cell damage and infections in the body [52]. Free radicals' steals electron from a healthy cell causing breakdown in activities [21]. Antioxidant defenses suppress free radicals that are toxic to the cells [1]. The high concentration in vitamin A, C and E recorded

in DPML can reduce and detoxify oxygen intermediates in cells of animals. The antioxidant activity of DPML and DPSM is lower than values recorded for Phoenix dactylifera fruit (35.64 to 67.56%), Soursop pulps (61.22%), soursop peels and seeds (51.10%, 48.09%) recorded by Safia, et al. [53]; Hakime, et al. [54]. The variation in these results can be attributed to differences chemical compounds, extraction and storage techniques, species amongst other [55-57].

Variables	*DPML (mg/g)	**DPSM (mg/g)	p value
1,1-diphenyl-2-picryl hydrazyl (DPPH)***	31.82 ± 0.05a	26.07 ± 0.03b	0.002

*Doum palm pulp meal: DPML; **Doum palm seed meal: DPSM; ***DPPH: 1,1-diphenyl-2-picryl hydrazyl

Table 5: Antioxidant properties of doum palm pulp meal (DPML) and doum palm seed meal (DPSM).

Conclusion

It was concluded that doum palm pulp meal (DPML) and doum palm seed meal (DPSM) have nutritional and medicinal properties due to the presence of phytochemicals which promotes multiple biological activities such as: antimicrobial, antifungal, hepato-protective, immune-stimulatory, antiviral, antioxidant properties amongst others. DPML also contains higher antioxidant properties compared to doum palm seed meal making it possible to scavenge free radicals, thus preventing infection. Therefore, both DPML and DPSM can be utilized by livestock.

References

1. Singh S, Alagbe OJ, Xing L, Ram S, Amita K, et al. (2022) Comparative analysis of ethanolic *Juniperus thurifera* leaf, stem bark and root extract using gas chromatography and mass spectroemetry. International Journal of Agriculture and Animal Production 2(6): 18-27.
2. Alagbe JO, Anuore DN (2023) Effect of Doum palm mesocarp meal (*Hyphaene thebaica*) as partial replacement for maize on growth performance and hematological indices of weaned pigs. Journal of Biotechnology and Bioinformatics Research 5(3): 1-6.
3. Shittu MD, Alagbe JO, Adejumo DO, Ademola SG, Abiola AO, et al. (2021) Productive Performance, Caeca Microbial Population and Immune-Modulatory Activity of Broiler Chicks Fed Different Levels Sida Acuta Leaf Extract in Replacement of Antibiotics. Bioinformatics and Proteomics Open Access Journal 5(1): 000143.
4. Alagbe JO, Muhammad RS, Shittu MD, Christiana OO (2022) Effect of *Trichilia monadelpha* stem bark extract

- on the fatty acid composition of rabbit's thigh meat. *Journal of Environmental Issues and Climate Change* 1(1): 63-71.
5. Alagbe JO (2022) Gas chromatography and mass spectroscopy of *Juniperus phoenice* stem bark extract and its influence on the haemato-biochemical values of growing rabbits. *British Scientific Periodical* 2(5): 18-33.
 6. Alagbe JO, Habiba Z, Adedeji OM, Bamigboye S, Agbonika D, et al. (2022) Influence of *Juniperus thurifera* root extract on the nutrient digestibility and caecal microbial count of growing rabbits. *Web of Synergy International Interdisciplinary Research Journal* 1(1): 5-17.
 7. Faten MA (2009) Antioxidant and Anticancer Activities of Doum Fruit Extract (*Hyphaene thebaica*). *African Journal of Pure and Applied Chemistry* 3(10): 197-201.
 8. Alagbe JO, Shittu MD, Ramalan SN, Tanimomo KB, Adekunle DA, et al. (2022) Growth performance, semen quality characteristics and hormonal profile of male rabbit bucks fed *Rubia cordifolia* root extracts. *International Journal of Biological Engineering and Agriculture* 1(1): 1-13.
 9. Fletcher R (1997) Listing of useful plants of the world. *Australian New crops*.
 10. Orwa C, Mutua A, Kindt R, Jamnadass R, Simons A, et al. (2009) Agroforestry database: a tree reference and selection guide version 4.0.
 11. Moussa H, Hank A, Margolis HA, Dube P, Odongo J, et al. (1998) Factors Affecting the Germination of Doum Palm (*Hyphaene thebaica* Mart.) Seeds from the Semi-Arid Zone of Niger, West Africa. *For Ecol Management* 104(1-3): 27-41.
 12. Wendakoon C, Calderon P, Gagnon D (2011) Evaluation of Selected Medicinal Plants Extracted in Different Ethanol Concentrations for Antibacterial Activity against Human Pathogens. *Journal of Medicinally Active Plants* 1(2): 60-68.
 13. Shehu BB, Gidado A, Buratai LB, Benisheikh AAG (2015) Sub-Acute Toxicity Studies of Methanol Extract of *Hyphaene thebaica* (L) Mart Fruit Pulp on Normal Wister Albino Rats. *Intl J of Res* 2(6): 460-467.
 14. Reda AA (2015) Physicochemical Properties of Doum (*Hyphaene thebaica*) Fruits and Utilization of its Flour in Formulating Some Functional Foods Alexandria. *Journal of Food Science and Technology* 12(2): 29-39.
 15. Ghada AT, Ibrahim BA, Hassan AE, Usama AM, Mohamed GS, et al. (2020) Metabolomic Profiling and Antioxidant, Anticancer and Antimicrobial Activities of *Hyphaene thebaica*. *Processes* 8(3): 1-13.
 16. Datti Y, Ibrahim SI, Abdulhadi M, Mohammad SM, Abubakar SA, et al. (2020) Mineral content, proximate composition and the antioxidant properties of the ethanol extract of *Hyphaena thebiaca* from Gazewa, Kano State, Nigeria. *Asian Journal of Applied Chemistry Research* 6(2): 33-40.
 17. Babiker HAM, Makki HMM (2013) Nutritional Value of Doum Fruits (*Hyphaene thebaica*) and Their Suitability for Production of Ready-to- Use Concentrated Drink. M.Sc. Thesis. College of Agriculture Studies Sudan University of Science and Technology.
 18. Bonde SD, Agate VV, Kulkarni DK (1990) Nutritional Composition of the Fruits of Doum Palms (*Hyphaene*) from the West Coast of India. *Principles* 34(1): 21-23.
 19. HusseinAM, SalahZA, HegazyNA (2010) Physicochemical, Sensory and Functional Properties of Wheat-Doum Fruit Flour Composite Cakes. *Polish Journal of Food and Nutrition Sciences* 60(3): 239-244.
 20. Lamiaa AG, Laith ZF (2018) Antioxidant Activity of Two Different Extracts From Doum (*Hyphaene thebaica*) Fruits. *Journal of Pharmacy and Biological Sciences* 14(4): 1-4.
 21. Alagbe JO, Shittu MD, Ushie FT (2021) GC-MS analysis of methanolic stem bark extract of *Zollingeriana indigofera*. *Asian Journal of Advances in Research* 4(1): 1265-1267.
 22. Alagbe JO, Adeoye A, Oluwatobi OA (2020) Proximate and mineral analysis of *Delonix regia* leaves and roots. *International Journal on Integrated Education* 4(1): 144-149.
 23. Alagbe JO, Shittu MD, Bamigboye SO, Oluwatobi AO (2020) Proximate and mineral composition of *Pentadiplandra brazzeana* stems bark. *Electronic Research Journal of Engineering Computer and Applied Sciences* 3(11): 91-99.
 24. Datti Y, Ibrahim SI, Abdulhadi M, Mohammad SM, Abubakar SA, et al. (2020) Mineral content, proximate composition and the antioxidant properties of the ethanol extract of *Hyphaena thebiaca* from Gazewa, Kano State, Nigeria. *Asian Journal of Applied Chemistry Research* 6(2): 33-40.
 25. Eimad TB, Chakib A, Jamal E, Mohammed B, Addi NM, et al. (2015) Phytochemical compositions and antioxidant capacity of three date (*Phoenix dactylifera*) seeds variety grown in the southern east of morocco. *Journal of the*

- Saudi Society of Agricultural Sciences 16(4): 350-357.
26. Alagbe JO (2019) Proximate, mineral and phytochemical analysis of *Piliostigma thonningii* stems bark and roots. *International Journal of Biological Physical and Chemical Studies* 1(1): 1-7.
 27. Alagbe JO (2019) Growth response and bacteria count of broiler starter given *Delonix regia* leaf extract as a natural alternative to antibiotics. *Food and Nutrition Current Research* 2(3): 197-203.
 28. Aboshora W, Lianfu Z, Dahir M, Gasmalla MA, Musa A, et al. (2014) Physicochemical, Nutritional and Functional Properties of the Epicarp, Flesh and Pitted Sample of Doum Fruit (*Hyphaene thebaica*). *Journal of Food and Nutrition Research* 2(4): 180-186.
 29. Ogunwolu SO, Henshaw FO, Mock HP, Matros A (2010) Production of protein concentrate and isolate from cashew nut. *African Journal of Food Agriculture Nutrition and Development* 10(5): 2502-2514.
 30. Samson BW, Safiya S (2013) Assessment of the nutritional and anti-nutritional components of Tiger nut meal. *International Journal of Science and research* 4(6): 342-344.
 31. Hall D, Cromwell G, Stahly T (1991) Effects of dietary calcium, phosphorus, calcium: phosphorus ratio and vitamin K on performance, bone strength and blood clotting status of pigs. *J Anim Sci* 69(2): 646-655.
 32. Hans KB, Jana T (2018) Micronutrients in the life cycle: Requirements and sufficient supply. *NFS Journal* 11: 1-11.
 33. Fairweather-Tait S, Hurrell RF (1996) Bioavailability of minerals and trace elements: Members of EC flair concerted action no. 10: Measurements of micronutrient absorption and status. *Nutr Res Rev* 9(1): 295-324.
 34. Angelova MG, Petkova-Marinova TV, Pogorielov MV, Loboda AN, Nedkova-Kolarova VN, et al. (2014) Trace element status (iron, zinc, copper, chromium, cobalt, and nickel) in iron-deficiency anaemia of children under 3 years. *Anemia* 2014: 718089.
 35. Abbaspour N, Hurrell R, Kelishadi R (2014) Review on iron and its importance for human health. *J Res Med Sci* 19(2): 164-174.
 36. Miller GD, Jarvis JK, McBean LD (2001) The importance of meeting calcium needs with foods. *J Am Coll Nutr* 20(2): 168S-185S.
 37. Booth SL, Johns T, Kuhnlein HV (1992) Natural food sources of vitamin A and provitamin A. *UNU Food Nutr Bull* 14: 6-19.
 38. Nair R, Maseeh A (2012) Vitamin D: the "sunshine" vitamin. *J Pharmacol Pharmacother* 3(2): 118-126.
 39. Lykstad J, Sharma S (2019) *Biochemistry, Water Soluble Vitamins*. StatPearls, StatPearls Publishing, Treasure Island, Florida, USA.
 40. Marcelina P, Seth S, Hanjo H (2018) Vitamin B6 and its role in cell metabolism and physiology. *Cell* 7(7): 84.
 41. He FJ, MacGregor GA (2008) Beneficial effects of potassium on human health. *Physiol Plant* 133(4): 725-735.
 42. Jeurnink TJ, De Kruif KG (1995) Calcium concentration in milk in relation to heat stability and fouling. *Neth Milk Dairy J* 49: 151-151.
 43. Hoffmann PR, Berry MJ (2008) The influence of selenium on immune responses. *Mol Nutr Food Res* 52(11): 1273-1280.
 44. Birringer M, Blumberg JB, Eggersdorfer M, Frank J, Weber P, et al. (2019) History of Vitamin E research. In: Weber P, et al. (Eds.), *Vitam E Human Health*. Nutrition and Health. Humana Press, pp: 7-18.
 45. Alagbe JO, Ushie FT (2022) Growth performance of broiler chicks fed diets containing different levels of aqueous *Citrus aurantium* stem bark extracts. *Discovery* 58(319): 735-741.
 46. Shittu MD, Alagbe JO (2020) Phyto-nutritional profiles of broom weed (*Sida acuta*) leaf extract. *International Journal of Integrated Education* 3(2): 119-124.
 47. Mohamed AA, Khalil AA, El-Beltagi HES (2010) Antioxidant and Antimicrobial Properties of Kaff Maryam (*Anastatica hierochuntica*) and Doum Palm (*Hyphaene thebaica*). *Grasas y Aceites* 61(1): 67-75.
 48. Alagbe JO, Sharma R, Abidemi OE, Shittu MD, Atanda BK, et al. (2020) Chemical evaluation of the proximate, minerals, vitamins and phytochemical analysis of *Daniellia oliveri* stem bark. *International Journal of Biological Physical and Chemical Studies* 2(1): 16-22.
 49. Wendakoon C, Calderon P, Gagnon D (2011) Evaluation of Selected Medicinal Plants Extracted in Different Ethanol Concentrations for Antibacterial Activity against Human Pathogens. *Journal of Medicinally Active Plants* 1(2): 60-68.
 50. Bidlack WR, Omaye ST, Meskin MS, Topham DW (2000)

Phytochemicals as Bioactive Agents 1st(Edn.), CRC Press, Boca Raton, Florida, USA.

51. Simões-Pires CA, Queiroz EF, Henriques AT, Hostettmann K (2005) Isolation and online identification of antioxidant compounds from three *Baccharis* species by HPLC-UV-MS/MS with post-column derivatisation. *Phytochemical Analysis* 16(5): 307-314.
52. Alagbe JO (2022) Use of medicinal plants as a panacea to poultry production and food security: A review. *Gospodarka I Innowacje* 22(2022): 1-12.
53. Safia AH, Rachida A, Abdelaziz M (2016) Antioxidant and anti-inflammatory properties of widely consumed date palm (*Phoenix dactylifera*) fruit varieties in Algerian Oases. *Journal of Food Biochemistry* 40(4): 463-471.
54. Hakime HO, Ilayda SB, Temine S (2019) Antioxidant activity of Soursop (*Annona muricata*) leaves, fruits pulps, peels and seeds. *Polish Journal of Food and Nutrition Sciences* 69(4): 359-366.
55. Singh J (2008) Maceration, Percolation and Infusion Techniques for the Extraction of Medicinal and Aromatic Plants. *Extraction Technologies for Medicinal and Aromatic Plants* 67: 32-35.
56. Kakkar P, Kumari A (2008) Screening of antioxidant potential of selected barks of Indian medicinal plants by multiple in- vitro essays. *Biomed Environ Sci* 21(1): 24-29.
57. Pihlanto A, Akkanen S, Korhonen HJ (2008) ACE-inhibitory and antioxidant properties of potato (*Solanum tuberosum*). *Food Chem* 109(1): 104-112.