



# Antioxidant and Angiotensin-Converting Enzyme (ACE) Inhibitory Activity of Cereal Grain

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## Research Article

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## Abstract

Phenolic compounds are important products from secondary metabolism in plants. These cannot be synthesized in the human body and are mainly taken from plant source. Cereal grains are important sources of dietary polyphenols. Phenolic compounds are the main source of antioxidant that reduces the incidence of chronic diseases including heart disease, blood cholesterol, blood pressure, diabetic etc. Angiotensin-converting enzyme (ACE) inhibitors are medications that help to relax the veins and arteries to lower blood pressure. The aim of this experiment was to study the phenolic compounds identified in eight cereal grain (white rice, viscosity rice, brown rice, black rice, sticky rice, oatmeal, millet and sorghum) with their antioxidant and angiotensin-converting enzyme (ACE) inhibitory activity level. The phenolic content determined according to the Folin Ciocalteu's method. Black rice contains maximum phenolic compound (94.90 mg Tannic acid equivalent/g of extract) that was statistically different from other cereal grains ( $p < 0.05$ ) but followed by oatmeal and sorghum (58.82 and 58.57 mg Tannic acid equivalent/g of extract respectively). Antioxidant activities were comparatively assessed by ABTS (2, 2'-azino-bis-3-ethylbenzthiazoline-6-sulphonic acid) free radical decolorization assay method. It was observed that maximum cereal grain has no antioxidant activity. Among eight cereal grain, only black rice, oatmeal and sorghum showed considerable free radical scavenging capacity by ABTS radical. Angiotensin-converting enzyme (ACE) inhibition rate was determined by using L-hippuryl-histidyl-leucine (HHL). Maximum angiotensin-converting enzyme (ACE) inhibitory activity was also found in black rice (66.80%) followed by oatmeal (48.62%) and sorghum (37.45%). It might be concluded that polyphenol content was closely related to antioxidant activity and angiotensin-converting enzyme (ACE) inhibitor activity. Black rice, oatmeal and sorghum can be used as functional food for their high phenolic content, antioxidant activity as well as angiotensin-converting enzyme (ACE) inhibitory activity against chronic diseases.

**Keywords:** Polyphenol; Antioxidant; Angiotensin-converting enzyme; Cereal grain

## Introduction

Cereals are the most important primary sources of human diet throughout the world [1,2] that possesses great nutritional and bioactive properties such as phenolic acids-antioxidants, carbohydrate, protein, free amino acid, fibers, vitamins and minerals. Some cereals have been used as staple food both directly for human consumption and indirectly via livestock feed since the beginning of civilization [3]. It is the major source of energy for the world population. Based on the World Health Organization report for 2012–2016 [4], consumption of cereal grains may decrease the risk of non-communicable diseases (type 2 diabetes, cardiovascular disease, hypertension, and obesity). Instead, they comprise most of the micronutrients, fiber, and phytochemicals of the grain that could significantly impact on the nutritional quality of human food if integrated in flours or used as food ingredients [5].

Now a days, there has been a renewed interest in polyphenols as “life span essentials” due to their role in maintaining body functions and health throughout the adult and later phases of life [6]. Polyphenols are a large and diverse class of compounds, many of which occur naturally in a range of food plants. Natural polyphenols are mostly found in plants which are a kind of compounds with phenolic hydroxyl structure widely existing in nature [7]. Polyphenols not only have a strong antioxidation characteristic [8] but also have anticancer [9], bacteriostatic [10], liver protecting [11], anti-infection [12], cholesterol lowering [13] and immunity enhancing [14] properties. Moreover, these can prevent various biological activities such as type 2 diabetes [15,16]. Although, these compounds are not well known for direct role in nutrition (non-nutrients), many of them have antioxidant [17], anti-mutagenic, anti-osteogenic, anti-carcinogenic and anti-inflammatory, antiviral and platelet aggregation inhibitory activity that might potentially be beneficial in preventing or minimizing the incidence of diseases [18].

Angiotensin converting enzyme (ACE) is a multifunctional enzyme present in the rennin-angiotensin system that elevates blood pressure by generating the vasoconstrictor, angiotensin II [19]. Inhibition of ACE activity leads to a decrease in the concentration of angiotensin II, which decreases the tension in blood vessels and consequently reduces blood pressure [20]. The influence of ACE on blood pressure has made it an ideal target, and various synthetic medications such as captopril, enalapril, and lisinopril that inhibit angiotensin converting enzyme (ACE) are widely prescribed in the treatment and prevention of cardiovascular disease. Although, these drugs are often accompanied by undesirable side effects, no such side effects have been

observed for ACE inhibitors derived from food peptides [21]. In this respect, the search for diet-related preventive agents for hypertension is obviously of interest within the scope of functional foods. Food-derived ACE inhibitory peptides are just the ideal candidates for such products; offering many advantages including safety of the natural product, low cost, and the additional nutritional benefits of the peptides as source of essential amino acids.

Most studies for deriving ACE-inhibitory peptides from enzymatic hydrolysis of food proteins have focused on milk proteins [21-23] and fish proteins [24-26] while a few have explored the potential of alternatives such as soybean, mushroom garlic and pea [27-30]. With the growing demand for natural health products that can help to maintain healthy blood pressure, there is a need to consider other possible sources that can tailor to the wide spectrum of consumers with different dietary restrictions. Plant seeds, especially cereals are one of the most important sources of proteins worldwide and cereal proteins have been found to be potential precursors of antihypertensive peptides [31]. Therefore, it is necessary to proceed more research on cereal grain to obtain proper functional foods with high content of bioactive compounds and ACE-inhibitory peptides with strong antioxidant activity.

## Materials and Methods

### Plant materials

Eight cereal grain samples were selected for the study namely white rice, viscosity rice, brown rice, black rice, sticky rice, Oatmeal, Millet and sorghum. All seed samples were collected from Republic of Korea seed market.

### Chemicals and other materials

Tannic acid, Folin & Ciocalteu's phenol, 2,2'-azinobis-(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS) were obtained from Sigma-Aldrich (Saint Louis, MO, USA). HHL (L-hippuryl-histidyl-leucine), HA (Hippuric acid), ACE enzyme extract and captopril were also purchased from Sigma-Aldrich (Saint Louis, MO, USA).

### Preparation of sample extraction

Dried cereal grain samples were ground with a laboratory grinder to make flour and sieved with a 100-mesh sieve. One gram of sample was extracted in 10 mL 70% ethanol by shaking for three hours (h) at 180 rpm, and then centrifuged 10 min on 3000 rpm. Extracted supernatant sample was prepared after passing through syringe-filter and preserved in refrigerator.

### Polyphenol content analysis

Total phenolic content (TPC) of cereal grain sample extract was determined following the reported method [32] with slight modification. Firstly, 100 µL of each sample was mixed with 100 µL of Folin-Denis reagent, after 3 minutes 1 mL of 0.7 M Sodium carbonate was added in the mixture and then incubated at room temperature for 1 h. Absorbance of sample mixture was measured at 750 nm. Total phenolic compounds in the cereal grain extracts were determined using an equation obtained from a standard curve of tannic acid (0–500 µg/mL,  $Y = 0.0028x - 0.0351$ ,  $R^2 = 0.9968$ ) where concentrations of tannic acid on the X-axis and their corresponding absorbance values on the Y-axis was given. The results are expressed as mg Tannic acid equivalents (TA eq) per g of extract of cereal grain sample. All determinations were carried out in triplicate.

### Determination of antioxidant activity using free radical-scavenging ability by the use of ABTS radical

Antioxidant activity was determined by the ABTS (2, 2'-azino-bis-3-ethylbenzthiazoline-6-sulphonic acid) free radical decolorization assay method developed by Re, et al. [33]. The ABTS positive (+) radical cation was progenerated by mixing 7 mM ABTS stock solution with 2.45 mM potassium persulfate (final concentration) and incubating for 12-16 h in the dark at room temperature until the reaction was complete and the absorbance was stable. The absorbance of the ABTS positive (+) solution was equilibrated to 0.70 ( $\pm 0.02$ ) by diluting with water at room temperature, then 100µl was mixed with 50 µl of the test sample and the absorbance was measured at 734 nm after 6 min. All experiments were repeated three times. The radical scavenging activities of cereal grain sample were calculated by the following equation:

$$ABTS \text{ radical scavenging activity (\%)} = \left( 1 - \frac{\text{sample absorbance}}{\text{Control absorbance}} \right) \times 100$$

Then, curves were constructed by plotting percentage of inhibition against concentration in µg/mL. The equation of this curve allowed to calculate the  $IC_{50}$  corresponding to the sample concentration that reduced the initial ABTS absorbance of 50 %. A smaller  $IC_{50}$  value corresponds to a higher antioxidant activity. All test analyses were realized in triplicate.

### In Vitro Colorimetric Assay of ACE

ACE activity was assayed following the method of Jimsheena and Lalitha [34] by monitoring the release of HA (Hippuric acid) from the hydrolysis of the substrate HHL

(L-hippuryl-histidyl-leucine). The assay mixture contained 0.125 mL of 0.05 M sodium borate buffer pH 8.2 containing 0.3 M NaCl, 0.05 mL of 5 mM HHL, and 0.025 mL of ACE enzyme extract. The reaction was arrested after incubation at 37°C for 30 min by the addition of 0.2 mL of 1 M HCl. After stopping the reaction, 0.4 mL of pyridine was added followed by 0.2 mL of BSC, and the solution was mixed by inversion for 1 min and cooled on ice. After developing yellow color, absorbance was measured at 410 nm. One unit of ACE activity is defined as the amount of enzyme that releases 1 µmol of HA per min at 37°C and pH 8.2. For control group captopril was used.

### Statistical Analysis

All assay data for continuous variables were conducted in triplicates. The values were expressed as the mean  $\pm$  standard deviation (SD) calculated using Microsoft Excel 2010 and Sigma plot 12.5. All assay data were subjected to one-way analysis of variance (ANOVA) using PROC GLM in SAS program [35]. Mean values were compared with Duncan's Multiple Range Test at 0.05 level of Type I error.

## Results and Discussion

### Polyphenol content of cereal grain

In humans, polyphenols in the diet can enhance the immune defense ability of the body, reduce the incidence of chronic diseases, and have significant effects such as anti-allergy, anti-arterial atherosclerosis, anti-inflammation, antioxidation, antibacterial, antithrombotic, and protecting heart and blood vessels [36]. The health benefits of polyphenols on the human body are mainly due to their oxidation resistance. Polyphenols in grains have a stronger antioxidant effect in the body through the synergistic effect of multiple bioactive compounds than the single active ingredient and can eliminate too many oxidations free radicals in the body as anti-oxidants or after the intestinal digestion. These are known to have antioxidant activity and it is likely that the activity of these extracts is due to this phenolic compound [37]. The result of phenolic content of cereal grain was presented in Table 1. The results showed that black rice contained maximum polyphenol content (94.90 mg/g extract) that was statistically different from other cereal grain ( $p < 0.05$ ). Oatmeal and sorghum contain 58.75 and 58.82 mg/g extract of polyphenol content that was statistically similar. While sticky rice contained minimum polyphenol content (0.11 mg/g extract) that was statistically different from other cereal grains. Similar result was described by Dykes and Rooney [38] where black rice had the height level of polyphenol content and antioxidant activity whereas non pigmented cereals i.e. white rice, wheat and waxy barley had the lowest levels. Pedro, et al. [39]

reported that black rice contains 52.02 mg/g of polyphenol. Bolea, et al. [40] observed that total polyphenol content of black rice is 48.3 mg/g fresh weight. The phenolic content values of red rice varieties reported by Sompong, et al. [41] were between 0.14 and 0.34 mg GA/g, whereas the black rice varieties displayed 4-fold higher values (0.74-0.105 mg GA/g). The analysis of polyphenols from black rice assessed by Murdifi, et al. [42] presented a value of  $11.97 \pm 9.00$  mg GAE/g. The deviation in results can be due to the difference in extraction technique, unit and assay method. Black rice has a higher content of phenolic compounds as compared to white rice and sticky rice [43]. Mira, et al. [44] found that total phenolic compound was four times higher in pigmented rice than in non-pigmented rice.

### Antioxidant activity of cereal grain

The concentration of total polyphenol content in cereal grain has been positively associated with the antioxidant

activity [45-47] with potential beneficial effects on health, such as reduction of oxidative stress [48,49], aid in the prevention of cancer [50,51], in the control of blood lipids and related diseases, which may help in the prevention of cardiovascular problems [48], and in the prevention of the complications of diabetes [43,52,53]. Nam, et al. [54] reported that cereal grains with red and black pericarp presented higher antioxidant activity than those with light brown pericarp color. ABTS radical is widely used to evaluate the free-radical scavenging capacity of antioxidants level according to their hydrogen donating ability [55]. In addition to that, reactions involved in these methods are fully unaffected by side reactions [56]. The antioxidant properties of extracts were measured in terms of their efficient  $IC_{50}$  concentration corresponding to the sample concentration that reduced the initial ABTS radical absorbance of 50%. These  $IC_{50}$  values for ABTS is given in Table 1.

Sample	Polyphenol content	Radical scavenging activity
	(mg/g extract)	( $RC_{50}$ )
		ABTs ( $\mu\text{g/mL}$ )
White Rice	$11.29 \pm 1.25^d$	ND
Viscosity Rice	$3.11 \pm 1.07^f$	ND
Brown Rice	$32.89 \pm 1.79^c$	ND
Black Rice	$94.90 \pm 1.16^a$	211.25
Sticky rice	$0.11 \pm 0.19^g$	ND
Oatmeal	$58.82 \pm 0.44^b$	282.67
Millet	$4.12 \pm 0.96^e$	ND
Sorghum	$58.57 \pm 1.60^b$	291.24

Each value represents the mean  $\pm$  SD (n = 3). Means having different letters are significantly different (DMRT,  $p < 0.05$ ).  $IC_{50}$ : Inhibition capacity for 50% reduction of ABTS radical. ND means not detected.

**Table 1:** Polyphenol content and radical scavenging activity of different cereal grain by using ABTS radical.

In this study, it was reported that all cereal grain did not show antioxidant activity except black rice, Oatmeal and sorghum. Maximum ABTS radical scavenging ability was shown in black rice that was 50% inhibition at 211.25  $\mu\text{g/ml}$  concentration. In same way, oatmeal and sorghum also showed 50% radical inhibition at 282.67 and 291.24  $\mu\text{g/ml}$  concentrations respectively. The antioxidant properties of black rice are more effective than red rice varieties [41]. Black rice and sorghum showed a stronger scavenging activity than other cereal grain. Black rice might be a potential material for antioxidants. Yildirim, et al. [57] reported that some potent antioxidants applied in food might cause serious side effects. Therefore, it is important to find more natural anti-oxidative ingredients for food industries. A study of Saenkod *et al.* [58] on Chinese black rice (Brown Himi variety) showed a

high antioxidant activity of 70.82%. Antioxidant activity of Korean black rice is 40.39 to 55.20% obtained by Park, et al. [59]. The genetic and environmental factors, the processing conditions [60,61], as well as the extraction procedures may all affect the antioxidant levels of cereal grain sample.

In recent years, pigmented rice has been classified as one of the most consumed products among functional foods, due to the fact that it contains a high amount of phenolic compounds that have antioxidant properties [62,63]. Besides, grains with darker pericarp colour, such as red and black rice, contain higher amounts of polyphenols [64]. The concentration of total phenolics in the grain has been positively associated with the antioxidant activity [46,47], with potential beneficial effects on health.

### ACE inhibition ability

Angiotensin converting enzyme (ACE) is a proteolytic enzyme that regulates blood pressure (BP) by its hydrolytic actions. ACE converts angiotensin-I (that has no direct effect on BP) to angiotensin-II, which constricts blood vessels (vasoconstrictor) and thereby elevates BP. Since ACE is an enzyme, it can be inactivated by blocking its active site with selective enzyme inhibitors. Synthetic ACE-inhibitors are widely used in the pharmacological treatment of hypertension. Certain peptides with suitable structures are also able to inhibit the activity of ACE by binding to its active site. In this experiment maximum ACE inhibition was observed in Black rice (66.80%) followed by oatmeal (48.62%) that was statistically different from other cereal grain. Minimum ACE inhibition ability was observed in millet (8.50%).

Sample	ACE inhibition Rate (%)
	(Conc. 1,000 µg/mL)
White Rice	32.31±1.15 <sup>d</sup>
Viscosity Rice	26.16±2.82 <sup>e</sup>
Brown Rice	32.30±1.63 <sup>d</sup>
Black Rice	66.80±2.45 <sup>a</sup>
Sticky rice	25.00±2.36 <sup>e</sup>
Oatmeal	48.62±1.97 <sup>b</sup>
Millet	8.50±2.04 <sup>f</sup>
Sorghum	37.45±2.71 <sup>c</sup>

Each value represents the mean ± SD (n = 3). Means having different letters are significantly different (DMRT,  $p < 0.05$ ).

**Table 2:** Angiotensin converting enzyme (ACE) inhibition ability of different cereal grain.

In a literature data base survey, twenty-two potential ACE-inhibitory peptides in cereal proteins have been reported [65]. In a recent study, *in silico* digestion of oat and barely by thermolysin resulted in generating 6 and 3 potent peptides from their parent proteins, respectively [66]. Furthermore, several cereal food-derived peptides have been shown to have *in vitro* ACE inhibitory activity that included corn, wheat, oat and rice [67-72].

### Conclusion

Black rice contained maximum polyphenol content (94.90 mg Tannic acid equivalent/g of extract) that are responsible for maximum antioxidant activity as well as maximum ACE inhibition ability in black rice. Maximum angiotensin converting enzyme (ACE) inhibitory activity was also shown in black rice 66.80% following by oatmeal

(48.62%) and sorghum (37.45%). It can be concluded that polyphenol content was closely related with antioxidant activity and angiotensin-converting enzyme (ACE) inhibitory activity. Black rice, oatmeal and sorghum can be used as functional ingredient with their high phenolic content, antioxidant activity as well as angiotensin-converting enzyme (ACE) inhibitory activity.

### Author Contribution

Soyema Khatun: Methodology, Investigation, Experiment conduction, writing original draft, Visualization.

Hyunhwa Lee : Help to analysis ACE by using colorimetric method.

Md. Mahi Imam Mollah: Data analysis, Review of Manuscript

### Conflict of Interest

The authors declare that they have no known competing interests or personal relationships that could have appeared to influence the work reported in this paper.

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### References

1. Liu RH (2007) Whole grain phytochemicals and health. *Journal of Cereal Science* 46(3): 207-219.
2. FAO (Food and Agriculture Organisation) (2002) *World Agriculture: Towards 2015/2030. Summary Report.* FAO, Rome.
3. BNF (British Nutrition Foundation) (1994) *Starchy Foods in the Diet.* BNF, London.
4. World Health Organization (WHO) report (2018) *Prevention and Control of Noncommunicable Diseases in the European Region: A Progress Report.*
5. Brouns F, Hemery Y, Price R, Anson NM (2012) Wheat aleurone: Separation, composition, health aspects and potential food use. *Crit Rev Food Sci Nutr* 52(6): 553-568.
6. Chandrasekara A, Shahidi F (2010) Content of insoluble bound phenolics in millets and their contribution to antioxidant capacity. *J Agric Food Chem* 58(11): 6706-6714.

7. Lewandowska U, Szewczyk K, Hrabec E, Janecka A, Gorchach S (2013) Overview of metabolism and bioavailability enhancement of polyphenols. *Journal of Agricultural and Food Chemistry* 61(50): 12183-12199.
8. Zhang R, Zeng Q, Deng Y, Zhang M, Wei Z, et al. (2013) Phenolic profiles and antioxidant activity of litchi pulp of different cultivars cultivated in Southern China. *Food Chemistry* 136(4): 1169-1176.
9. Noratto G, Porter W, Byrne D, Cisneros-Zevallos L (2009) Identifying peach and plum polyphenols with chemopreventive potential against estrogen-independent breast cancer cells. *Journal of Agricultural and Food Chemistry* 57(12): 5119-5126.
10. Ng KR, Lyu X, Mark R, Chen WN (2019) Antimicrobial and antioxidant activities of phenolic metabolites from flavonoid-producing yeast: potential as natural food preservatives. *Food Chemistry* 270: 123-129.
11. Callcott ET, Santhakumar AB, Luo J, Blanchard CL (2018) Therapeutic potential of rice-derived polyphenols on obesity-related oxidative stress and inflammation, *Journal of Applied Biomedicine* 16 (4): 255-262.
12. Pešić MB, Milinčić DD, Kostić AZ, Stanisavljević NS, Vukotić GN, et al. (2019) In vitro digestion of meat- and cereal-based food matrix enriched with grape extracts: how are polyphenol composition, bioaccessibility and antioxidant activity affected? *Food Chemistry* 284: 28-44.
13. Liu S, You L, Zhao Y, Chang X (2018) Wild *Lonicera caerulea* berry polyphenol extract reduces cholesterol accumulation and enhances antioxidant capacity *in vitro* and *in vivo*. *Food Research International* 107: 73-83.
14. Cuevas A, Saavedra N, Salazar LA, Abdalla DSP (2013) Modulation of immune function by polyphenols: possible contribution of epigenetic factors: possible contribution of epigenetic factors, *Nutrients* 5(7): 2314-2332.
15. Vitale M, Vaccaro O, Masulli M, Bonora E, Prato SD, et al. (2017) Polyphenol intake and cardiovascular risk factors in a population with type 2 diabetes: the TOSCA.IT study. *Clinical Nutrition* 36(6): 1686-1692.
16. Xiao J, Kai G, Yamamoto K, Chen X (2013) Advance in dietary polyphenols as  $\alpha$ -glucosidases inhibitors: a review on structure-activity relationship aspect. *Critical Reviews in Food Science and Nutrition* 53(8): 818-883.
17. Sripriya G, Chandrasekharan K, Murty VS, Chandra TS (1996) ESR spectroscopic studies on free radical quenching action of finger millet (*Eleusine coracana*). *Food Chem* 57(4): 537-540.
18. Ferguson LR (2001) Role of plant polyphenols in genomic stability. *Mutat Res* 475(2): 89-111.
19. Fujita H, Yokoyama K, Yoshikawa M (2000) Classification and antihypertensive activity of angiotensin I-converting enzyme inhibitory peptides derived from food proteins. *Journal of Food Science* 65(4): 564-569.
20. Ondetti MA, Rubin B, Cushman DW (1977) Design of specific inhibitors of angiotensin-converting enzyme: new class of orally active antihypertensive agents. *Science* 196(4288): 441-444.
21. Hata Y, Yamamoto M, Ohni M, Nakajima K, Nakamura Y, et al. (1996) A placebo-controlled study of the effect of sour milk on blood pressure in hypertensive subjects. *Am J Clin Nutr*. 64(5): 767-771.
22. Phelan M, Kerins D (2011) The potential role of milk-derived peptides in cardiovascular disease. *Food Funct* 2: 153-167.
23. Mao XY, Ni JR, Sun WL, Hao PP, Fan L (2007) Value-added utilization of yak milk casein for the production of angiotensin-I-converting enzyme inhibitory peptides. *Food Chemistry* 103(4): 1282-1287.
24. Raghavan S, Kristinsson HG (2009) ACE-inhibitory activity of tilapia protein hydrolysates. *Food Chemistry* 117(4): 582-588.
25. Ono S, Hosokawa M, Miyashita K, Takahashi K (2006) Inhibition properties of dipeptides from salmon muscle hydrolysate on angiotensin I-converting enzyme. *International Journal of Food Science and Technology* 41(4): 383-386.
26. Qian ZJ, Je JY, Kim SK (2007) Antihypertensive effect of angiotensin I-converting enzyme-inhibitory peptide from hydrolysates of Bigeye tuna dark muscle, *Thunnus obesus*. *J Agric Food Chem* 55(1): 8398-8403.
27. Rho SJ, Lee JS, Chung Y, Kim YW, Lee HG (2009) Purification and identification of an angiotensin I-converting enzyme inhibitory peptide from fermented soybean extract. *Process Biochemistry* 44(4): 490-493.
28. Hyoung LD, Ho KJ, Sik PJ, Jun CY, Soo LJ (2004) Isolation and characterization of a novel angiotensin I-converting enzyme inhibitory peptide derived from the edible mushroom *Tricholoma giganteum*. *Peptides* 25(4): 621-627.
29. Suetsuna K (1998) Isolation and characterization of angiotensin I-converting enzyme inhibitor dipeptides

- derived from *Allium sativum* L (garlic)," *Journal of Nutritional Biochemistry* 9(7): 415-419.
30. Li H, Prairie N, Udenigwe CC, Adebisi AP, Tappia PS, et al. (2011) Blood pressure lowering effect of a pea protein hydrolysate in hypertensive rats and humans. *J Agric Food Chem* 59(18): 9854-9860.
  31. Anantharaman K, Finot PA (1993) Nutritional Aspects of Food Proteins in Relation to Technology. *Food Reviews International* 9(4): 629- 655.
  32. Singleton VL, Orthofer R, Lamuela Raventós RM (1999) Analysis of total phenols and other oxidation substrates and antioxidants by means of folin ciocalteu reagent. *Methods in Enzymology* 299: 152-178.
  33. Re R, Pellegrini N, Proteggente A, Pannala A, Yang M, et al. (1999) Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Rad Biol Med* 26: 1231-1237.
  34. Jimsheena VK, Lalitha RG (2009) Colorimetric, high-throughput assay for screening Angiotensin I-Converting Enzyme Inhibitors. *Anal Chem* 81: 9388-9394.
  35. SAS Institute Inc (1989) SAS/STAT user's guide, Release 6.03, Ed. Cary, NC.
  36. Sae-Leaw T and Benjakul S (2019) Prevention of melanosis in crustaceans by plant polyphenols: a review. *Trends in Food Science & Technology* 85: 1-9.
  37. Tepe B, Sokmen M, Akpulat HA, Sokmen A (2006) Screening of the antioxidant potentials of six *Salvia* species from Turkey. *Food Chem* 95(2): 200-204.
  38. Dykes L, Rooney LW (2007) Phenolic compounds in cereal grains and their health benefits. *Cereal Food World* 52(3): 105-111.
  39. Pedro AC, Granato D, Rosso ND (2016) Extraction of anthocyanins and polyphenols from black rice (*Oryza sativa* L.) by modeling and assessing their reversibility and stability. *Food Chemistry* 191: 12-20.
  40. Bolea C, Vizireanu C (2017) Polyphenolic content and antioxidant properties of black rice flour. *AUDJG – Food Technology* 41(2): 75-85.
  41. Sompong R, Siebenhandl Ehn S, Linsberger Martin G, Berghofer E (2011) Physicochemical and antioxidative properties of red and black rice varieties from Thailand, China and Sri Lanka. *Food Chemistry* 124(1): 132-140.
  42. Murdifi M, Pakki E, Rahim A, Syaiful SA, Evary IYM, et al. (2015) Physicochemical properties of Indonesian pigmented rice (*Oryza sativa* Linn.) varieties from South Sulawesi. *Asian Journal of Plant Sciences* 14(2): 59-65.
  43. Yawadio R, Tanimori S, Morita N (2007) Identification of phenolic compounds isolated from pigmented rices and their aldose reductase inhibitory activities. *Food Chemistry* 101(4): 1616-1625.
  44. Mira NVM, Massaretto IL, Pascual CS CI, Marquez UML (2009) Comparative study of phenolic compounds in different Brazilian rice (*Oryza sativa* L.) genotypes. *Journal of Food Composition and Analysis* 22(5): 405-409.
  45. Itani T, Tatemoto H, Okamoto M, Fujii K, Muto N (2002) A comparative study on antioxidative activity and polyphenol content of colored kernel rice. *J Jpn Soc Food Sci* 49(8): 540-543.
  46. Goffman FD, Bergman CJ (2004) Rice kernel phenolic content and its relationship with antiradical efficiency. *J Sci Food Agr* 84(10): 1235-1240.
  47. Zhang M, Guo B, Zhang R, Chi J, We Z, et al. (2006) Separation, purification and identification of antioxidant compositions in black rice. *Agric Sci China* 5(6): 431-440.
  48. Ling WH, Cheng QX, Ma J, Wang T (2001) Red and black rice decrease atherosclerotic plaque formation and increase antioxidant status in rabbits. *J Nutr* 131(5): 1421-1426.
  49. Hu C, Zawistowski J, Ling W, Kitts DD (2003) Black rice (*Oryza sativa* L. indica) pigmented fraction suppresses both reactive oxygen species and nitric oxide in chemical and biological model systems. *J Agric Food Chem* 51(18): 5271-5277.
  50. Hudson EA, Dinh PA, Kokubun T, Simmonds MSJ, Gescher A (2000) Characterization of potentially chemopreventive phenols in extracts of brown rice that inhibit the growth of human breast and colon cancer cells. *Cancer Epidem Biomar* 9(11): 1163-1170.
  51. Hyun JW, Chung HS (2004) Cyanidin and malvidin from *Oryza sativa* cv. Heugjinjubyeo mediate cytotoxicity against human monocytic leukemia cells by arrest of G2/M phase and induction of apoptosis. *J Agric Food Chem* 52(8): 2213-2217.
  52. Hu FB (2003) Plant-based foods and prevention of cardiovascular disease: an overview. *Am J Clin Nutr* 78(3): 544S-551S.
  53. Orlich MJ, Fraser GE (2014) Vegetarian diets in the Adventist Health Study 2: a review of initial published

- findings. *Am J Clin Nutr.* 100(1): 353S–358S.
54. Nam SH, Choi SP, Kang MY, Kozukue N, Friedman M (2005) Antioxidative, antimutagenic, and anticarcinogenic activities of rice bran extracts in chemical and cell assays. *J Agric Food Chem* 53(3): 816-822.
  55. Nuutila AM, Puupponen Pimiä R, Aarni M, Kirsi Marja Oksman Caldentey (2003) Comparasion of antioxidant activities of onion and garlic extracts by inhibition of lipid peroxidation and radical scavenging activity, *Food Chem* 81(4): 485-493.
  56. Livani F, Ghorbanli M, Sateyi A (2013) Changes in antioxidant activity and control of phenolic compounds during the ripening process of elm-leaved blackberry fruit. *Int. J Agron Plant Prod* 4(1): 88-93.
  57. Yildirim A, Oktay M, Bilaloglu V (2001) The antioxidant activity of the leaves of *Cydonia vulgaris*. *Turk J Med Sci* 31: 23-27.
  58. Saenkod C, Liu Z, Huang J, Gong Y (2013) Anti-oxidative biochemical properties of extracts from some Chinese and Thai rice varieties. *African Journal of Food Science* 7(9): 300-305.
  59. Park SY, Kim SJ, Chang HI (2008) Isolation of anthocyanin from black rice (*Heuginjubyeo*) and screening of its antioxidant activities. *Korean Journal of Microbiology and Biotechnology* 36(1): 55-60.
  60. Mpofu A, Sapirstein HD, Beta T (2006) Genotype and environmental variation in phenolic content, phenolic acid composition, and antioxidant activity of hard spring wheat. *J Agric Food Chem* 54(4): 1265–1270.
  61. Mpofu A, Beta T, Sapirstein HD (2007) Effects of Genotype, Environment and Genotype Environment Interaction on the Antioxidant Properties of Wheat. In: Yu L (Eds.), *Wheat Antioxidants*, John Wiley and Sons Inc.: Hoboken, NJ, USA, pp: 24-41.
  62. Mira NVM, Massaretto IL, Pascual CS CI, Marquez UML (2009) Comparative study of phenolic compounds in different Brazilian rice (*Oryza sativa* L.) genotypes. *Journal of Food Composition and Analysis* 22(55): 405-409.
  63. Shen Y, Jin L, Xiao P, Lu Y, Bao J (2009) Total phenolics, flavonoids, antioxidant capacity in rice grain and their relations to grain color, size and weight. *Journal of Cereal Science* 49(1): 106-111.
  64. Zhou Z, Robard K, Helliwell S, Blanchard C (2004) The distribution of phenolic acids in rice. *Food Chemistry* 87(3): 401-406.
  65. Loponen J (2004) Angiotensin converting enzyme inhibitory peptides in Finnish cereals: A database survey. *Agricultural and Food Science* 13(2): 39-45.
  66. Gu Y, Majumder K, Wu J (2011) QSAR-aided in silico approach in evaluation of food proteins as precursors of ACE inhibitory peptides. *Food Research International* 44(8): 2465-2474.
  67. Yang Y, Tao G, Liu P, Liu J (2007) Peptide with angiotensin I-converting enzyme inhibitory activity from hydrolyzed corn gluten meal. *J Agric Food Chem.* 55(19): 7891-7895.
  68. Lu WL, Hu JH, Li FQ (2009) Research on the antihypertensive effect of corn angiotensin converting enzyme inhibitory peptides. *Pharmaceutical Care and Research* 9: 126-129.
  69. Thewissen BG, Pauly A, Celus I, Brijs K, Delcour JA (2011) Inhibition of angiotensin I-converting enzyme by wheat gliadin hydrolysates. *Food Chemistry* 127(4): 1653-1658.
  70. Jia J, Ma H, Zhao W, Wang Z, Tian W, et al. (2010) The use of ultrasound for enzymatic preparation of ACE-inhibitory peptides from wheat germ protein. *Food Chemistry* 119(1): 336-342.
  71. Ma H, Geng J, Luo L, Yang Q, Zhu W (2010) Effect of oat-seed protein pretreatment by ultrasonic on preparation of ACE inhibitory peptides. *Nongye Jixie Xuebao/ Transactions of the Chinese Society of Agricultural Machinery* 4: 133-137.
  72. He GQ, Xuan GD, Ruan H, Chen QH, Xu Y (2005) Optimization of angiotensin I-converting enzyme (ACE) inhibition by rice dregs hydrolysates using response surface methodology. *J Zhejiang Univ Sci B* 6(6): 508-513.

