

Atherogenic Dyslipidemia: An Important Risk Factor for Cardiovascular Disease in Metabolic Syndrome and Type 2 Diabetes Mellitus Patients

Tangvarasittichai S*

Department of Medical Technology, Naresuan University, Thailand

***Corresponding author:** Surapon Tangvarasittichai, Department of Medical Technology, Faculty of Allied Health Sciences, Naresuan University, Phitsanulok, 65000 Thailand, Tel: +66-08-9638-8382; E-mail: surapon14t@yahoo.com

Review Article

Volume 2 Issue 1 Received Date: December 30, 2016 Published Date: February 03, 2017 DOI: 10.23880/doij-16000144

Abstract

Both metabolic syndrome (MetS) and type 2 diabetes mellitus (T2DM) are increased risk of cardiovascular disease (CVD) and coronary heart events. Lipid abnormalities in MetS and T2DM patients are the major role in atherogenesis development. These lipid disorders include both quantitative and qualitative abnormalities of lipoproteins the potentially atherogenic. Hypertriglyceridemia is the main quantitative abnormalities, correlated with hepatic VLDL over production and the reduction of both VLDL and IDL catabolism in circulation, and accelerated HDL catabolism to cause decreased HDL-Cholesterol levels. Increased triglyceride content in VLDL particles cause VLDL1 (large VLDL) and also increase in triglyceride content of LDL and HDL particle, small dense LDL particles, easy to cause LDL oxidation and apolipoproteins glycation. Although the LDL-cholesterol level in circulation is usually normal in MetS and T2DM patients, but these types of LDL particles show easy oxidized and reduced turn-over, which is potentially harmful. However, insulin resistance and the insulin deficiency are occurred in MetS and T2DM patients, may play the important role of dyslipidemia. Insulin has the important function and regulation in lipid metabolism. In addition with adipokine, both in the pro-inflammatory adipokines, it could play the important role in the pathophysiology of dyslipidemia.

Keywords: Atherogenic dyslipidemia; Lipoprotein; Insulin resistance; Metabolic syndrome; Type 2 diabetes mellitus

Atherogenic Dyslipidemia: An Important Risk Factor for Cardiovascular Disease in Metabolic Syndrome and Type 2 Diabetes Mellitus Patients

Introduction

The International Diabetes Federation (IDF) reported estimation of 246 million adults worldwide had type 2diabetes mellitus (T2DM) in 2008 and provided the expectation of the prevalence to reach 380 million by 2025 [1]. Estimation of the prevalence of T2DM in the USA was from 21.9 million to 30.3 (11.2%) by the year 2025 [2,3]. This increase in T2DM results from increase of the incidence or rise in new patients of T2DM, which is a consequence of obesity particularly in abdominal obesity, ageing and lack of exercise population. The World Health Organization (WHO) has estimated that over 1 billion persons are overweight and more than 300 million of obese worldwide [4]. In the United States, adults more than 60% are overweight and/or obese, and also increased in the number of both obese children and adolescents [5]. In the Lower Northern Region (seven provinces) of Thailand, adults aged ≥ 40 years of abdominal obesity (AO) men and women were 37% and 41.8% [6]. Obesity is now become an important health problem worldwide. Clinical evidence proved that AO is an effect or of cardiovascular disease (CVD) and risk of T2DM [7.8]. The development of AO promotes insulin resistance and inflammation, and/or altered hemostasis as risk factors for CVD [9]. The increasing prevalence of obesity and abdominal obesity is considered as the pandemic levels. This has been attributed to increase the adoption of energy-dense diets, physical inactivity and sedentary lifestyles as the consequence of economic globalization and urbanization [4]. Most health care systems are concerned or based on the drug treatment diseases which caused by the specific organism or agents after diseases merge. However, what is really needed for the pandemic of obesity.

Metabolic syndrome (MetS) is defined as the cluster of cardio metabolic disorders that individuals are increase risk of T2DM, coronary heart disease (CHD), and cardiovascular disease (CVD). The major components of MetS are abdominal obesity, glucose intolerance or diabetes, pre-hypertension (HT) or HT and dyslipidemia characterized by hypertriglyceridemia and low levels of high-density lipoprotein cholesterol (HDL-C). The definition of MetS is varies slightly different between each guidelines from the expert groups such as the World Health Organization (WHO), the European Group for the Study of Insulin Resistance, the National Cholesterol Education Program Third Adult Treatment Panel (NCEP ATP III), the International Diabetes Federation (IDF) and the American Heart Association/National Heart, Lung, and Blood Institute. However, all definitions include AO, as a major risk factor, with in the IDF guidelines AO as a prerequisite (Table 1). The guidelines of NCEP ATP III define MetS as the presence of ≥ 3 of the following abnormalities: waist circumference (WC) ≥102 cm (~40 in) in men or \geq 88 cm (~35 in) in women, elevated blood pressure (BP) (systolic BP ≥130 mm Hg or diastolic BP \geq 85 mm Hg), triglycerides (TG) \geq 150 mg/dL, HDL-C <40 mg/dL in men and <50 mg/dL in women, and fasting blood glucose \geq 110 mg/dL. By the American Diabetes Association (ADA) criteria modified the threshold for fasting glucose $\geq 100 \text{ mg/dL}$ for impaired fasting glucose. International Diabetes Federation (IDF) also modified the threshold for WC to ≥90 cm. in men and ≥80 cm. in women for Asian population. When using the NCEP ATP III definition, the prevalence of MetS in US adults is estimated 22% [10] to 34% [11], 39% as the IDF definition [11]. The IDF definition also identify a greater prevalence of MetS than, (approximately 80% increase) the NCEP ATP III definition do in many countries [12,13]. Increasing of the obesity especially AO consequence glowing the MetS and T2DM will correspond increase the incidence of CVD. There is the urgent need to improve the therapeutic strategies for CVD management in MetS and/or T2DM patients. The prevention of MetS and T2DM is based on the changes in lifestyle. However, neither private companies nor governments have provided any funds for these approaches. Obesity especially AO is an established risk factor for HT and T2DM, and as the major component of MetS. Not surprisingly, about the increasing prevalence of obesity is paralleled with the increasing of the number of HT and T2DM and/or MetS patients.

Central obesity(defined as waist circumference* with ethnicity specificvalues); plus any two of these following				
factors:				
Elevated TC levels	≥50 mg/dL (1.7 mmol/L)			
	or specific medication for lipid abnormality			
Reduced HDL-C levels	<40 mg/dL (1.03 mmol/L) in males			
	<50 mg/dL (1.29 mmol/L) in females			
	or specific medication for lipid abnormality			
Elevated Blood Pressure (BP)	systolic BP ≥130 or diastolic BP ≥85 mm Hg			
	or medication for hypertension			
Elevated fasting plasma glucose(FPG)	≥100 mg/dL (5.6 mmol/L),			
	or diagnosed and treated with type 2 diabetes mellitus			
	If above 5.6 mmol/L or 100 mg/dL, OGTT is strongly recommended.			
Country/Ethnic group	waist circumference			
Europids*In the USA, the ATP III	Male	≥94 cm		
values(102 cm male; 88 cm female)are				
likely to continue to be used for clinical	Female	≥80 cm		
purposes				
South Asians:	Male	≥90 cm		
South Asians: Chinese based, Malay and	Male	≥90 cm		
South Asians: Chinese based, Malay and Asian-Indian population	Male Female	≥90 cm ≥80 cm		
South Asians: Chinese based, Malay and Asian-Indian population	Male Female Male	≥90 cm ≥80 cm ≥90 cm		
South Asians: Chinese based, Malay and Asian-Indian population Chinese	Male Female Male Female	≥90 cm ≥80 cm ≥90 cm ≥80 cm		
South Asians: Chinese based, Malay and Asian-Indian population Chinese	Male Female Male Female Male	≥90 cm ≥80 cm ≥90 cm ≥80 cm ≥90 cm		

Table 1: The International Diabetes Federation (IDF) definition for the persons having the metabolic syndrome.

Cardiovascular Disease Risk in MetS and/or T2DM Patients

There are many clinical research evidences demonstrated that patients with MetS and/orT2DM are at the state of increased CVD risk [11,14-19]. In MetS definition is useful for the physicians to identify patients at high risk for CVD. In population-based cohort study of Finnish men (aged 42-60 years) without CVD or T2DM at baseline, the risk of death from CHD or CVD (≥12-yrs follow-up period) was significantly higher in participants with MetS [14]. In the epidemiologic studies in the United States have identified individuals with MetS also elevated risk of CVD [11,17-19]. The incidence of cardiovascular mortality was 12.0% in patients with MetS and 2.2% in MS individuals without (P<.001) [15]. Manv epidemiologic studies and clinical trials have demonstrated that T2DMpatients are at increased risk for CVD [20-23]. In the Multiple Risk Factor Intervention Trial, a cohort study of a large population (aged 35-57 yrs) without T2DM and with T2DM (same ages), the incidence of the >12 yrs follow-up for cardiovascular mortality was 11.7% in T2DM patients and 2.6% in non-T2DM individuals [21]. In the generally accepted that T2DM increases the CVD risk by 2- to 4-fold.Generally, we

known that the elevated CVD risk in MetS and T2DM patients is multi factorial, one of the risk factor in particular-atherogenic dyslipidemia. The excess CVD risk in patients with MetS and/or T2DM is due to several risk factors including both un-modifiable risk factors (age, gender and genetics) and traditional risk factors (HT, dyslipidemia, hyperglycemia and smoking). The overall cardio-metabolic risk is modified by the disorders of these lipoproteins complex and demonstrated the same components of the MetS and T2DM. The characteristics of atherogenic dyslipidemia which consists of elevated triglyceride-rich lipoproteins (TRLs), small dense lowdensity lipoprotein (sdLDL) and low high-density lipoprotein (HDL) cholesterol concentrations in plasma in both fasting and postprandial. This review aims to summarize our understanding of the pathophysiology of dyslipidemia and abnormalities of lipid metabolism in MetS and/or T2DM are the major risk factors of the CVD [20,24].Cardiovascular disease risk in T2DM patients is 2 -4 greater than in non-diabetic subjects [20,25-27], CVD in T2DM patients is the major cause of morbidity and mortality. Insulin resistance plays the major role in the pathophysiology of lipid abnormalities in MetS and T2DM. Addition with adipokines (leptin, and adiponectine) could be associated in the development of lipid abnormalities in these patients.

Brief Normal Human Lipoprotein Metabolism

Lipid is transported as lipoproteins, the biochemical complex for all hydrophobic lipids molecules transportation in the circulation. They have a single layer on the surface as monolayer of phospholipids, free cholesterol and apolipoproteins as the hydrophilic portions oriented outward surrounding by the water. The central inner is lipophilic portions, the non-water soluble cholesterol esters and triglycerides (TG) as spherical particles composed of a central core of each molecule oriented inwards the lipids molecules within the lipoprotein particles in the circulations. General, lipoproteins are classified according to their density following as: (i) chylomicron, (ii) very low density lipoprotein (VLDL), (iii) intermediate density lipoprotein (IDL), (iv) low density lipoprotein (LDL) and (v) high density lipoprotein (HDL). The brief schematic of normal lipoprotein metabolism is shown in (Figure 1).



Figure 1: Briefs chematic of normal human lipoprotein metabolism: LPL; lipoprotein lipase, HL; hepatic lipase, VLDL; very low density lipoprotein, IDL; intermediate density lipoprotein, LDL; low density lipoprotein, TG; triglycerides, CETP; cholesteryl ester transfer protein, CE; cholesteryl ester, HDL; high density lipoprotein, LCAT; lecithin-cholesterol acyl transferase, ABCA1; ATP binding cassette A1 transporter.

Chylomicrons

Chylomicrons are the largest lipoprotein particles. The major purpose is responsible for the dietary triglycerides and cholesterol transportation in the circulation. Chylomicrons are composed with cholesterol esters, TG (85%-90%), phospholipids and apolipoproteins (apoB48, apoA-I, apoA-IV). Chylomicrons are synthesized in the enterocytes, and the processing of the lipid components (TG, cholesterol esters, phospholipids) and apoB48 association is performed by the microsomal transfer protein (MTP). Chylomicrons are secreted into the lymphatic circulation before entering the circulation. In circulation, TG in chylomicrons is hydrolyzed by LPL releasing FFA for energy production or for reesterification or storage in adipose tissues. This chylomicron depleted TG leading to form chylomicronremnants, the smaller, TG-poorer particles. Chylomicronremnants are cleared by the liver via LDL B/E receptor or LDL-receptor related protein (LRP) and do not accumulated in the circulation.

VLDLs and IDLs

VLDL particles are the first lipoprotein particles that are synthesized and secreted by the liver. VLDL particles are composed endogenous cholesterol, TG (55% to 65%), phospholipids and apolipoproteins (apoB100, apoC and apoE). The synthesis of VLDL in the hepatocyte occurs in two major steps as follow: (i) pre-VLDL formation, is takes place in the rough endoplasmic reticulum. ApoB is co-translational and post-translational lipid mediated by the MTP. MTP transfers lipids (majorin TG but also cholesterol esters, phospholipids) to apoB [28]. (ii) This step is the conversion step of pre-VLDL to VLDL in the smooth membrane compartment by the ADP ribosylation factor-1 (ARF-1) and phospholipase deactivation [28]. Triglycerides, the major component of VLDLs are hydrolyzed by lipoprotein lipase in circulation. Triglycerides in VLDLs are become progressively reduced, while phospholipids and apolipoprotein C and E on the surface are transferred to HDLs. This metabolic process causes the IDL particles formation, which are cleared by the liver (via LDL B/E receptors) or further metabolized to form LDLs particles. For this metabolic process of LDL particles formation from IDLs is cause by hepatic lipase triglyceride consist with lipase and enzyme, phospholipase activities.

LDLs

LDL particles are the main cholesterol-carrying lipoprotein in circulation. LDL particles are the final production of the VLDL-IDL-LDL process. In each LDL particle contains one molecule of apoB100, which plays the major role in the LDL metabolism. LDL particles are cleared in the plasma mediated via the LDL B/E receptor. LDL B/E receptors are located on hepatic cells (70%) and the other cells of the body (30%).These cells can take up LDL-C from circulation. All cells of extra-hepatic tissues cannot be process these cholesterol which may accumulate in these cells.

HDLs

HDL particles are secreted as the small lipid-poor lipoproteins by the hepatocyte, a major lipoprotein containing is apoA-I, which receive in the circulation, and the others are apoC, apoE and phospholipids from chylomicrons and VLDLs. The original HDL lipoprotein particles (nascent or lipid-poor HDLs) are synthesized by the liver as the complexes of apolipoproteins and phospholipid, which form with cholesterol-free flattened spherical particles. These complexes are capable to pick up cholesterol from the cells cytoplasm by interaction with the ATP-binding cassette transporter A1 (ABCA1) [42]. Lecithin-cholesterol acyltransferase (LCAT) enzyme in circulation converts the free cholesterol into cholesteryl ester (a more hydrophobic form of cholesterol), which is sequestered into the core of the HDL lipoprotein leading to the formation of HDL3

particles. The fusion of 2 HDL3 particles formed a one larger size HDL2 particle that is promoted by phospho lipid transfer protein (PLTP). These HDL2 lipoproteins, cholesteryl ester rich, are degraded by hepatic lipase and endothelial lipase to form HDL remnant particles that are cleared by the liver via recognition of scavenger receptor class B type 1(SR-B1) receptor as direct pathway. HDL particles increase in size by more cholesterol and phospholipid molecules incorporation from cells and other lipoproteins, as they circulate through the circulation. Liver and the steroidogenic organs (adrenals, ovary, and testes) are the sites that HDL transports cholesterol to degrade by both direct and indirect pathways. In humans, cholesteryl ester transfer protein (CETP), the one indirect pathway exchanges TG of VLDL against cholesteryl esters of HDL. After that VLDLs are hydrolyzed to LDL and removed from the circulation via the LDL receptor. Triglycerides in HDL are not stable and degraded by hepatic lipase to generate small HDL particles and restart to uptake cholesterol from the cells. The HDL cholesterol delivery to other organs (adrenals, ovaries, and testes) is important for steroid hormones synthesis. The delivered cholesterol in the liver is excreted into the bile and intestine after conversion into bile acids. The importance step of the HDL participate in cholesterol metabolism may the transportation from the foam cells (lipidladen macrophages of atherosclerotic arteries) to the liver for secretion into the bile. This step has been termed reverse cholesterol transport as the protective function of HDL for atherosclerosis.

There are two major lipid transfer proteins (CETP and PLTP) involved in lipoprotein metabolism. Among of these, CETP facilitates the transfer of TG from VLDL (TG-rich lipoproteins) to HDL and LDL particles, and also transfer cholesteryl esters from HDL and LDL to VLDL particles. While, PLTP will facilitates the transfer of phospholipids and α -tocopherol between the lipoproteins particles and also involved in the HDL2 particles formation from HDL3 particles. Any modification from CETP or PLTP activities will promote the qualitative abnormalities of lipoproteins particles.

Role of Insulin on Lipid Metabolism

Insulin plays the regulation role of the lipid metabolism. Figure 2 demonstrated the main sites of insulin action in lipoprotein metabolism. Insulin inhibits hormone-sensitive lipase in adipose tissue as the anti-lipolytic action and promotes TG storage in the adipocytes and reduces the secretion of free fatty acids from adipose tissue into the circulation. Insulin also inhibits VLDL

Tangvarasittichai S. Atherogenic Dyslipidemia: An Important Risk Factor for Cardiovascular Disease in Metabolic Syndrome and Type 2 Diabetes Mellitus Patients. Diabetes Obes Int J 2017, 2(1): 000144.

synthesis from the liver. It has been demonstrated that insulin decreased the VLDL-TG synthesis (by 67%) and decreased of VLDL-apoB synthesis (by 52%) in normal subjects [30,31]. Insulin diminish free fatty acids, the substrates of VLDL in circulation (as the anti-lipolytic effect) resulting in reduced VLDL synthesis, and also by the direct inhibition effect in the hepatocytes [31]. Insulin is the activator on the lipoprotein lipase (LPL) gene to promote LPL synthesis [32]. Insulin is also an activator and enhancer of LPL activity to promote the TG-rich lipoproteins catabolism [33]. Insulin is also enhances LDL B/E receptor expression and activity to promote the clearance of LDL in the circulation [34,35]. Insulin is also activates on LCAT and hepatic LPL activities to promote in the action on HDL metabolism [36]. It has been demonstrated that insulin has an inhibitory effect on the PLTP activity both in normal subjects and T2DM patients [37]. Arii et al. demonstrated that insulin reduces CETP activity, it is not a direct inhibitory action on CETP but this inhibitory effect depend on the consequence of the insulin-induced reduction of free fatty acids in the circulation [38].

Atherogenic Dyslipidemia Pattern

Lipoproteins in circulation exist as the spectrum of particles size difference and the difference in

atherogenicity or anti-atherogenic potential (Figure 2). Metabolic syndrome and T2DM diabetes mellitus as the insulin-resistant state, exhibit the characteristic pattern of abnormalities in serum lipids: low levels of HDL-C and elevated TG and sdLDL [39,40]. This dyslipidemia pattern is also demonstrated sapolipo protein B (apoB) elevations and the sdLDL particles are depleted in cholesteryl ester (Table 2) [41]. The elevation of apoB-carrying lipoproteins of dyslipidemia is also decreases in apolipoprotein A-I-carrying lipoproteins as the central abnormalities. This complex dyslipidemia is termed dyslipidemia of insulin resistance or diabetic dyslipidemia. Then, insulin resistance reflects the underlying dyslipidemia state or plays the major role in the increased cardiovascular risk in MetS and T2DM patients with increased atherogenic dyslipidemia. Dvslipidemia pattern demonstrated lipoprotein abnormalities in both quantitative and qualitative phenomena in these patients [42-48]. Hypertriglyceridemia and low HDL-cholesterol levels is the main quantitative lipid abnormalities while the qualitative lipid abnormalities are occur in all lipoproteins. These lipoprotein abnormalities may promote atherosclerosis in these patients. Main lipoprotein modifications are shown in Table 2 and in (Figure 3).

Lipoproteins	Plasma level	Abnormalities	Qualitative or type of abnormalities
VLDL	Hypertriglyceridemia	Increased production Decreased catabolism	-Large VLDL (VLDL1) -Glycation
LDL	Normal or slightly increased	Decreased catabolism Decreased turn-over	-sdLdL (TG-rich LDL) -Oxidation -Glycation
HDL	Decreased HDL	Increased catabolism	-TG-rich HDL -Glycation

Table 2: The major lipid abnormalities in MetS and T2DM.



Figure 2: The major point effects by insulin on lipoprotein metabolism:

For insulin activates [+ve]: LPL; HL; LCAT; LDL B/E receptors.

For insulin inhibits [-ve]: hepatic VLDL production, hormone-sensitive lipase.LPL; lipoprotein lipase, HL; hepatic lipase, VLDL; very low density lipoprotein, IDL; intermediate density lipoprotein, LDL; low density lipoprotein, TG; triglycerides, CETP; cholesteryl ester transfer protein, CE; cholesteryl ester, HDL; high density lipoprotein, LCAT; lecithin-cholesterol acyl transferase, ABCA1; ATP binding cassette A1 transporter.



Figure 3: The major qualitative or types of lipid abnormalities in MetS and T2DM: Qualitative or type of lipid abnormalities

- Hypertriglyceridemia (increased VLDL production),
- -Increased HDL catabolism due to low HDL-C levels,
- -Increased large VLDL (VLDL1) production,
- -Increased sdLDL (TG-rich) and oxidized LDL,
- -Increased CETP activity (due to increased transfer TG to LDLs and HDLs)
- -Increased TG content in HDL,
- -Increased taken up of VLDL1, sdLDL and oxidized LDL by macrophages.
- -Decreased LDL B/E receptors (reduction of LDL turn over),
- -Decreased TG catabolism in TG-rich lipoprotein (due to reduced LPL activity).

Triglyceride-rich Lipoproteins

Triglyceride levels are increased in both MetS and T2DM, due to the elevation of the number of VLDL particles in circulation [42,46] and also in the number of IDL particles was observed [49,50]. The major determinant of the hypertriglyceridemia is the VLDL-TG overproduction in the circulation of both MetS and T2DM patients [51]. This may cause increase and accumulate a large number of lipids in the hepatocytes and also to cause hepatic resistance to insulin for the inhibitory effect of VLDL production [52-54]. In this stage, increasing of lipid pool in hepatocytes is the increase flux of free fatty acids (FFA) into the liver [55] and may accelerate to increase de novo lipogenesis, which correlates with the accumulation of fat in liver. This evidence was demonstrated by the observation of sterol regulatory element-binding protein-1c (SREBP-1c) elevation in the liver of insulin-resistant animals [56,57]. The SREBP-1c expression in the liver is associated with the 2 enzymes of de novo lipogenesis: (i) acetylCoA carboxylase and (ii) fatty acid synthase activation. Many studies demonstrate the decreasing of the inhibitory effect of insulin on the hepatic VLDL production is the major role [31,54] and elevated FFA level in circulation [58]. Many studies have been demonstrated these inhibitory effects of insulin on hepatic VLDL production [30,31,59-65] and also demonstrated the inhibition of VLDL particles formation and secretion by increased apoB degradation and reduction of the MTP expression in hepatocytes [61,62,64]. Elevation of MTP expression lead to enhance VLDL assembly and secretion has been demonstrated in insulin-resistant hamsters, obese diabetic mice [65,66] and T2DM patients. In normal subject, insulin also activated enzyme phosphoinositide 3-kinase (PI 3-kinase) using for the phosphoinositol biphosphate (PIP2) transformation to phosphoinositol triphosphate (PIP3). The reduction of PIP2 was induced by insulin is the activator of ADP ribosylation factor-1 (ARF-1) and phospholipase D which leads to decrease in the ARF-1 and phospholipase D activation that were involved in VLDL formation[28]. In T2DM present the defective in PI 3kinase activation from the insulin resistant state. This situation in T2DM causes an elevation of PIP2 that activates ARF-1 and Phospholipase D leading to increase VLDL formation. Thus, VLDL overproduction, in both MetS and T2DM may due to hepatic resistance to insulin action of the inhibitory effect of insulin on hepatic VLDL production [30,67]. In T2DM, overproduction of VLDL-TG may be greater than in the production of VLDL-apoB result from the larger TG-rich VLDL particles (VLDL1) formation [68] and also reduced in catabolism of VLDL and IDL particles as the factors in promoting diabetic hypertriglyceridemia in T2DM [51,68,69]. Both increased VLDL and IDL particles mainly effects on the reduction of lipoprotein lipase activity. Many studies have been demonstrated the reduction of adipose tissue lipoprotein lipase activity in T2DM [68,70,71]. Since insulin action is a major activator of lipoprotein lipase. Then, reduction in insulin activity may be relates with insulin deficiency and/or insulin resistance, observed in T2DM and MetS. There are several qualitative abnormalities of VLDL particles have been demonstrated in T2DM patients. The mainly overproduction of VLDL is an increased large VLDL particles synthesis (VLDL1), characterized by large amount of TG than the smaller VLDL2 particles. VLDL1 are easily taken up by the scavenger receptors of macrophages and accumulate of lipid within macrophages to cause foam cells formation in vessel walls [72]. Type 2 diabetes mellitus patients demonstrate both in fasting hypertriglyceridemia and postprandial TG-rich hypertriglyceridemia. The majority of lipoproteins during the postprandial state in T2DM patients are VLDL1 particles [73,74]. Apolipoproteins (apoB, apoC, apoE) glycation in VLDL particles may occur in T2DM and cause this glycated VLDL particle and the B/E receptor binding reduction on the catabolism [75].

LDL

Low density lipoprotein-cholesterol (LDL-C) is usually normal level in T2DM patients but the modifications in LDL particles metabolism are observed. In vivo study has been demonstrated the featuring of LDL-cholesterol values in T2DM similar with non-diabetic controls [69]. However, T2DM demonstrate the reduction in the turnover of LDL particles by the reduction of LDL catabolism, leading to increase LDL circulation and to promote cholesterol deposition in the vessel wall. Duvillard et al. [76] reported insulin treatment in T2DM patients can be normalized LDL catabolism. The reduction of the number of LDL B/E receptors is the other caused of impaired LDL catabolism in T2DM. Indeed, T2DM patients demonstrate the significant LDL B/E receptors reduction on cell surface [77], but insulin treatment can restore the number of LDL B/E receptors on cell surface in T2DM patients [77]. Apo Bglycation on the LDL particles could be decrease affinity of LDLs with their receptors to cause the reduction in LDL catabolism [78]. The important atherogenic LDLs in T2DM patients are small dense, triglyceride-rich, LDL particles [79,80] and this appears to be mainly related to hypertriglyceridemia phenotype in T2DM [81]. Then, VLDL1-TG is the major predictor of LDL size in T2DM [73]. Elevation of TG-rich lipoproteins stimulates CETP activity in T2DM to promote TG transfer to LDLs to form TG-rich LDL particles which are favor

Tangvarasittichai S. Atherogenic Dyslipidemia: An Important Risk Factor for Cardiovascular Disease in Metabolic Syndrome and Type 2 Diabetes Mellitus Patients. Diabetes Obes Int J 2017, 2(1): 000144.

substrate for hepatic lipase to produce sdLDL particles. Many studies have been demonstrated presence of sdLDL particles is associated with increased cardiovascular risk [82-84]. Many research studies indicate that sdLDL particles have atherogenic properties as follow: (i) sdLDL particles reduce affinity for the LDL B/E receptor and are easy taken up by the scavenger receptor of macrophages to form the foam cells (ii) sdLDL particles have higher affinity for intimal proteoglycans to penetrate into the vessel wall than large LDL particles [85,86] (iii) patients with sdLDL particles elevation demonstrated an impaired response to the endothelium acetylcholine vasodilatator [87,88] (iv) sdLDL particles are susceptible for oxidation [88,89], LDL oxidation is the marked atherogenic potential that observed in increased in T2DM [47,48,90,91]. Oxidized LDLs are rapid uptake by macrophages resulting in foam cell formation. Oxidized LDLs produce chemotactic stimulates monocytes by increasing the adhesion molecules formation (such as intercellular adhesion molecule 1; ICAM-1) from endothelial cells. Oxidized LDLs also stimulate the macrophages cytokines production including $TNF\alpha$, IL1, as the amplifiers of the inflammation of atherosclerotic process. Indeed, glycated LDL (glycation of apoB within LDL particles) observed in T2DM patients to cause the reduction in affinity of LDL B/E receptor effect in LDL metabolism [92-94] and are also favor taken up by macrophages to cause foam cells formation [95]. Furthermore, this glycated LDL is easy oxidized in the circulation [96,97].

HDL

The decrease in HDL-cholesterol in T2DM patients related to the reduction in HDL2 sub-fraction [98,99] and elevation of HDL particles catabolism [100,101]. The HDL2 level reduction in T2DM has been demonstrated correlation with both hypertriglyceridemia and obesity. Many studies indicated the elevation of hepatic lipase activity in HDL catabolism, is observed in T2DM patients [100,102]. Elevation of HDL catabolism is associated with insulin-resistant state and obese insulin resistant nondiabetic patients [103,104]. This elevation of HDL particles catabolism in these states will increase TG-rich lipoproteins pool (mainly VLDL). The large amount of TGrich lipoproteins in circulation, CETP can transfer TG from TG-rich lipoproteins to HDL particles to form TG-rich HDL particles [105]. Now, TG-rich HDL particles become the good substrate for hepatic lipase which increased HDL particles catabolism in insulin resistant states and T2DM. This catabolism process is responsible for increased HDL particles catabolism. ApoA-I dissociated from TG-rich HDL particles, is filtered and degraded by the renal glomerular and renal tubular cells. Furthermore, HDL particles in T2DM can be glycated and demonstrated the correlation between glucose level with apoA-I glycation [106]. The apoA-I glycation may reduce the binding of HDL to its receptor [107]. These abnormalities of HDL particles may cause HDL dysfunction in mediated cholesterol efflux and the process of reverse cholesterol transport [108-110].

Lipid Transfer Proteins

In the qualitative abnormalities of lipoproteins such as TG-rich LDL and TG-rich HDL particles are occur in insulin resistance state and T2DM patients. This may indicate an increased activity of CETP in the transfer of TG and cholesterol esters between lipoproteins [111,112]. Both hypertriglyceridemia (TG-rich lipoproteins) and the hyperglycemia accelerated lipoproteins glycation may direct stimulate and increase CETP activity in insulin resistance state and T2DM [113]. Many research studies reported increased PLTP mass and activity in T2DM [112,114]. But the exact process of this increased PLTP activity remains unclear.

Some Roles of Adipokines Relate the Pathophysiology of Dyslipidemia

Beyond the storage fat, visceral, subcutaneous depots and widely dispersed throughout the body of adipose tissue may be participate in influencing cardiovascular disease. In the present day, white adipose tissue is well recognized as an endocrine tissue function. Its can produce several adipokines levels in circulation and are altered in obesity and T2DM. The difference of adipose tissue depots in one with another is based on the relative adipokine levels production. In obesity will favor to produce pro-inflammatory adipokines [115-117]. Interestingly, in research studies of visceral adipose tissue in aging mice demonstrate the expression of pro inflammatory adipokines (TNF α and IL-6) even the absence of diet-induced obesity [118,119]. We can classify adipokines to pro-inflammatory and anti- proinflammatory adipokines. Most adipokines are identified as pro-inflammatory and they are up-regulated in the obese state and T2DM.These adipokines functions are promoting metabolic and cardiovascular diseases in these conditions. Pro-inflammatory adipokines include $TNF\alpha$, leptin, IL-6, resist in, RBP4, lipocalin 2, IL-18 and ANGPTL2. The smaller number adipokines are anti-proinflammatory factors. These adipokines include adiponectin [120,121] and SFRP5 [122]. Inflammatory stimulation increase leptin levels both in adipose tissue and plasma [123].

Leptin can stimulate the multiple types of immune cells (monocytes/macrophages, neutrophils, and T cells), to release the inflammatory cytokines [124-127]. Leptin increases TH1-type cytokines production and suppresses TH2-type cytokine IL-4 production in T cells [124], leading T cells polarizing to TH1 cell phenotype. Consistent with the observations of leptin deficiency can protect against liver damage as in the models of T cellmediated hepatitis and autoimmune encephalomyelitis [124,128,129]. By these evidences accepted that leptin plays the major role in pro-inflammatory adipokine. Many research studies demonstrated the important role of leptin in cardiovascular diseases. Elevation of leptin levels are identified in patients with myocardial infarction [130] and heart failure [131]. Faraj et al. reported adiponectin and/or TNF α involve in lipid metabolism [132]. Thus, adipokines could play the pathophysiology role in dyslipidemia of obese and T2DM. Adiponectin increases FFA uptake and oxidation in muscle and decreases TG content in muscle, liver and also reduces FFA level in plasma [132-134]. Many research studies reported the reduction of adiponectin levels and expression in white adipose tissue of obese and T2DM patients [135,136] and also demonstrate the negative correlation with plasma TG levels and positive correlation with plasma HDLcholesterol in T2DM and non-T2DM subjects [137-140]. The associations of adiponectin and plasma lipids are independence from insulin-resistance indexes [137-139]. Ng et al. reported the positive correlation of circulating adiponectin level with VLDL apoB catabolism, independent from HOMA-IR index [141].

These research data suggest the action of adiponectin on lipid metabolism independent from the action of insulin. However, the opposite mechanisms to decrease adiponectin may affect lipid metabolism in obese and T2DM still unclear. Adiponectin may reduce plasma triglyceride level via promoting of FFA oxidation through acetyl CoA oxidase, carnitine palmitoyltransferase-1 and AMP kinase activation [142]. Adiponectin may indirectly stimulate lipoprotein lipase activity [143] via the expression of PPAR- α in the liver and adipocytes [134]. Production of tumor necrosis factora (TNFa), proinflammatory adipokine, from white adipose tissue is increased in obesity and T2DM [132,144,145]. TNF α has been demonstrated involved in lipid metabolism. TNFa reduces FFA uptake in the adipose tissue, promotes lipolysis and FFA efflux [132]. TNF α also suppresses lipoprotein lipase, fatty acid transport protein and acetyl CoA synthetase production which involved in triglyceride accumulation [132, 139]. Indeed, no correlations were found between plasma TNF α and lipids levels in obese or T2DM patients [140] and has not found the association of TNF α and TG-rich lipoprotein metabolism [141,146]. Resist in, another Pro-inflammatory adipokine is found negative correlation with HDL-cholesterol levels in T2DM subjects, after adjustment for BMI and HbA1c [140]. However, also found no correlations between resist in and TG-rich lipoprotein metabolism [141,146]. Apelin is another adipokine effects on feeding behavior and glucose utilization. Sörhede Winzell et al. demonstrated that apelin can activate both apelin receptor and APJ receptor expression in islets for inhibiting the insulin secretion [147], and PI3K–phosphodiesterase 3B activation [148]. Ringström et al. [149] demonstrated that apelin can express by itself in β - and α -cells of pancreatic islets possible for autocrine/paracrine effects.Sfrp5 is the one of the anti-inflammatory adipokine.

Sfrp5 was identified as the soluble modulator of Wnt proteins to protect against metabolic dysfunction [150]. Sfrp5-deficient mice with high caloric diet feeding display impaired glucose catabolism and increased lipid accumulation in liver, even metabolically normal and on regular diet [150]. Association of Sfrp5-deficiency with increased lipid accumulation in macrophages and increased pro-inflammatory cytokines production. The mechanism causes by the deletion of JUN N-terminal kinase 1 (JNK1). Because of the Sfrp5 suppress Wnt5a which mediated the JUN N-terminal kinase 1 (JNK1) activation in adipose tissue. Then, in Sfrp5-deficient mice reverses the metabolic and inflammatory phenotypes. Thus, the over activation of JNK1 signaling in Sfrp5deficient mice induces increased inflammation and metabolic dysfunction in adipose tissue. These consist with the role of JNK1 in insulin resistance regulation and lipid inflammation [151,152,153]. Many studies have been reported the detection of Wnt5a expression in lesions of atherosclerosis in mouse and human [154,155]. Furthermore studies are needed to identify the putative role of adipokines on lipid abnormalities.

Using Lipoprotein Ratios for Insulin Resistance Estimation

According in described above, the major characteristics of quantitative dyslipidemia in MetS and T2D patients are increase plasma TG levels, reduce HDL-C levels and sdLDL particles, increased TG-rich remnant lipoprotein (TGRLs) and increase insulin levels in circulation[156]. The major change is increased TGRLs and decreased HDL-C levels are associated with insulin resistance syndrome. Insulin involves in the role of TG metabolism (Figure 2). In normal condition, TGRLs particles were less synthesized by the distinct pathways than VLDL particles synthesis [73,157]. In insulin resistant state or T2DM, insulin fails

Tangvarasittichai S. Atherogenic Dyslipidemia: An Important Risk Factor for Cardiovascular Disease in Metabolic Syndrome and Type 2 Diabetes Mellitus Patients. Diabetes Obes Int J 2017, 2(1): 000144.

to suppress the synthesis of VLDL particles [52-54], and associate with increased FFAs flow to the liver and increased lipid synthesis in the liver and decreased VLDL particles clearance from the circulation resulting in the elevated VLDL particles [54,158]. These phenomena indicated the problems of VLDL and HDL particles and concurrent with increased insulin levels. Reduction of HDL-C level is correlated with the insulin resistance or hyperinsulinemia and defected in insulin signaling for insulin-mediated glucose disposal [159]. These features are associated with the risk factors for coronary heart disease in obesity, MetS and T2DM patients. We can use these lipoprotein ratios such as TC/HDL-C, TG/HDL-C ratios and non-HDL-C (as TC - HDL-C) to estimate insulin resistance. Then, TC/HDL-C, TG/HDL-C ratios and non-HDL-C (as TC - HDL-C) were used as the markers for insulin resistance estimation. Tangvarasittichai et al. reported the using of TC/HDL-C, TG/HDL-C ratios and non- HDL-C as markers of insulin resistance and CVD risk factor [157,160] and reported the cut-off points of the highest % sensitivity and % specificity of TC/HDL-C, TG/HDL-C ratios and non-HDL-C corresponding to 3.58, 2.48 and 130.4, respectively [157]. These results of lipoprotein ratios were from Asian subjects and lower than the subjects from Western [161-163]. All of these lipoprotein ratios, TC/HDL-C, TG/HDL-C ratios and non-HDL-C are the simple mathematical analysis and easy to calculate and order with the lipid profiles available for clinician and no costs gain. Then, we can use these lipoprotein ratios as markers of insulin resistance estimation. For atherosclerotic risk assessment in higher risk subjects; obesity, MetS and T2DM patients need more attention for lipid screening.

Conclusion

Increased prevalence of the MS and T2DM is global as the consequence of the obesity and abdominal obesity pandemic. Several factors in the definition of MetS are the major characteristics of atherogenic dyslipidemia same as T2DM. The same lipid disorder in both cases contributes to increase the CVD risk in these individuals. These characteristics of lipid disorders include quantitative and qualitative abnormalities. Hypertriglyceridemia and low HDL-cholesterol levels are the main lipid quantitative abnormalities, include large VLDL particles (VLDL1), sdLDLs, increase TG content in LDLs and HDLs, increase apolipoproteins glycation and increase oxidized LDL. The involvement of adipokines in pathophysiology of lipid abnormalities in MetS and T2DMis complex and not complete explained. Need more studies to get insight into the in the detail mechanisms of these dyslipidemia. The improvement in understanding of lipid abnormalities and disorders in MetS and T2DM will get the better treatment and outcome of these dyslipidemia.

References

- 1. Editorial (2008) The global challenge of diabetes. Lancet 371(9626): 1723.
- 2. Wild SH, Farouhi NG (2007) What is the scale of the future diabetes epidemic and how certain are we about it? Diabetologica 50(5): 903-905.
- 3. Wild S, Roglic G, Green A, Sicree R, King H (2004) Global prevalence of diabetes. Estimates for the year 2000 and projections for 2030. Diabetes Care 27(5): 1047-1053.
- 4. World Health Organization (2003) Obesity and Overweight: World Health Organization global strategy on diet, physical activity and health fact sheet.
- 5. Wyatt SB, Winters KP, Dubbert PM (2006) Overweight and obesity: prevalence, consequences, and causes of a growing public health problem. Am J Med Sci 331(4): 166-174.
- Tangvarasittichai S, Seangsuk C, Chaisomboon C, Meemark S, Tangvarasittichai O (2015) Association of abdominal obesity, hypertriglyceridemia, and hypertriglyceridemic waist phenotype with hypertension and type 2 diabetes mellitus. Int J Diabetes Dev Ctries 35(4): 439-447.
- Okosun IS, Chandra KM, Choi S, Christman J, Dever GE, Prewitt TE (2001) Hypertension and type 2 diabetes comorbidity in adults in the United States: risk of overall and regional adiposity. Obes Res 9(1): 1-9.
- 8. Okosun IS, Dever GE (2002) Abdominal obesity and ethnic differences in diabetes awareness, treatment, and glycemic control. Obes Res 10(12): 1241-1250.
- Karter AJ, D'AgostinoJr RB, Mayer-Davis EJ, Wagenknecht LE, Hanley AJ, et al. (2005) Abdominal obesity predicts declining insulin sensitivity in nonobese normoglycaemics: the Insulin Resistance Atherosclerosis Study (IRAS). Diabetes Obes Metab 7(3): 230-238.
- 10. Ford ES, Giles WH, Dietz WH (2002) Prevalence of the metabolic syndrome among US adults: findings from

the third National Health and Nutrition Examination Survey. JAMA 287(3): 356-359.

- 11. Ford ES (2005) Prevalence of the metabolic syndrome defined by the International Diabetes Federation among adults in the U.S. Diabetes Care 28(11): 2745-2749.
- 12. Athyros VG, Ganotakis ES, Elisaf M, Mikhailidis DP (2005) The prevalence of the metabolic syndrome using the National Cholesterol Educational Program and International Diabetes Federation definitions. Curr Med Res Opin 21(8): 1157-1159.
- 13. Harzallah F, Alberti H, Ben Khalifa F (2006) The metabolic syndrome in an Arab population: a first look at the new International Diabetes Federation criteria. Diabet Med 23(4): 441-444.
- 14. Lakka HM, Laaksonen DE, Lakka TA, Niskanen LK, Kumpusalo E, et al. (2002) The metabolic syndrome and total and cardiovascular disease mortality in middle-aged men. JAMA 288(21): 2709-2716.
- 15. Isomaa B, Almgren P, Tuomi T, Forsén B, Lahti K, et al. (2001) Cardiovascular morbidity and mortality associated with the metabolic syndrome. Diabetes Care 24(24): 683-689.
- 16. Hu G, Qiao Q, Tuomilehto J, Balkau B, Borch-Johnsen K, et al. (2004) Prevalence of the metabolic syndrome and its relation to all-cause and cardiovascular mortality in nondiabetic European men and women. Arch Intern Med 164(10): 1066-1076.
- 17. Alexander CM, Landsman PB, Teutsch SM, Haffner SM, Third National Health and Nutrition Examination Survey (NHANES III); National Cholesterol Education Program (NCEP) (2003) NCEP-defined metabolic syndrome, diabetes, and prevalence of coronary heart disease among NHANES III participants age 50 years and older. Diabetes 52(5): 1210-1214.
- Hunt KJ, Resendez RG, Williams K, Haffner SM, Stern MP, et al. (2004) National Cholesterol Education Program versus World Health Organization metabolic syndrome in relation to all cause and cardiovascular mortality in the San Antonio Heart Study. Circulation 110(10): 1251-1257.
- 19. McNeill AM, Rosamond WD, Girman CJ, Golden SH, Schmidt MI, et al. (2005) The metabolic syndrome and 11-year risk of incident cardiovascular disease in

the atherosclerosis risk in communities study. Diabetes Care 28(2): 385-390.

- 20. Haffner SM, Lehto S, Ronnemaa T, Pyörälä K, Laakso M (1998) Mortality from coronary heart disease in subjects with type 2 diabetes and in nondiabetic subjects with and without prior myocardial infarction. N Engl J Med 339(4): 229-234.
- 21. Stamler J, Vaccaro O, Neaton JD, Wentworth D, The Multiple Risk Factor Intervention Trial Research Group (1993) Diabetes, other risk factors, and 12-yr cardiovascular mortality for men screened in the Multiple Risk Factor Intervention Trial. Diabetes Care 16(2): 434-444.
- 22. Almdal T, Scharling H, Jensen JS, Vestergaard H (2004) The independent effect of type 2 diabetes mellitus on ischemic heart disease, stroke, and death: a population-based study of 13,000 men and women with 20 years of follow-up. Arch Intern Med 164(13): 1422-1426.
- 23. Koskinen P, Manttari M, Manninen V, Huttunen JK, Heinonen OP, et al. (1992) Coronary heart disease incidence in NIDDM patients in the Helsinki Heart Study. Diabetes Care 15(7): 820-825.
- Turner RC, Millns H, Neil HAW, Stratton IM, Manley SE, et al. (1998) Risk factors for coronary artery disease in non-insulin dependent diabetes mellitus: United Kingdom prospective diabetes study (UKPDS: 23). BMJ 316(7134): 823-828.
- 25. Pyorola K, Laakso M, Uusitupa M (1987) Diabetes and atherosclerosis: an epidemiologic view. Diabetes Metab Rev 3(2): 463-524.
- 26. deVegt F, Dekker JM, Ruhe HG, Stehouwer CD, Nijpels G, et al. (1999) Hyperglycaemia is associated with allcause and cardiovascular mortality in the Hoorn population: the Hoorn Study. Diabetologia 42(8): 926-931.
- 27. Kannel WB, McGee DL (1979) Diabetes and cardiovascular risk factors: the Framingham study. Circulation 59(1): 8-13.
- OlofssonSO, Stillemark-Billton P, Asp L (2000) Intracellular assembly of VLDL: two major steps in separate cell compartments. Trends Cardiovasc Med 10(8): 338-345.
- 29. Huang CX, Zhang YL (2013) The target of regulating the ATP-binding cassette A1 protein (ABCA1):

¹² H

Tangvarasittichai S. Atherogenic Dyslipidemia: An Important Risk Factor for Cardiovascular Disease in Metabolic Syndrome and Type 2 Diabetes Mellitus Patients. Diabetes Obes Int J 2017, 2(1): 000144.

promoting ABCA1-mediated cholesterol efflux in different cells. Curr Pharm Biotechnol 14 (6): 623-631.

- Lewis GF, Uffelman KD, Szeto LW, Weller B, Steiner G (1993) Effects of acute hyperinsulinemia on VLDL triglyceride and VLDL apo B production in normal weight and obese individuals. Diabetes 42(6): 833-842.
- 31. Malmström R, Packard CJ, Caslake M, Bedford D, Stewart P, et al. (1998) Effects of insulin and acipimox on VLDL1 and VLDL2 apolipoprotein B production in normal subjects. Diabetes 47(5): 779-787.
- 32. Fried SK, Russell CD, Grauso NL, Brolin RE (1993) Lipoprotein lipase regulation by insulin and glucocorticoid in subcutaneous and omental adipose tissues of obese women and men. J Clin Invest 92: 2191-2198.
- 33. Brunzell JD, Schwartz RS, Eckel RH, Goldberg AP (1981) Insulin and adipose tissue lipoprotein lipase activity in humans. Int J Obes 5(6): 685-694.
- 34. Chait A, Bierman EL, Albers JJ (1979) Low-density lipoprotein receptor activity in cultured human skin fibroblasts. Mechanism of insulin induced stimulation. J Clin Invest 64(5): 1309-1319.
- 35. Mazzone T, Foster D, Chait A (1984) *In vivo* stimulation of low-density lipoprotein degradation by insulin. Diabetes 33(4): 333-338.
- 36. Ruotolo G, Parlavecchia M, Taskinen MR, Galimberti G, Zoppo A, et al. (1994) Normalization of lipoprotein composition by intraperitoneal insulin in IDDM. Role of increased hepatic lipase activity. Diabetes Care 17(1): 6-12.
- 37. Riemens SC, van Tol A, Sluiter WJ, Dullaart RP (1999) Plasma phospholipid transfer protein activity is lowered by 24-h insulin and acipimox administration: blunted response to insulin in type 2 diabetic patients. Diabetes 48(8): 1631-1637.
- 38. Arii K, Suehiro T, Yamamoto M, Ito H, Hashimoto K (1997) Suppression of plasma cholesteryl ester transfer protein activity in acute hyperinsulinemia and effect of plasma nonesterified fatty acid. Metabolism 46(10): 1166-1170.
- 39. Krauss RM (2004) Lipids and lipoproteins in patients with type 2 diabetes. Diabetes Care 27(6):1496-1504.

- 40. Carr MC, Brunzell JD (2004) Abdominal obesity and dyslipidemia in the metabolic syndrome: importance of type 2 diabetes and familial combined hyperlipidemia in coronary artery disease risk. J ClinEndocrinolMetab 89(6): 2601-2607.
- 41. Brunzell JD, Ayyobi AF (2003) Dyslipidemia in the metabolic syndrome and type 2 diabetes mellitus. Am J Med 115(suppl 8A):24S–28S.
- 42. Reaven GM (1987) Abnormal lipoprotein metabolism in non insulin dependent diabetes mellitus. Am J Med 83(suppl. 3A): 31-40.
- 43. Taskinen MR (1992) Quantitative and qualitative lipoprotein abnormalities in diabetes mellitus. Diabetes 41(2): 12-17.
- 44. Howard BV, Howard WJ (1994) Dyslipidemia in non insulin-dependent diabetes mellitus. Endocrine Rev15(3): 263-274.
- 45. Tomkin GH, Owens D (1994) Insulin and lipoprotein metabolism with special reference to the diabetic state. Diabetes Metab Rev10(3): 225-252.
- 46. Howard BV (1995) Pathogenesis of diabetic dyslipidemia. Diabetes Rev 3:423-32.
- 47. Lopes-Virella MF, Klein RL, Virella G (1996) Modification of lipoproteins in diabetes. Diabetes Metab Rev 12(1): 69-90.
- 48. Vergès B (1999) Dyslipidemia in diabetes mellitus. Review of the main lipoprotein abnormalities and their consequences on the development of atherogenesis. Diabetes Metab 25 (3):32-40.
- 49. Steiner G (1994) Thedyslipoproteinemias of diabetes. Atherosclerosis 110(suppl):27-33.
- 50. Steiner G, Tkac I, Uffelman KD, Lewis GF (1998) Important contribution of lipoprotein particle number to plasma triglyceride concentration in type 2 diabetes. Atherosclerosis 137(1): 211-214.
- 51. Kissebah AH, Alfarsi S, Evans DJ, Adams PW (1982) Integrated regulation of very low density lipoprotein triglyceride and apolipoprotein B kinetics in non insulin dependent diabetes mellitus. Diabetes 31(3): 217-225.
- 52. Reaven GM, Greenfield DS (1981) Diabetic hypertriglyceridemia. Evidence for three clinical syndromes. Diabetes 30 (2):66-75.

Tangvarasittichai S. Atherogenic Dyslipidemia: An Important Risk Factor for Cardiovascular Disease in Metabolic Syndrome and Type 2 Diabetes Mellitus Patients. Diabetes Obes Int J 2017, 2(1): 000144.

- 53. Tobey TA, Greenfield DS, Kramer F, Reaven GM (1981) Relationship between insulin resistance, insulin secretion, very low density lipoprotein kinetics and plasma triglyceride levels in normo triglyceridemic man. Metabolism 30(2): 165-171.
- 54. Malmström R, Packard LJ, Caslake M, Bedford D, Stewart P, et al. (1997) Defective regulation of triglyceride metabolism by insulin in the liver in NIDDM. Diabetologia 40(4): 454-462.
- 55. Lewis GF, Carpentier A, Adeli K, Giacca A (2002) Disordered fat storage and mobilization in the pathogenesis of insulin resistance and type 2 diabetes. Endocr Rev 23(2): 201-209.
- 56. Shimomura I, Matsuda M, Hammer RE, Bashmakov Y, Brown MS, et al. (2000) Decreased IRS-2 and increased SREBP-1c lead to mixed insulin resistance and sensitivity in livers of lipodystrophic and ob/ob mice. Mol Cell 6(1): 77-86.
- 57. Tobe K, Suzuki R, Aoyama M, Yamauchi T, Kamon J, et al. (2001) Increased expression of the sterol regulatory element-binding protein-1 gene in insulin receptor substrate-2(-/-) mouse liver. J Biol Chem 276: 38337-38340.
- Cummings MH, Watts GF, Umpleby AM, Hennessy TR, Kelly JM, et al. (1995) Acute hyperinsulinemia decreases the hepatic secretion of very-low-density lipoprotein apolipoprotein B-100 in NIDDM. Diabetes 44(9): 1059-1065.
- 59. Sparks JD, Sparks CE (1994) Insulin regulation of triacylglycerol-rich lipoprotein synthesis and secretion. BiochimBiophysActa 1215(1-2): 9-32.
- 60. Theriault A, Cheung R, Adeli K (1992) Expression of apolipoprotein B *in vitro* in cell-free lysates of HepG2 cells: evidence that insulin modulates ApoB synthesis at the translational level. Clin Biochem 25(5): 321-323.
- 61. Chireac DV, Chireac LR, Corsetti JP, Cianci J, Sparks CE, et al. (2000) Glucose-stimulated insulin secretion supresses hepatic triglyceride-rich lipoprotein and apob production. Am J EndocrinolMetab 279(5): E1003-E1011.
- 62. Sparks JD, Phung TL, Bolognino M, Sparks CE (1996) Insulin-mediated inhibition of apolipoprotein B secretion requires an intracellular trafficking event and phosphatidylinositol 3-kinase activation: studies

with brefeldin A and wortmannin in primary cultures of rat hepatocytes. Biochem J 313(pt2): 567-574.

- 63. Phung TL, Roncone A, Jensen KL, Sparks CE, Sparks JD (1997) Phosphoinositide 3-kinase activity is necessary for insulin-dependent inhibition of apolipoprotein B secretion by rat hepatocytes and localizes to the endoplasmic reticulum. J Biol Chem 272: 30693-30702.
- 64. Lin MC, Gordon D, Wetterau JR (1995) Microsomal triglyceride transfer protein (MTP) regulation in HEPG2 cells: insulin negatively regulates MTP gene expression. J Lipid Res 36(5): 1073-1081.
- 65. Taghibiglou C, Carpentier A, Van Iderstine SC, Chen B, Rudy D,et al. (2000) Mechanisms of hepatic very low density lipoprotein overproduction in insulin resistance. Evidence for enhanced lipoprotein assembly, reduced intracellular ApoB degradation, and increased microsomal triglyceride transfer protein in a fructose-fed hamster model. J Biol Chem 275(12): 8416-8425.
- 66. Bartels ED, Lauritsen M, Nielsen LB (2002) Hepatic expression of microsomal triglyceride transfer protein and *in vivo* secretion of triglyceriderich lipoproteins are increased in obese diabetic mice. Diabetes 51(4): 1233-1239.
- 67. Pont F, Duvillard L, Florentin E, Gambert P, Vergès B (2002) Early kinetic abnormalities of apoBcontaining lipoproteins in insulin-resistant women with abdominal obesity. Arterioscler Thromb Vasc Biol 22: 1726-1732.
- 68. Taskinen MR, Beltz WF, Harper I, Fields RM, Schonfeld G, et al. (1986) The effects of noninsulin dependent diabetes mellitus on VLDL triglyceride and VLDL apoB metabolism : studies before and after sulfonlyurea therapy. Diabetes 35(11): 1268-1277.
- 69. Duvillard L, Pont F, Florentin E, Galland-Jos C, Gambert P, Vergès B (2000) Metabolic abnormalities of apolipoprotein B-containing lipoproteins in noninsulin-dependent diabetes: a stable isotope kinetic study. Eur J Clin Invest 30(8): 685-694.
- 70. Taskinen MR, Nikkila EA, Kuusi T, Harno K (1982) Lipoprotein lipase activity and serum lipoproteins in untreated type 2 diabetes associated with obesity. Diabetologia 22(1): 46-50.

Tangvarasittichai S. Atherogenic Dyslipidemia: An Important Risk Factor for Cardiovascular Disease in Metabolic Syndrome and Type 2 Diabetes Mellitus Patients. Diabetes Obes Int J 2017, 2(1): 000144.

- 71. Panarotto D, Remillard P, Bouffard L, Maheux P (2002) Insulin resistance affects the regulation of lipoprotein lipase in the postprandial period and in an adipose tissue-specific manner. Eur J Clin Invest 32(2): 84-92.
- 72. Gianturco SH, Bradley WA, Gotto AM Jr, Morrisett JD, Peavy DL (1982) Hypertriglyceridemic very low density lipoproteins induce triglyceride synthesis and accumulation in mouse peritoneal macrophages. J Clin Invest 70(1): 168-178.
- Taskinen MR (2003) Diabetic dyslipidaemia: from basic research to clinical practice. Diabetologia 46(6): 733-749.
- 74. Vakkilainen J, Mero N, Schweizer A, Foley JE, Taskinen MR (2002) Effects of nateglinide and glibenclamide on postprandial lipid and glucose metabolism in type 2 diabetes. Diabetes Metab Res Rev 18(6): 484-490.
- 75. Mamo JCL, Szeto L, Steiner G (1990) Glycation of very low density lipoprotein from rat plasma impairs its catabolism. Diabetologia 33(6): 339-345.
- 76. Duvillard L, Pont F, Florentin E, Gambert P, Vergès B (2000) Significant improvement of apolipoprotein Bcontaining lipoprotein metabolism by insulin treatment in patients with non-insulin-dependent diabetes mellitus. Diabetologia 43(1): 27-35.
- 77. Duvillard L, Florentin E, Lizard G, Petit JM, Galland F, et al. (2003) Cell surface expression of LDL receptor is decreased in type 2 diabetic patients and is normalized by insulin therapy. Diabetes Care 26(5): 1540-1544.
- 78. Witzum JL, Mahoney EM, Bronks MJ, Fisher M, Elam R, et al. (1982) Non enzymatic glucosylation of low density lipoprotein alters its biologic activity. Diabetes 31(4pt1): 283-291.
- 79. Feingold KR, Grunfeld C, Pang M, Doerrler W, Krauss RM (1992) LDL subclass phenotypes and triglyceride metabolism in non insulin dependent diabetes. Arterioscler Thromb 12(12): 1496-1502.
- Haffner SM, Mykkanen L, Stern MP, Paidi M, Howard BV (1994) Greater effect of diabetes on LDL size in women than in man. Diabetes Care 17(10): 1164-1171.
- 81. Lamdenpera S, Sane T, Vuorinen-Markkola H, Knudsen P, Taskinen MR (1995) LDL particle sizes in midly hypertriglyceridemic subjects : no relation to

insulin-resistance or diabetes. Atherosclerosis 113: 227-236.

- 82. Austin MA, King MC, Vranizan KM, Krauss RM (1990) Atherogenic lipoprotein phenotype: a proposed genetic marker for coronary heart disease risk. Circulation 82(2): 495-506.
- 83. Lamarche B, Tchernof A, Moorjani S, Cantin B, Dagenais GR, et al. (1997) Small, dense low-density lipoprotein particles as a predictor of the risk of ischemic heart disease in men. Prospective results from the Quebec Cardiovascular Study. Circulation 95(1): 69-75.
- 84. Gardner CD, Fortmann SP, Krauss RM (1996) Association of small lowdensity lipoprotein particles with the incidence of coronary artery disease in men and women. JAMA 276(11): 875-881.
- 85. Chapman MJ, Guerin M, Bruckert E (1998) Atherogenic, dense low-density lipoproteins. Pathophysiology and new therapeutic approaches. Eur Heart J 19 (A): A24-30.
- 86. Anber V, Griffin BA, McConnell M, Packard CJ, Shepherd J (1996) Influence of plasma lipid and LDLsubfraction profile on the interaction between low density lipoprotein with human arterial wall proteoglycans. Atherosclerosis 124(2): 261-271.
- 87. Vakkilainen J, Makimattila S, Seppala-Lindroos A, Vehkavaara S, Lahdenpera S, et al. (2000) Endothelial dysfunction in men with small LDL particles. Circulation 102(7): 716-721.
- 88. Makimattila S, Liu ML, Vakkilainen J, Schlenzka A, Lahdenperä S, et al. (1999) Impaired endothelium dependent vasodilation in type 2 diabetes. Relation to LDL size, oxidized LDL, and antioxidants. Diabetes Care 22(6): 973-981.
- 89. Tribble DL, Holl LG, Wood PD, Krauss RM (1992) Variations in oxidative susceptibility among six low density lipoprotein subfractions of differing density and particle size. Atherosclerosis 93(3): 189-199.
- 90. Cominacini L, Garbin U, Pastorino AM, FrattaPasini A, Campagnola M, et al. (1994) Increased susceptibility of LDL to *in vitro* oxidation in patients with insulin-dependent and non-insulin-dependent diabetes mellitus. Diabetes Res 26(4): 173-184.
- 91. Vergès B, Galland F, Duvillard L (2003) Direct measurement of plasma oxidized LDL levels in type 2

Tangvarasittichai S. Atherogenic Dyslipidemia: An Important Risk Factor for Cardiovascular Disease in Metabolic Syndrome and Type 2 Diabetes Mellitus Patients. Diabetes Obes Int J 2017, 2(1): 000144.

diabetic patients before and after insulin therapy. Diabetologia 46 (2):73.

- 92. DeGraaf J, Hak-Lemmers HLM, Hectors MPC, Demacker PNM, Hendricks JCM, et al. (1991) Enhanced susceptibility to *in vitro* oxidation of the dense low density lipoprotein subfraction in healthy subjects. Arterioscler Thromb 11(2): 298-306.
- Kim HJ, Kurup IV (1982) Non enzymatic glycosylation of human plusma low density lipoprotein. Evidence for *in vitro* and *in vivo* glucosylation. Metabolism 31(4): 348-353.
- 94. Lyons TJ, Jenkins AJ (1997) Lipoprotein glycation and its metabolic consequences. Curr Opin Lipidol 8: 174-180.
- 95. Steinbrecher UP, Witzum JL (1984) Glucosylation of low density lipoproteins to an extent comparable to that seen in diabetes slows their catabolism. Diabetes 33: 130-134.
- 96. Hunt JV, Smith CCT, Wolff SP (1990) Autointoxidative glycosylation and possible involvement of peroxides and free radicals in LDL modification by glucose. Diabetes 39(11): 1420-1424.
- 97. Bowie A, Owens D, Collins P, Johnson A, Tomkin GH (1993) Glycosylated low density lipoprotein is more sensitive to oxidation: implications for the diabetic patient? Atherosclerosis 102(1): 63-67.
- 98. Verges B, Brun JM, Vaillant G, Quantin C, Brunet-Lecomte P, et al. (1992) Influence of obesity and hypertriglyceridemia on low HDL2-cholesterol level and its relationship with prevalence of atherosclerosis in type 2 diabetes. Diabetes Metab 18(4): 289-297.
- 99. Taskinen MR, Harno K, Nikkila EA (1984) Serum lipids and lipoproteins in type 2 diabetes. ActaEndocrinol (Kbh) 262(suppl):9 5-99.
- 100. Golay A, Zech L, Shi MZ, Chiou YA, Reaven GM, et al. (1987) High density lipoprotein (HDL) metabolism in non insulin dependent diabetes mellitus: measurement of HDL turn-over using tritiated HDL. J Clin Endocrinol Metab 65(3): 512-518.
- 101. Duvillard L, Pont F, Florentin E, Gambert P, Verges B (2000) Inefficiency of insulin therapy to correct apolipoprotein A-I metabolic abnormalities in

non insulin-dependent diabetes mellitus. Atherosclerosis 152(1): 229-237.

- 102. Nikkila EA, Huttunen JK, Ehnholm C (1977) Postheparin plasma lipoprotein lipase and hepatic lipase in diabetes mellitus. Relationship to plasma triglyceride metabolism. Diabetes 26(1): 11-21.
- 103. Verges B, Duvillard L, Pont F, Florentin E, Gambert P (2000) Respective effects of insulin resistance and "relative" insulin deficiency on apoB and apoA1 metabolism in NIDDM. Diabetologia 43(S1): A38.
- 104. Pont F, Duvillard L, Florentin E, Gambert P, Verges B (2002) High-density lipoprotein apolipoprotein A-I kinetics in obese insulin resistant patients. An *in vivo* stable isotope study. Int J Obes Relat Metab Disord 26(9): 1151-1158.
- 105. Caste C, Kuiper S, Blake W, Paigen B, Marotti K, et al. (1998) Remodeling of the HDL in NIDDM a fundamental role for cholesteryl ester transfer protein. Am J Physiol 274(6): E1091-E1098.
- 106. Calvo C, Talussot C, Ponsin G, Berthézène F (1988) Non enzymatic glycation of apolipoprotein A1. Effects on its self-association and lipid binding properties. Biochem Biophys Res Com 153(3): 1060-1067.
- 107. Duell PB, Oram JF, Bierman EL (1991) Nonenzymatic glycosylation of HDL and impaired HDL-receptor-mediated cholesterol efflux. Diabetes 40(3): 377-384.
- 108. Cavallero E, Brites F, Delfly B, Nicolaïew N, Decossin C, et al. (1995) Abnormal reverse cholesterol transport in controlled type II diabetic patients. Studies on fasting and postprandial LpA-I particles. Arterioscler Thromb Vasc Biol 15(12): 2130-2135.
- 109. Quintao EC, Medina WL, Passarelli M (2000) Reverse cholesterol transport in diabetes mellitus. Diabetes Metab Res Rev 16(4): 237-250.
- 110. Syvänne M, Castro G, Dengremont C, De Geitere C, Jauhiainen M, et al. (1996) Cholesterol efflux from Fu5AH hepatoma cells induced by plasma of subjects with or without coronary artery disease and non-insulin-dependent diabetes: importance of LpA-I:A-II particles and phospholipid transfer protein. Atherosclerosis 127(2): 245-253.

Tangvarasittichai S. Atherogenic Dyslipidemia: An Important Risk Factor for Cardiovascular Disease in Metabolic Syndrome and Type 2 Diabetes Mellitus Patients. Diabetes Obes Int J 2017, 2(1): 000144.

- 111. Bagdade JD, Lane JT, Subbaiah PV, Otto ME, Ritter MC (1993) Accelerated cholesteryl ester transfer in non insulin-dependent diabetes mellitus. Atherosclerosis 104(1-2): 69-77.
- 112. Riemens S, Vantoll A, Sluiter W, Dullaart R (1998) Elevated plasma cholesteryl ester transfer in NIDDM relationships with apo B-containing lipoproteins and phospholipid transfer protein. Atherosclerosis 140(1): 71-79.
- 113. Passarelli M, Catanozi S, Nakandakare ER, Rocha JC, Morton RE, et al. (1997) Plasma lipoproteins from patients with poorly controlled diabetes mellitus and *"in vitro"* glycation of lipoproteins enhance the transfer rate of cholesteryl ester from HDL to apo-B-containing lipoproteins. Diabetologia 40(9): 1085-1093.
- 114. Desrumaux C, Athias A, Bessede G, Vergès B, Farnier M, et al. (1999) Mass concentration of plasma transfer phospholipid protein in normolipidemic, type Ilahyperlipidemic, type IIbhyperlipidemic, and non-insulin-dependent diabetic subjects as measured by a specific ELISA. Arterioscler Thromb Vasc Biol 19(2): 266-275.
- 115. Samaras K, Botelho NK, Chisholm DJ, Lord RV (2010) Subcutaneous and visceral adipose tissue gene expression of serum adipokines that predict type 2 diabetes. Obesity (Silver Spring) 18(5): 884-889.
- 116. Fried SK, Bunkin DA, Greenberg AS (1998) Omental and subcutaneous adipose tis-sues of obese subjects release interleukin-6: depot difference and regulation by glucocorticoid. J Clin Endocrinol Metab 83(3): 847-850.
- 117. Chatterjee TK, Stoll LL, Denning GM, Harrelson A, Blomkalns AL, et al. (2009) Proinflammatory phenotype of perivascular adipocytes: influence of high-fat feeding. Circ Res 104(4): 541-549.
- 118. Wu D, Ren Z, Pae M, Guo W, Cui X, et al. (2007) Aging up-regulates expression of inflammatory mediators in mouse adipose tissue. J Immunol 179(7): 4829-4839.
- 119. Starr ME, Evers BM, Saito H (2009) Ageassociated increase in cytokine production during systemic inflammation: adipose tissue as a major source of IL-6. J Gerontol A Biol Sci Med Sci 64(7): 723–730.

- 120. Ouchi N, Kihara S, Funahashi T, Matsuzawa Y, Walsh K (2003) Obesity, adiponectin and vascular inflammatory disease. Curr Opin Lipidol 14(6): 561-566.
- 121. Berg AH, Scherer PE (2005) Adipose tissue, inflammation, and cardiovascular disease. Circ Res 96(9): 939-949.
- 122. Ouchi N, Higuchi A, Ohashi K, Oshima Y, Gokce N, et al. (2010) Sfrp5 is an anti-inflammatory adipokine that modulates metabolic dysfunction in obesity. Science 329 (5990): 454–457.
- 123. Grunfeld C, Zhao C, Fuller J, Pollack A, Moser A, et al. (1996) Endotoxin and cytokines induce expression of leptin, the ob gene product, in hamsters. J Clin Invest 97(9): 2152–2157.
- 124. Lord GM, Matarese G, Howard JK, Baker RJ, Bloom SR, et al. (1998) Leptin modulates the T-cell immune response and reverses starvation-induced immunosuppression. Nature 394(6696): 897-901.
- 125. Santos-Alvarez J, Goberna R, Sanchez-Margalet V (1999) Human leptin stimulates proliferation and activation of human circulating monocytes. Cell Immunol 194(1): 6-11.
- 126. Kiguchi N, Maeda T, Kobayashi Y, Fukazawa Y, Kishioka S (2009) Leptin enhances CC-chemokine ligand expression in cultured murine macrophage. Biochem Biophys Res Commun 384(3): 311-315.
- 127. Zarkesh-Esfahani H, Pockley AG, Wu Z, Hellewell PG, Weetman AP, et al. (2004) Leptin indirectly activates human neutrophils via induction of TNF-alpha. J Immunol 172(3): 1809-1814.
- 128. Faggioni R, Jones-Carson J, Reed DA, Dinarello CA, Feingold KR, et al. (2000) Leptin-deficient (ob/ob) mice are protected from T cell-mediated hepatotoxicity: role of tumor necrosis factor alpha and IL-18. Proc Natl Acad Sci USA 97(5): 2367-2372.
- 129. Matarese G, Di Giacomo A, Sanna V, Lord GM, Howard JK, et al. (2001) Requirement for leptin in the induction and progression of autoimmune encephalomyelitis. J Immunol 166(10): 5909-5916.
- 130. Fujimaki S, Kanda T, Fujita K, Tamura J, Kobayashi I (2001) The significance of measuring plasma leptin in acute myocardial infarction. J Int Med Res 29(2): 108-113.

Tangvarasittichai S. Atherogenic Dyslipidemia: An Important Risk Factor for Cardiovascular Disease in Metabolic Syndrome and Type 2 Diabetes Mellitus Patients. Diabetes Obes Int J 2017, 2(1): 000144.

- 131. Schulze PC, Biolo A, Gopal D, Shahzad K, Balog J, et al. (2011) Dynamics in insulin resistance and plasma levels of adipokines in patients with acute decompensated and chronic stable heart failure. J Card Fail 17(12): 1004-1011.
- 132. Faraj M, Lu HL, Cianflone K (2004) Diabetes, lipids, and adipocyte secretagogues. Biochem Cell Biol 82(1): 170-190.
- 133. Motojima K, Passilly P, Peters JM, Gonzalez FJ, Latruffe N (1998) Expression of putative fatty acid transporter genes are regulated by peroxisome proliferator-activated receptor alpha and gamma activators in a tissue- and inducer-specific manner. J Biol Chem 273(27): 16710-16714.
- 134. Yamauchi T, Kamon J, Waki H, Terauchi Y, Kubota N, et al. (2001) The fat-derived hormone adiponectin reverses insulin resistance associated with both lipoatrophy and obesity. Nat Med 7(8): 941-946.
- 135. Arita Y, Kihara S, Ouchi N, Takahashi M, Maeda K, et al. (1999) Paradoxical decrease of an adipose specific protein, adiponectin, in obesity. Biochem Biophys Res Commun 257(1): 79-83.
- 136. Weyer C, Funahashi T, Tanaka S, Hotta K, Matsuzawa Y, et al. (2001) Hypo-adiponectinemia in obesity and type 2 diabetes: close association with insulin resistance and hyperinsulinemia. J Clin Endocrinol Metab 86(5): 1930-1935.
- 137. Schulze MB, Rimm EB, Shai I, Rifai N, Hu FB (2004) Relationship between adiponectin and glycemic control, blood lipids, and inflammatory markers in men with type 2 diabetes. Diabetes Care 27(7): 1680-1687.
- 138. Cnop M, Havel PJ, Utzschneider KM, Carr DB, Sinha MK, et al. (2003) Relationship of adiponectin to body fat distribution, insulin sensitivity and plasma lipoproteins: evidence for independent role of age and sex. Diabetologia 46(4): 459-469.
- 139. Tschritter O, Fritsche A, Thamer C, Haap M, Shirkavand F, et al. (2003) Plasma adiponectin concentrations predict insulin sensitivity of both glucose and lipid metabolism. Diabetes 52(2): 239-243.

- 140. Shetty G, Economides P, Horton E, Mantzoros C, Veves A (2004) Circulating adiponectin and resistin levels in relation to metabolic factors, inflammatory markers and vascular reactivity in diabetic patients and subjects at risk for diabetes. Diabetes Care 27(10): 2450-2457.
- 141. Ng T, Watts G, Farvid M, Chan D, Barrett PH (2005) Adipocytokines and VLDL metabolism. Independent regulatory effects of adiponectin, insulin resistance and fat compartments on VLDL apoB-100 kinetics? Diabetes 54(3): 795-802.
- 142. Yamauchi T, Kamon J, Minokoshi Y, Ito Y, Waki H, et al. (2002) Adiponectin stimulates glucose utilization and fatty acid oxidation by activating AMP-activated protein kinase. Nat Med 8(11): 1288-1295.
- 143. Combs TP, Pajvani UB, Berg AH, Lin Y, Jelicks LA, et al. (2004) A transgenic mouse with a deletion in the collagenous domain of adiponectin displays elevated circulating adiponectin and improved insulin sensitivity. Endocrinology 145(1): 367-383.
- 144. Ruan H, Lodish HF (2003) Insulin resistance in adipose tissue: direct and indirect effects of tumor necrosis factor-alpha. Cytokine Growth Factor Rev 14(5): 447-455.
- 145. Sethi JK, Hotamisligil GS (1999) The role of TNF alpha in adipocyte metabolism. Semin Cell DevBiol 10(1): 19-29.
- 146. Chan DC, Watts GF, Ng TW, Uchida Y, Sakai N, et al. (2005) Adiponectin and other adipocytokines as predictors of markers of triglyceride-rich lipoprtein metabolism. Clin Chem 51(3): 578-585.
- 147. SörhedeWinzell M, Magnusson C, Ahrén B (2005) Theapj receptor is expressed in pancreatic islets and its ligand, apelin, inhibits insulin secretion in mice. Regul Pept 131(1-3): 12–17.
- 148. Guo L, Li Q, Wang W, Yu P, Pan H, et al. (2009) Apelin inhibits insulin secretion in pancreatic b-cells by activation of PI3-kinase-phosphodiesterase 3B. Endocr Res 34(4): 142–154.
- 149. Ringström C, Nitert MD, Bennet H, Fex M, Valet P, et al. (2010) Apelin is a novel islet peptide. Regul Pept 162(1-3): 44–51.

- 150. Ouchi N, Higuchi A, Ohashi K, Oshima Y, Gokce N, et al. (2010) Sfrp5 is an anti-inflammatory adipokine that modulates metabolic dysfunction in obesity. Science 329(5990): 454-457.
- 151. Sabio G, Das M, Mora A, Zhang Z, Jun JY, et al. (2008) A stress signaling pathway in adipose tissue regulates hepatic insulin resistance. Science 322(5907): 1539-1543.
- 152. Solinas G, Vilcu C, Neels JG, Bandyopadhyay GK, Luo JL, et al. (2007) JNK1in hematopoietically derived cells contributes to diet-induced inflammation and insulin resistance without affecting obesity. Cell Metab 6(5): 386-397.
- 153. Vallerie SN, Furuhashi M, Fucho R, Hotamisligil GS (2008) A predominant role for parenchymal c-Jun amino terminal kinase (JNK) in the regulation of systemic insulin sensitivity. PLoS One 3(9): e3151.
- 154. Kim J, Kim J, Kim DW, Ha Y, Ihm MH, et al. (2010) Wnt5a induces endothelial inflammation via beta-catenin-independent signaling. J Immunol 185(2): 1274-1282.
- 155. Christman 2nd MA, Goetz DJ, Dickerson E, McCall KD, Lewis CJ, et al. (2008) Wnt5a is expressed in murine and human atherosclerotic lesions. Am J Physiol Heart Circ Physiol 294(6): H2864-H2870.
- 156. Sheu WH, Shieh SM, Fuh MM, Shen DD, Jeng CY, et al. (1993) Insulin resistance, glucose intolerance, and hyperinsulinemia. Hypertriglyceridemia versus hypercholesterolemia. Arterioscler Thromb Vasc Biol 13(3): 367-370.

- 157. Tangvarasittichai S, Poonsub P, Tangvarasittichai O (2010) Association of serum lipoprotein ratios with insulin resistance in type 2 diabetes mellitus. Indian J Med Res 131: 641-648.
- Adiels M, Olofsson SO, Taskinen MR, Borén J (2006) Diabetic dyslipidaemia. Curr Opin Lipidol 17(3): 238-246.
- 159. Laws A, Reaven GM (1992) Evidence for an independent relationship between insulin resistance and fasting plasma HDL-cholesterol, triglyceride and insulin concentrations. J Intern Med 231(1): 25-30.
- 160. Stampfer MJ, Sacks FM, Salvini S, Willett WC, Hennekens CH (1991) A prospective study of cholesterol, apolipoproteins, and the risk of myocardial infarction. N Engl J Med 325(6): 373-381.
- 161. Jeppesen J, Facchini FS, Reaven GM (1998) Individuals with high total cholesterol/HDL cholesterol ratios are insulin resistant. J Intern Med 243(4): 293-298.
- 162. Li C, Ford ES, Meng YX, Mokdad AH, Reaven GM (2008) Does the association of the triglyceride to high-density lipoprotein cholesterol ratio with fasting serum insulin differ by race/ethnicity? Cardiovasc Diabetol 7: 4.
- 163. McLaughlin T, Reaven G, Abbasi F, Lamendola C, Saad M, et al. (2005) Is there a simple way to identify insulin-resistant individuals at increased risk of cardiovascular disease? Am J Cardiol 96(3): 399-404.



Tangvarasittichai S. Atherogenic Dyslipidemia: An Important Risk Factor for Cardiovascular Disease in Metabolic Syndrome and Type 2 Diabetes Mellitus Patients. Diabetes Obes Int J 2017, 2(1): 000144.