



Albizia Lebbeck Stem Bark Aqueous Extract as Alternative to Antibiotic Feed Additives in Broiler Chicks Diets: Haematology, Serum Indices and Oxidative Status

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Abstract

A total of Three hundred and seventy five (375) one day old (Ross 308) broiler chicks with mixed sex were used to examine the effects of *Albizia lebbeck* stem bark (ATSM) aqueous extract as alternative to antibiotic feed additives in broiler chick's diets: haematology, serum biochemical indices and oxidative status. Birds were divided to five treatments with five replicates of fifteen (15) birds in a completely randomized design. Treatment 1 (basal diet + 0 % ATSM), treatment 2 (basal diet + 1.2 grams Oxytetracycline per litre of water), treatment 3 (basal diet + 10 ml ATSM per liter of water), treatment 4 (basal diet + 20 ml ATSM per litre of water) and treatment 5 (basal diet + 30 ml ATSM per liter of water) and the trial lasted for 56 days. Results on some haematological parameters revealed that red blood cell (RBC), pack cell volume (PCV), haemoglobin (Hb), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC), white blood cell (WBC) and its differentials were significantly ($P < 0.05$) different among the treatments. Total protein, glucose, urea, cholesterol, creatinine, aspartate aminotransaminase (AST) and alanine aminotransferase (ALT) were significantly ($P < 0.05$) affected by ATSM. Activities of superoxide dismutase (SDA), glutathione peroxidase (GPx), catalase (CAT) and malonyldialdehyde (MLA) were significantly influenced by ATSM ($P < 0.05$). It was concluded that ATSM could be administered to broiler chicks at 30 ml/litre without any negative effect on the general performance of birds.

Keywords: *Albizia Lebbeck*; Broiler Chicks; Haematology; Serum Biochemical Indices

Abbreviations: RBC: red blood cell; PCV: pack cell volume; Hb: haemoglobin; MCV: mean corpuscular volume; MCH: mean corpuscular haemoglobin; MCHC: mean corpuscular haemoglobin concentration; WBC: white blood cell; AST: aspartate aminotransaminase; ALT: alanine aminotransferase.

Introduction

Phytogenics are heterogeneous group of feed additives emanating from plants and consists of herbs, fruit, spices and other plant parts [1]. According to Veerschari et al. there are over 100, 000 species of plants used globally for medicinal purposes, many have been used in the form of therapy for

livestock among resource poor smallholder farmers to treat variety of conditions of animals [2]. Most medicinal plants have been found to be abundant in minerals, vitamins, amino acid and bioactive chemicals [phytochemicals] [3]. However, only a small percentage have been explored or studied for their pharmacological properties. Nutrients in plants have great influence on responses of animals to a disease challenge and it has a direct correlation to the immune system [4]. One of the numerous plant used for therapeutic purposes is *Albizia lebbbeck*. (Mimosaceae) is a perennial tree native to tropical and subtropical regions of Asia and Africa. The genus *Albizia* comprises of almost 150 species spread all over India, China, Nigeria, Senegal, Ghana, Togo, Congo, Benin, Angola, Uganda, Botswana among others Ukpabi and Offor, Karuppannan Karuppannan, [5]. The plant parts (stem, leaf and seeds) have been found to be loaded with minerals (calcium, phosphorus, iron, copper, zinc, selenium, molybdenum and potassium), vitamins and amino acids [6,7]. The leaf and stem has traditionally been used for the treatment of fever, tooth ache, wounds, leprosy, ulcer, cold, leprosy, sexually transmitted diseases and other respiratory infections [8,9].

Several reports on the biological activity of *Albizia lebbbeck* revealed that the plant performs antimicrobial [8], anti-inflammatory [10], antioxidant [11], analgesic [12], antihelminthic [5], hepato-protective [13], antidiabetic, immuno-modulatory [12] and antihyperlipidemic properties due to the presence of several bioactive chemicals such as alkaloids, flavonoids, saponins, phenols, tannin etc [8]. In view of these abundant potential, administration of *Albizia lebbbeck* stem bark to birds will possibly supply nutrients to meet all the body's need during a time of challenge.

Therefore, this experiment was designed to determine the effects of *Albizia lebbbeck* stem bark aqueous extract as alternative to antibiotic feed additives in broiler chick's diets: haematology, serum biochemical indices and oxidative status.

Materials and Methods

Study Area

The experiment was carried out at Division of Animal Nutrition, Sumitra Research Institute, Gujarat, India during the month of April to June, 2019.

Sources, Collection and Preparation of *Albizia Lebbbeck* Stem Bark Extract

The stem of *Albizia lebbbeck* stems were obtained from different plants in Gujarat, India and authenticated by a botanist Sharma Xing. The stem bark were cut into pieces and

thoroughly washed with distilled water, air dried under the shade to maintain the bioactive chemicals in the test material. The dried samples were pulverized into powder using pestle and mortar, thereafter 250 grams of the sample was soaked into 1000 litres of water, sample was continuously stirred and kept in the refrigerator at 4°C for 72 hours. All mixtures were filtered using Whatman filter paper and the filterates (ATSM) were collected into a clean labelled container.

Experimental Animals and Management

Three hundred and seventy five one day old (Ross 308) broiler chicks with mixed sex were used for the experiment. The birds were purchased from a commercial hatchery in India and weighed on arrival on the farm to obtain their initial body weight and thereafter weekly. A deep litter housing system was used, it was fumigated two weeks prior to the commencement of the study, and the surrounding environment was also cleaned daily to ensure proper hygiene. Birds were divided to five treatments with five replicates of fifteen (15) birds in a completely randomized design. Electric brooders were used and wood shavings serve as the litter material. Daily feed intake (g) was calculated as a difference between feed offered and left-over. Vaccines were administered according to the prevailing disease condition in the environment and all other management practices were strictly adhered to throughout the experiment which lasted for 56 days.

Ration Formulation

Three (3) basal diets were formulated at different stages of production to meet up with the requirements of birds according to NRC (1994). Broiler starter's mash (0-21 days), growers mash (22-35 days) and finishers mash (36-56 days). Treatment 1 (basal diet + 0 % ATSM), treatment 2 (basal diet + 1.2 grams Oxytetracycline per litre of water), treatment 3 (basal diet + 10 ml ATSM per liter of water), treatment 4 (basal diet + 20 ml ATSM per litre of water) and treatment 5 (basal diet + 30 ml ATSM per liter of water)

Parameters Measured

Proximate compositions of experiment diet were determined by using official method of analysis by AOAC [14]. Amino acid analysis was carried out using amino acid analyzer with ion exchange chromatographic method (Model NH-09b) India.

Haematological and Serum Biochemical Analysis

Blood samples were collected very early in the morning from the wing vein from three (3) randomly selected birds

per replicate into a 5 ml sterile syringe using 23 gauge needles and transferred into an ethylene diamine tetra acetic acid (EDTA) bottle. Haematological parameters: pack cell volume (PCV), red blood cell (RBC), haemoglobin (Hb), mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC), mean corpuscular volume (MCV), white blood cell (WBC) and its differentials were analyzed using an automated machine (Sysmex, Model KU-30 HG, India).

Serum analysis was carried out using bottles free from EDTA, blood were analyzed for total protein, albumin, globulin, glucose, cholesterol, creatinine, alanine transaminase (ALT) and aspartate transaminase (AST) were assayed using diagnostic kit manufactured by Merck India Ltd (Model PS-09R) as described by Olubukola, et al. [15].

Antioxidant Status

Activity of superoxide dismutase (SDA), glutathione

peroxidase (GPx), catalase (CAT) and malonyldialdehyde (MLA) were carried out using method outlined by Mahipal, et al. [16].

Statistical Analysis

All data were subjected to one -way analysis of variance (ANOVA) using SPSS (18.0) and significant means were separated using Duncan multiple range tests [17]. Significant was declared if $P \leq 0.05$.

Result and Discussion

The proximate composition of experimental diet (Table 1) revealed that it contains crude protein of 23.08 %, 20.11 % and 19.33 %; energy of 2936.0 kcal, 3000.8 kcal and 3100.2 kcal for starter, growers and finisher mash. The ether extract range between (4.28 – 5.03 %) and crude fibre range between (3.06 – 3.95 %). The proximate components meet the nutritional needs of birds according to NRC [18].

Materials	Starter (1-21 days)	Grower (22-35 days)	Finisher (36-56 days)
Maize	50	56	60.5
Wheat offal	8	7	8.05
Soya meal	28.55	22	21
Groundnut cake	10	11.55	6.05
Fish meal	2	2	2
Bone meal	0.35	0.4	0.4
Limestone	0.2	0.2	0.2
Lysine	0.15	0.15	0.15
Methionine	0.2	0.2	0.2
Premix	0.25	0.25	0.25
Salt	0.3	0.3	0.3
TOTAL	100	100	100
Calculated analysis			
Crude protein	23.08	20.11	19.33
Ether extract	5.03	4.87	4.28
Crude fibre	3.06	3.95	3.42
Calcium	0.98	1	1.1
Phosphorus	0.47	0.4	0.51
Lysine	1.17	1.29	1.6
Meth +Cyst	0.87	0.82	0.51
ME (Kcal/kg)	2936	3000.8	3100.2

Table 1: Chemical composition of experimental diets.

*Premix supplied per kg diet: - vit A, 13,000 I.U; vit E, 5mg; vit D3, 3000I.U, vit K, 3mg; vit B2, 5.5mg; Niacin, 25mg; vit B12, 16mg; choline chloride, 120mg; Mn, 5.2mg; Zn, 25mg; Cu, 2.6g; folic acid, 2mg; Fe, 5g; pantothenic acid, 10mg; biotin, 30.5g; antioxidant, 56mg.

The crude fibre and ether extract range also conforms to the report of Teodora, et al. [19] in feeding broilers *Hermetia illucens* meal. The calcium (0.98 – 1.10 %) and phosphorus (0.47 – 0.51 %) range in the experimental diet is in line with the reports of Aduku [20]. Proper feeding is one of the key cardinals of management in livestock production, for animals to perform at their optimum, there is need to furnish them with proper balanced diet which contains all the necessary nutrients [21].

The amino acid composition of *Albizia lebbbeck* stem bark is presented in Table 2. Results revealed the presence of threonine (1.12%), leucine (1.00 %), lysine (0.86 %), valine (0.65 %), glycine (1.00 %), phenylalanine (0.34 %), histidine (3.45 %), methionine (0.02 %), alanine (3.00 %), serine (0.78 %), proline (0.03 %), aspartic acid (2.00 %), arginine (1.65 %), tyrosine (0.10 %), isoleucine (2.00 %), aspartic acid (2.00 %) and cysteine (4.11 %). The sample contains high concentration of histidine and tyrosine has the lowest concentration. Amino acids are building blocks of protein which are necessary for gene expression and cell signal transduction regulation. Phenylalanine plays a vital role in insulin secretion and fat oxidation. Lysine ensures effective production of hormones, enzymes and energy [22]. Alanine and glutamic acid enables a healthy skeletal system and energy production for the body Marc and Wu [23]; Kimura [24]. Regulation of blood sugar is been assisted by isoleucine [25,26]. Adequate arginine ensures healthy immune system and maintains the visceral organs in the body [27,28]. Serine and cysteine play a key role as neuromodulator and

antioxidant respectively [29,30,22]. Methionine maintains the integrity of the liver, feather formation and egg size or production in birds [31,32] Table 3.

Amino acids	Composition (%)	*Reference level
Lysine	0.86	5.5
Arginine	1.65	1
Aspartic acid	2	-
Threonine	1.12	0.65
Histidine	3.45	0.3
Serine	0.78	-
Glycine	1	1.2
Alanine	3	-
Cystine	4.11	0.35
Valine	0.65	0.82
Leucine	1	1.2
Phenylalanine	0.34	0.5
Tyrosine	0.1	0.45
Isoleucine	2	0.6
Proline	0.03	0.2
Methionine	0.02	0.35

Table 2: Amino Acid Compositions of *Albizia Lebbbeck* Stem Bark.

*NRC (1994)

Parameters	T1	T2	T3	T4	T5	SEM
PCV (%)	26.50 ^c	29.31 ^b	31.02 ^b	33.56 ^a	34.00 ^a	0.37
Hb (g/dl)	9.12 ^c	10.21 ^b	11.93 ^b	12.11 ^a	12.50 ^a	0.64
RBC ×10 ⁶ /μl	1.88 ^c	2.00 ^b	2.10 ^b	2.60 ^a	2.97 ^a	0.07
MCV (fl)	111.2 ^b	119.8 ^a	120.5 ^a	123.6 ^a	130.4 ^a	8.1
MCH (pg)	34.51 ^c	50.43 ^b	56.11 ^b	57.67 ^a	59.00 ^a	2.51
MCHC (g/dl)	29.80 ^b	35.60 ^a	39.00 ^b	39.10 ^a	40.03 ^a	0.88
WBC×10 ³ /μl	20.41 ^b	20.62 ^b	22.74 ^b	22.88 ^b	30.04 ^a	0.12
Differentials (10 ³ /μl)						
Lymphocytes	10.45 ^c	14.08 ^b	15.44 ^b	18.71 ^b	20.04 ^a	1.96
Monocytes	0.07 ^c	1.11 ^b	1.20 ^b	1.26 ^b	1.72 ^a	0.01
Heterophils	4.23 ^b	5.06 ^a	5.40 ^a	5.89 ^a	6.11 ^a	0.41
Eosinophils	0.88 ^b	1.02 ^a	1.09 ^a	1.21 ^a	1.27 ^a	0.02

Table 3: Haematological parameters of broiler chicks fed different levels ATSM.

Means in the same row with different superscript are significantly different ($P<0.05$)

Haematological parameters of broiler chicks fed different levels of ATSM are presented in Table 4. PCV values

ranged between (26.50 – 34.00 %), Hb (9.12 – 12.50 g/dl), RBC 1.88 – 2.97 (10⁶/μl), MCV (111.2 – 130.4 fl), MCH (34.01

– 59.00 pg) and MCHC (29.80 – 40.03 g/dl). RBC, PCV, Hb, MCV, MCH and MCHC values were higher ($P<0.05$) in T3, T4 and T5 than for T2 and T1. WBC 20.41 – 30.04 ($10^3/\mu\text{l}$) were highest in T4 and T5 ($P<0.05$) compared to other treatments. Monocytes (0.07 – 1.72%), lymphocytes 10.45 – 20.04 ($10^3/\mu\text{l}$), heterophils 1.23 – 6.11 ($10^3/\mu\text{l}$) and eosinophils 0.88 – 1.27 ($10^3/\mu\text{l}$) were lowest ($P<0.05$) in T1 relative to other treatments. The haematological parameters measured follow similar pattern as it significantly ($P<0.05$) increased from T1 to T5. However, all values are within the physiological range for normal birds [33,34]. Islam, et al. [35]; Abdi-Hachesoo, et

al. [36] reported a RBC range (2.9 – 3.5 $10^6/\mu\text{l}$), this variation could simply be as a result of differences in age, sex, breed, environment, hormones and nutrition [37]. Haematological indices are used to in disease diagnosis as well as extent of damage to the blood [38,39]. PCV and MCH are useful indices for the diagnosis of anaemia Nse Abasi, et al. [38]; Alagbe [13]. A higher RBC level is an indication of adequate oxygen in the blood which gives room for effective nutrient transportation in the body [40,41]. WBC plays a major role in the immune system by the production of antibodies, animals with low WBC stands a high risk of infection Isaac, et al. [40].

Parameters	T1	T2	T3	T4	T5	SEM
Total protein (g/dl)	2.57 ^b	3.22 ^a	3.69 ^a	3.88 ^a	3.97 ^a	0.67
Albumin (g/dl)	1.34 ^b	1.55 ^b	1.91 ^b	2.00 ^a	2.03 ^a	0.02
Globulin (g/dl)	1.23 ^c	1.67 ^b	1.78 ^b	1.88 ^a	1.94 ^a	0.15
Creatinine (mg/dl)	0.15 ^c	0.45 ^b	0.81 ^a	0.87 ^a	0.91 ^a	0.01
Glucose (mg/dl)	196.1 ^c	204.3 ^a	234.1 ^a	241.5 ^a	250.6 ^a	4.33
Cholesterol (mg/dl)	101.4 ^a	98.6 ^b	90.4 ^a	89.4 ^a	87.5 ^a	2.87
Uric acid (mg/dl)	7.33 ^a	4.89 ^b	4.22 ^b	4.00 ^b	3.88 ^b	0.05
ALT (u/l)	74.1 ^a	70.5 ^a	61.6 ^b	58.1 ^b	50.7 ^b	1.45
AST (u/l)	300.7 ^a	288.5 ^b	230.4 ^b	218.0 ^b	200.9 ^b	9.45

Table 4: Serum analysis of broiler chicks fed different levels of ATSM. Means in the same row with different superscripts differ significantly ($P<0.05$)

The serum biochemical indices of the experimental birds are presented in Table 5. Total protein ranges (2.57 – 3.97 g/dl), globulin (1.23 – 1.97 g/dl), albumin (1.34 – 2.03g/dl), creatinine (0.51 - 0.91 mg/dl) and glucose (196.1 – 250.6 mg/dl) were lowest ($P<0.05$) for T1 compared to the other treatments while cholesterol (87.5 – 101.4 mg/dl), uric acid (50.7 – 74.1 mg/dl), ALT (50.7 – 74.1 u/l) and AST (200.9 – 300.7 u/l) was higher ($P<0.05$) for T1 and T2 than for the rest of the treatments. Total protein value in T4 and T5 were significantly higher ($P<0.05$) compared to the other treatment, this could be attributed to the presence of some relevant nutrients in ATSM [42]. Albumin content in the blood are generally influenced by protein shortages, however, the values reported fall within the range reported by Ibrahim [34]; Obajuluwa, et al. [43]; Olafadehan, et al. [44]; Livingston, et al. [45] reported a globulin and uric acid range of (1.6 – 1.9 g/dl) and (3.7-5.2 mg/dl) respectively. This result is also in agreement with the findings of Obikaonu, et al. [46] and Cholesterol, creatinine, uric acid, ALT and AST values follow similar pattern as it significantly ($P<0.05$) decreased from T1 to T5. However, all the values were within the range reported by Olafadehan, et al. [44]. Lower Creatinine and uric acid level is a sign that the kidney is not damage by feeding ATSM to the birds. According to Alagbe [2], ATSM is loaded with several minerals, vitamins and bioactive chemicals or secondary

metabolites (tannin, saponin, flavonoids, alkaloids, phenol etc.) which are within the lethal dose for broiler chicks. Urea levels is also reported to be influenced by dietary protein quality, quantity, bleeding time and are sensitive biomarkers employed in the diagnosis of renal damage [47]. ATSM can also be serves as a hypolipidemic substance because of its ability to lower blood cholesterol, thus preventing heart diseases [2]. ALT and AST are serum enzymes triggered due to the presence of a toxic substance in feed Oluwafemi, et al. [48]. The result obtain revealed that ATSM did not contain antinutrients or toxic substance which could hinder the general performance of birds, this result confirms the earlier report of Abdel, et al. [49]; Cho, et al. [50] on the effects phytogenic feed additive in broiler chicks.

The oxidative status as influenced by ATSM is presented in table. Superoxide dismutase [SDA; 35.7 – 45.3 U/mg Hb], glutathione peroxidase [GPx; 27.1 – 28.3 U/mg Hb], catalase [CAT; 40.1 – 54.2 U/mg Hb] and malonyldialdehyde [MLA; 1.85 – 3.11 U/mg Hb] values were lowest ($P<0.05$) in T1 than in other treatments. According to Alagbe, et al. [51], ATSM contains antioxidants which are capable of scavenging free radicals, thereby giving total protection to animals. The presence of phenol and flavonoids prevent oxidative damage to biomolecules, superoxide anions and lipid peroxy radicals

[52,53]. The same results were reported by Lin, et al. [54] who observed that intake of phytogetic feed additives resulted in the increase in serum antioxidant enzyme activities and a

decrease in MDA concentration. Conversely, Lan, et al. [55] reported that the concentration of blood glutathione was not affected by phytogetic feed additives.

Parameters	T1	T2	T3	T4	T5	SEM
MLA (U/mg Hb)	1.85 ^c	2.77 ^b	2.93 ^b	3.04 ^a	3.11 ^a	0.03
SDA (U/mg Hb)	35.7 ^b	39.8 ^b	40.7 ^a	43.5 ^a	45.3 ^a	1.21
GPx(U/mg Hb)	27.1 ^b	29.4 ^b	33.8 ^a	34.7 ^a	38.3 ^a	1.96
CAT (U/mg Hb)	54.2 ^a	45.7 ^b	42.5 ^b	41.6 ^b	40.1 ^b	0.52

Table 5: Antioxidant status of broiler chicks fed different levels of ATSM
Means in the same row with different superscripts differ significantly ($P < 0.05$)

Conclusion

Feed additives (plants extracts) have been reported to perform multiple biological activities including antibacterial, antifungal, antiviral, antihelminthic, antioxidant and immune modulator because they contain phytochemicals such as phenols, flavonoids, alkaloids, tannins, saponins, terpenoids etc. They are relatively cheap, safe and effective without any side effect on continuous use. The use of ATSM at 30 ml/litre of water have shown to be able to give total protection to the body and its metabolism against free radicals due to the presence of antioxidants and have no deleterious effect on the blood profile of broiler chicks [56-58].

References

- Santi DU, Kim IH (2017) Efficacy of phytogetic feed additive on performance production and health status of monogastric animals – A review. *Ann Anim Sci* 17(4): 929-948.
- Alagbe JO (2020) Performance, hematology and serum biochemical parameters of weaner rabbits fed different levels of fermented *Lagenaria brevifera* whole fruit extract. *Advances in Research and Reviews* 1: 5.
- Olafadehan OA, Oluwafemi RA, Alagbe JO (2020) Performance, haemato-biochemical parameters of broiler chicks administered Rolfe (*Daniellia oliveri*) leaf extract as an antibiotic alternative. *Drug Discovery* 14(33): 135-145.
- Gary DB, Richard DM (2002) Interrelationship between nutrition and immunity. VM 139 Series at the Veterinary Medicine – Large Animal Clinical Sciences Dept, UF/IFAS Extension.
- Karuppannan K, Priyadharshini SD, Sujatha V, (2013) Phytopharmacological properties of Albizia species: A review. *Int J Pharm Pharm Sci* 5(3): 70-73.
- Alagbe JO, Soares DM (2018) Effects of feeding different levels of *Azolla pinnata*, *Polyalthia longifolia*, *Tithonia diversifolia*, *Moringa olifera*, *Azadiracta indica* leaf meal infusion as an organic supplement on the performance and nutrient retention of growing grass cutters. *Greener Journal of Agricultural Sciences* 8(1): 1-11.
- Mohammed F, Singh PP, Irchhaiya R (2012) Review on *Albizia lebbecka* potent herbal drugs. *International Research Journal of Pharmacy* 3(5): 63-68.
- Ibrahim L, Afolabi LO, Adan AA, Usman JD, Muhammad U (2016) Analysis of some phytochemical and minerals found in aqueous stem bark extract of *Albizia lebbeck*. *Dutse Journal of Pure and Applied Sciences* 2(1): 231-237.
- Uwaya JO, Okunrobo LO, Igbinauwu PO (2017) *Albizia zygia*: A comparative investigation of phytochemical composition, proximate analysis and anti-seizure properties of methanol extracts and its leaves and stem bark. *Nigerian Journal of Pharmaceutical and Applied Science Research* 6(2): 76-80.
- Gupta M, Mazumder U, Kumar T, Gomathi P, Kumar S (2004) Antioxidant and hepato-protective effects of *Bulhinia racemosa* against paracetamol and carbon tetrachloride induced liver damage in rats. *Iran Journal of Pharmacological Therapy* 3(1): 12-20.
- Mc Donald SD, Prenzler P, Robards K, Antolovich M (2001) Phenolic content and antioxidant activity of olive extracts. *Food Chemistry* 73(1): 73-84.
- Saidu Y, Bilbis LS, Lawal M, Isezuo SA, Hassan SW, et al. (2007) Acute and sub chronic toxicity studies of crude aqueous extract of *Albizia chevalier*. *Asian Journal of Biochemistry* 2(4): 224-236.
- Alagbe JO (2019) Haematology, serum biochemistry, relative organ weight and bacteria count of broiler chicken given different levels of *Luffa aegyptiaca* leaf extracts. *International Journal of Advanced Biological*

and *Biomedical Research* 7(4): 370-380.

14. AOAC (2000) Association of Official Analytical Chemists. Official Methods of Analysis 19th Edition Washington, DC Pages 69-77.
15. Olubukola SO, Anthony JA, Adewale A (2015) Sub-chronic administration of methanolic whole fruit extracts of *Lagenaria breviflora* (Benth) roberly induces mild toxicity in rats. *Pharmacogn Mag* 11(Suppl4) : S516-S521.
16. Mahipal C, Ashak KP, Shalini B, Narayan D, Sunil EJ, et al. (2015) Dietary supplementation of a novel phytogetic feed additive: effects on nutrient metabolism, antioxidant status and immune response of goats. *Animal Production Science* 56(10): 1612-1621.
17. Duncan DB (1955) Multiple range and multiple F-test. *Biometrics* 11(1): 1-42.
18. National Research Council (1994) Nutrient requirement of poultry 9th Rev Edn, Washington DC National Academy Press.
19. Teodora P, Evgeni P, Maya I (2020) Effect of black soldier fly (*Hermetia illucens*) meals on the meat quality of broilers. *Agriculture and Food Science* 29(3): 177-188.
20. Aduku AO (2004) Animal nutrition in the tropics: Feeds and feeding in monogastric and ruminant nutrition. *Journal of Applied Poultry Research* 13: 628-638.
21. Alagbe JO, Oluwafemi RA (2019) Growth performance of weaner rabbits fed Noni (*Morinda citrifolia*) and *Moringa olifera* leaf mixture as partial replacement of soya bean meal. *Journal of Agriculture and Forest Meteorology Research* 2(4): 136-142.
22. Baker DH (2009) Advances in protein-amino acid nutrition of poultry. *Amino Acids* 37(1): 29-41.
23. Marc Rhoads J, Wu G (2009) Glutamine, arginine, and leucine signalling in the intestine. *Amino Acids* 37(1): 111-122.
24. Kimura H (2010) Hydrogen sulphide: from brain to gut. *Antioxid Redox Signal* 12(9): 1111-1123.
25. Tan B, Yin Y, Kong X, Li P, Li X, et al. (2010) L-Arginine stimulates proliferation and prevents endotoxin-induced death of intestinal cells. *Amino Acids* 38(4): 1227-1235.
26. Yin YL, Yao K, Liu ZJ, Gong M, Ruan Z, et al. (2010) Supplementing L-leucine to a low-protein diet increases tissue protein synthesis in weanling pigs. *Amino Acids* 39(5): 1477-1486.
27. Brosnan JT, Brosnan ME (2010) Creatine metabolism and the urea cycle. *Mol Genet Metab* 100(Suppl 1): S49-S52.
28. Wu X, Ruan Z, Gao YL, Yin YL, Zhou XH, et al. (2010) Dietary supplementation with L-arginine or N-carbamylglutamate enhances intestinal growth and heat shock protein-70 expression in weanling pigs fed a corn- and soybean meal-based diet. *Amino Acids* 39(3): 831-839.
29. Wu G, Bazer FW, Burghardt RC, Johnson GA, Kim SW, et al. (2010) Proline and hydroxyproline metabolism: implications for animal and human nutrition. *Amino Acids* 40(4): 1053-1063.
30. Wu G, Bazer FW, Burghardt RC, Johnson GA, Kim SW, et al (2010) Functional amino acids in swine nutrition and production. In: Doppenberg J, editor. Dynamics in animal nutrition. Wageningen (The Netherlands): Wageningen Academic Publishers; p. 69-98.
31. McKnight JR, Satterfield MC, Jobgen WS, Smith SB, Spencer TE, et al. (2010) Beneficial effects of L-arginine on reducing obesity: potential mechanisms and important implications for human health. *Amino Acids* 39(2): 349-357.
32. Pali SS, Kays CE, Deval C, Bruhat A, Fafournoux P, et al. (2009) Specificity of amino acid regulated gene expression: analysis of gene subjected to either complete or single amino acid deprivation. *Amino Acids* 37(1): 79-88.
33. Talebi A, Asri-Rezaei S, Rozeh-Chai R, Sahraei R (2005) Comparative studies on haematological values of broiler strains (Ross, Cobb, Arbo-acres and Arian). *International Journal of Poultry Science* 4(8): 573-579.
34. Subhadarsini M, Silpa MG (2020) Comparative haematology and biochemical parameters of Indigenous broiler chicken. *International Journal of Scientific Technology Research* 9(4): 972-979.
35. Islam MS, Lucky NS, Islam MR, Ahad A, Rahman MM, et al. (2004) Haematological parameters of fayoumi, assil and local chickens reared in Sylhet region in Bangladesh. *International Journal of Poultry Science* 3(2): 144-147.
36. Abdi-Hachesoo B, Talebi A, Asri-Rezaei S (2011) Comparative study on blood profile of indigenous and Ross-308 broiler breeders. *Global Veterinaria* 7(3): 238-241.
37. Fugdge AM (2000) Laboratory Medicine: Avian and Exotic Pets. WB Saunders Company, Philadelphia.
38. Nse Abasi NE, Mary EW, Uduak A, Edem EAO (2014)

- Haematological parameters and factors affecting their values. *Journal of Agricultural Science* 2(1): 37-47.
39. Omokore EO, Alagbe JO (2019) Efficacy of dried *Phyllanthus amarus* leaf meal as an herbal feed additive on the growth performance, haematology and serum biochemistry of growing rabbits. *International Journal of Academic Research and Development* 4(3): 97-104.
 40. Isaac LJ, Abah G, Akpan B, Ekaette IU (2013) Haematological properties of different breeds and sexes of rabbits. *Proceedings of the 18th Annual Conference of Animal Science Association of Nigeria*. (Pg 24-27).
 41. Ugwuene MC (2011) Effect of dietary palm kernel meal for maize on the haematology and serum chemistry of broiler turkey. *Nigerian J Anim Sci*13: 93-103.
 42. Alagbe JO, Shittu MD, Eunice Abidemi OJO (2020) Prospect of leaf extracts on the performance and blood profile of monogastric-A review. *International Journal of Integrated Education* 3(7): 122-127.
 43. Obajuluwa OV, Sanwo KA, Egbeyale LT, Fafiolu OA (2020) Performance, blood profile and gut morphometry of broiler chickens fed diets supplemented with Yohimbe (*Pausynistalia yohimbe*) and Larvacide. *Veterinary and Animal Science* 10: 100127.
 44. Olafadehan OA, Oluwafemi RA, Alagbe JO (2020) Carcass quality, nutrient retention and caeca microbial population of broiler chicks administered Rolfe (*Daniellia oliveri*) leaf extract as an antibiotic alternative. *Journal of Drug Discovery* 14(33): 146-154.
 45. Livingston ML, Cowieson AJ, Crespo R, Hoang V, Nogal B, et al. (2020) Effect of broiler genetics, age and gender on performance and blood chemistry. *Heliyon* 6(7): e 04400.
 46. Obiakaonu HO, Okoli IC, Opara MN, Okoro VMO, Ogbuewu IP, et al. (2012) Haematological and serum biochemical indices of starter broilers fed leaf meal of neem. *Journal of Agricultural Technology* 8(1): 71-79.
 47. Akande T, Odunsi AA (2012) Nutritive value and biochemical changes in broiler chickens fed detoxified castor kernel cake based diets. *African Journal of Biotechnology* 11(12): 2904-2911.
 48. Oluwafemi RA, Isiaka Olawale, Alagbe JO (2020) Recent trends in the utilization of medicinal plants as growth promoters in poultry nutrition- A review. *Research in: Agricultural and Veterinary Sciences* 4(1): 5-11.
 49. Abdel-Wareth AA, Lohakare JD (2014) Effect of dietary supplementation of peppermint on performance, egg quality and serum metabolic profile of Hy-Line brown hens during the late laying period. *Animal Feed Science and Technology* 197: 114-120.
 50. Krishnan S, Alden N, Lee K (2015) Pathways and functions of gut microbiota metabolism impacting host physiology. *Curr Opin Biotechnol* 36: 137-145.
 51. Alagbe JO (2019) Growth performance and haemato-biochemical parameters of broilers chicken fed different levels of *Parkia biglobosa* leaf extracts. *Academic Journal of Life Sciences* 5(12): 107-115.
 52. Hollman PC (2001) Evidence for health benefits of plant phenols: Local or systemic effects. *Journal of Science Food and Agriculture* 81(9): 842-852.
 53. Ojewuyi OB, Ajiboye TO, Adebajo EO, Balogun A, Mohammed AO (2014) Proximate composition, phytochemical and mineral contents of young and mature *Polyalthia longifolia* Sonn leaves. *Fountain Journal of Natural and Applied Sciences* 3(1): 10-19.
 54. Lin J, Hunkapiller AA, Layton AC, Chang YJ, Robbins KR (2013) Response of intestinal microbiota to antibiotic growth promoters in chickens. *Foodborne Pathog Dis* 10(4): 331-337.
 55. Lan PTN, Le Binh T, Benno Y (2003) Impact of two probiotic *Lactobacillus* strains feeding on fecal lactobacilli and weight gains in chicken. *J Gen Appl Microbiol* 49(1): 29-36.
 56. Cho JH, Kim HJ, Kim IH (2014) Effects of phytogenic feed additive on growth performance, digestibility, blood metabolites, intestinal microbiota, meat colour and relative weight after oral challenge with *Clostridium perfringens* in broilers. *Livestock Science* 160: 82-88.
 57. Oh J, Giallongo F, Frederick T, Pate J, Walusimbi S, et al. (2014) Effects of dietary Capsicum oleoresin on productivity and immune responses in lactating dairy cows. *J Dairy Sci* 98(9): 6327-6339.
 58. OIabanji RO, GO Farinu, JA Akinlade, OO Ojebiyi, AA Odunsi, et al.(2007) Studies on Haematological and Serum Biochemical Characteristics of Weaner Rabbits Fed Different Levels of Wild Sunflower (*Tithonia diversifolia* Hemsl A Gray) Leaf- Blood Meal Mixture. *International Journal of Applied Agriculture and Apiculture Research* 4 (1&2): 80-89.

