



# Exploring Forensic Challenges and Insights: Implications of Genetic Analyses on Degraded DNA Samples

**Rathnayake RWRK\*, Marasinghe E, Perera LRT and Bandaranayake VJ**

Department of Government Analyst's, Senior Assistant Government Analyst, Srilanka

**\*Corresponding author:** Rathnayake Weerakoonge Ruchira Kalhari Rathnayake, Department of Government Analyst's, Senior Assistant Government Analyst, 31, Isuru Mawatha, Pelawatta, Battaramulla, Srilanka, Tel: +94112-786395; Fax: +94112-786394; Email: kalharigad@hotmail.com

## Case Report

Volume 9 Issue 1

Received Date: January 30, 2024

Published Date: February 14, 2024

DOI: 10.23880/ijfsc-16000359

## Abstract

Identifying missing persons from different types of body parts poses an intricate challenge for law enforcement and forensic experts. Identification through human remains is challenging when the available body parts don't contain distinctive features, commonly used for anthropological analysis. The complexities arise from the fragmentation of remains and difficulties in obtaining viable biological samples due to decomposition and exposure to other factors enhancing degradation. The absence of personal information and known reference samples for familial identification further amplifies the complexities in the process. When dealing with different types of human remains and lacking additional information of those missing individuals, the analysis centered on DNA as the most dependable and potentially definitive method for identification. In such scenario, it is imperative to select ideal tissue among different tissue types for DNA extraction followed by optimized DNA extraction protocols and STR amplification kits with different markers to achieve consistent DNA profiling in identification purposes.

**Keywords:** Decomposed Human Remains; Human Identification; DNA Extraction; STR Amplification

**Abbreviations:** DNA: Deoxyribo Nucleic Acid; STR: Short Tandem Repeats.

## Introduction

Identifying victims is a fundamental step in resolving a criminal case, and it becomes particularly challenging when the remains are badly decomposed or skeletonized. Utilizing multidisciplinary techniques, including anthropology, may not be as effective in identification because bone samples in human remains lack prominent morphological features [1,2]. Visual and traditional presumptive methods of identification

typically involve the examination of remains by relatives or associates of the missing person. However these methods should be exclusively relied upon for identification when the bodies are intact, without decomposition or mutilation. On the other hand the risk becomes more significant on these presumptive methods when the number of deceased individuals increased [3]. Criminals may engage in the mutilation and employ diverse methods of a dead body to remove any identifiable features.

This destructive behavior is intended to eliminate all traces that could be used for identification purpose [4].

All these common factors coupled with the absence of reference material, make difficulty for authorities to determine the identity of the deceased individual, creating challenges in the investigation process. As of our current knowledge, there are no standardized guidelines for such incident and analysis based on DNA as a primary identifier [5].

The current best practice following in the DNA laboratory in government Analyst's Department for human remains identification from damaged and degraded samples involves employing optimized methods for various tissue types. This involves using different extraction methods for different tissue types, expanded STR markers to enhance resolution and integrating lineage markers for specific applications. The varied methods for extraction and amplification provide more precise and effective approach in current investigations. However, we stay updated with the latest technologies and protocols to address challenges, especially in situations involving degraded human remains.

The article details three case studies in which human remains sent to the Government Analyst's Department

for identification of missing individuals. In all three cases, samples were retrieved from decomposed human remains by judicial medical officers and produced to Government Analyst's Department by police through court order. Relatives of missing persons were produced to the department in accordance with the court order to undergo blood collection.

### Case Study-1

Body parts, showing signs of damages and partial decomposition, were discovered in two distinct locations within proximity of nearly 1 kilometer, in Western Province in Sri Lanka, all within the duration of one week. Notably, there have been no reports of missing persons from public corresponding to these remains. In first location, a partially decomposed body lacking both head and limbs was found. Piece of bone sample and muscle sample was recovered. In the second location, partially decomposed lower limb was uncovered, and muscle sample was recovered.

Figure 1 and Figure 2 represents images of human remains discovered in two locations



Piece of bone



Sample of muscle

**Figure 1:** The Piece of Bone and Muscle Sample Recovered from Decomposed Body (Case Study-1, Location 1)

### Case Study-2

A man went missing in Colombo, Western Province in Sri Lanka. Mother is the sole close relative of the missing person and reported the disappearance to the police two days after the individual went missing. A person living close to the missing individual was arrested after about two years by the police following a comprehensive investigation. During questioning, the individual admitted of killing the missing person and burying the body in his garden. However, the person later removed the decayed skeletal remains from his garden and disposed them into nearby river, leaving only the skull with teeth accidentally at the burial site. Skull with teeth was produced for DNA identification. A blood sample from of missing person's mother was submitted for identification purposes. Figure 3 represents images of human remains discovered in the garden of suspect.



**Figure 2:** Sample of Muscle Recovered from Decomposed Lower Limb (Case Study-1-Location-2).



**Figure 3:** Skull with Teeth Found from Burial Site of Suspected Person's Garden (Case Study-2).

### Case Study-3

In a village near the jungle in North Province of Sri Lanka, a person was mysteriously disappeared, leaving no traces or clues. The only close relative, his father reported his son's disappearance to the police. After several months, someone hunting in a jungle near the village accidentally found sample of bones. Two long bones were recovered by police and produced to DNA analysis. Blood sample of missing person's father was produced for identification. Figure 4 represents images of human remains discovered in the jungle.



**Figure 4:** Two Bone Samples Found from Jungle (Case Study-3).

### Materials and Methods

#### Sample Information

To eliminate potential contaminants and dirt, bone samples were subjected to surface scraping using a variable speed dental milling drill (Dermal, Racine, WI, USA). Following this, the samples underwent two-step cleaning process, washed twice with 10% sodium hypochlorite followed by two rinses with sterile water for 5 minutes each and allowed to air-dry overnight. Similar cleaning procedure followed for teeth recovered from the skull by surface scraping using a scalpel blade and cleaned with a bleach solution, followed by rinsing with sterile water and left to air-dry. Dried bone and teeth were placed in plastic containers and immersed with 0.5M EDTA pH 8.0. The containers were incubated on a shaking rotator at 56°C for duration of two weeks. After first week, the EDTA solution was replaced with an equal volume of fresh 0.5M EDTA solution. After two-week incubation period, the samples were washed with distilled water and dried using blotting papers. Following this, the dried samples

were finely cut into small pieces for DNA extraction.

The muscle samples were cleaned to remove visible foreign materials from their surfaces by trimming with sterile forceps. Subsequently, the samples were subjected to washing procedure with sterile water at 56°C for 15 minutes under shaking conditions, followed by 3 minutes wash with 70% ethanol.

### DNA Analysis

Muscle samples, finely cut bone and teeth (approximately 2 grams) and reference blood samples were subjected to extraction using QIAmp DNA Investigator kit (Qiagen) following optimized extraction protocol for each sample type. DNA quantification was performed with Quantifier™ Human DNA Quantification Kit (Applied Biosystems, USA) according to manufacturer's protocol. PCR Amplification was carried out with currently available multiplex STR typing kits: AmpFISTR Identifiler Plus PCR Amplification Kit (Applied Biosystems, USA), PowerPlex® Fusion System (Promega Corp, Madison, WI, USA), PowerPlex® Y23 System (Promega Corp, Madison, WI, USA) and Qiagen, Investigator® Argus X-12 QS amplification kit. Separation of amplified PCR products was carried out on Genetic Analyser 3500 (Applied Biosystems, Life Technologies, Foster City, CA,

USA) and analyzed with GeneMapper ID, version 3.2, from Applied Biosystems using standard procedures.

### Results and Discussion

The primary objective in presenting these three case studies was to emphasize the significance of novel DNA identification methods in human remains identification and to highlight potential areas for future improvements in analyzing such challenging biological samples.

The quality and reliability of DNA testing is significantly influenced by setup of the DNA extraction laboratory. We consistently employ DNA-free chemicals and consumables while enforcing rigorous contamination control measures and assess all safety protocols. Through the development of advanced DNA extraction and typing procedures, currently we successfully conducted DNA analysis on these critical scenarios. However, in highly degraded or samples containing inhibitors, it is crucial to implement further improvements to ensure a more accurate identification on human remains.

The result of DNA profiles derived from human remains found in each location in case study-1 is summarized in the following Table 1.

Loci	Sample of bone Location-1	Sample of muscle Location-2
D8S1179	12,14	11,14
D21S11	28,30,2	29,30
D7S820	10,10	11,11
CSF1PO	12,12	12,12
D3S1358	18,18	15,16
TH01	7,10	8,9
D13S317	10,11	11,11
D16S539	10,12	8,10
D2S1338	18,23	16,21
D19S433	14,14	12,15
vWA	17,19	15,19
TPOX	8,9	11,11
D18S51	14,14	13,14
D5S818	10,13	11,12
FGA	24,26	23,25
Amel	X,Y	X,Y

**Table 1:** The Identifiler Plus, DNA Profiles Details for Human Remains Found on Location-1 and Location-2 (Case Study-1).

According to results from two locations two different male persons were identified. The muscle sample obtained

from location-1 not given positive results for none of the STR kits. This could be attributed to severe degradation or the

presence of DNA inhibitors in the extracted DNA. Additional efforts to search for the missing individuals cannot be initiated as the family members have not reported the disappearance of each person.

The DNA profiles obtained from human remains discovered in the garden in Case Study-2 are presented in the following Table 2.

Loci	Sample of teeth	Blood sample of Mother
D3S1358	14,15	14,17
D1S1656	16,17	16,17
D2S441	10,10	10,10
D10S1248	11,16	11,14
D13S317	9,12	11,12
Penta E	11,17	13,17
D16S539	10,14	11,14
D18S51	14,16	13,14
D2S1338	19,21	18,19
CSF1PO	11,12	11,12
Penta D	9,9	9,13
TH01	6,6	6,9
vWA	17,18	18,18
D21S11	30,31.2	31.2,33.2
D7S820	12,13	12,13
D5S818	11,12	11,12
TPOX	9,12	9,12
DYS391	10	-
D8S1179	13,14	13,15
D12S391	18,18	18,18
D19S433	14,15	15,16.2
FGA	20,21	20,20
D22S1045	15,16	11,16
Amel	X,Y	X,X

**Table 2:** The Powerplex Fusion System, DNA Profiles Details for Human Remains Found in the Garden and Reference Sample of Missing Person's Mother (Case Study-2).

A complete DNA profile was not obtained for the skull and teeth when using the Identifier kit containing minimum quantity of DNA. Skull being thin and often exposed is more susceptible to DNA degradation compared to teeth. Hence, it's imperative to understand the susceptibility of various skeletal elements to DNA degradation. This becomes effective when choosing skeletal elements that are less susceptible to degradation, like long bones, to enhance likelihood of obtaining viable DNA samples. However, complete DNA profile was obtained for teeth when using the PowerPlex Fusion kit. Hence, it can be assumed PowePlexFusion could be more suitable in scenario with low quantity of DNA,

offering not only successful profiling but also demonstrating high discriminating power.

The profile details of teeth from missing person revealed, each marker shared allele with the mother's genotype profile. The statistical analysis for maternal relationship was calculated using eDNA 2.3 software package, further revealed the relationship of the missing person with mother. The table below presents a Summary Matrix for the distribution of corresponding alleles along with the calculated probability value for both samples (Table 3).

Summary Matrix								
System	Child		Mother		PI	Mutation Code	Pattern	Rule
D3S1358	14	15	14	17	8.8246		/PQ/PR	1/4P wher P equals .0283
D1S1656	16	17	16	17	5.3655		/PQ/PQ	(P+Q)/4PQ wher P equals .1150 and Q equals .0783
D2S441		10		10	3.2258		/P/P	1/P wher P equals .3100
D10S1248	11	16	11	14	30.012		/PQ/PR	1/4P wher P equals .0083
D13S317	9	12	11	12	1.25		/PQ/QR	1/4Q wher Q equals .2000
penta E	11	17	13	17	2.2727		/PQ/QR	1/4Q wher Q equals .1100
D16S539	10	14	11	14	13.6388		/PQ/QR	1/4Q wher Q equals .0183
D18S51	14	16	13	14	0.8108		/PQ/PR	1/4P wher P equals .3083
D2S1338	19	21	18	19	1.7045		/PQ/PR	1/4P wher P equals .1467
CSF1PO	11	12	11	12	1.4923		/PQ/PQ	(P+Q)/4PQ wher P equals .29.0 and Q equals .3967
Penta D		9	9	13	1.7857		/P/PQ	1/2P Wher P equals .2800
TH01		6	6	9	2.1126		/P/PQ	1/2P Wher P equals .2367
Vwa	17	18		18	2.4793		/PQ/P	1/2Q Wher Q equals .2017
D21S11	30	31.2	31.2	33.2	2.2387		/PQ/QR	1/4Q wher Q equals .1117
D7S820	12	13	12	13	11.4151		/PQ/PQ	(P+Q)/4PQ wher P equals .1767 and Q equals .0250
D5S818	11	12	11	12	1.5647		/PQ/PQ	(P+Q)/4PQ wher P equals .3533 and Q equals .2917
TPOX	9	12	9	12	10.8162		/PQ/PQ	(P+Q)/4PQ wher P equals .1733 and Q equals .0267
D8S1179	13	14	13	15	1.8519		/PQ/PR	1/4P wher P equals .1350
D19S433	14	15	15	16.2	0.25		/PQ/QR	1/4Q wher Q equals 1.0000
FGA	20	21		20	3.5714		/PQ/P	1/2P wher P equals .1400
D22S1045	15	16	11	16	1.8073		/PQ/QR	1/4Q wher Q equals .1383
CRI POP = 4,428,132,371.8883								
Probability = 99.9999%								

**Table 3:** The Summary Matrix for Allele Distribution of Human Remains of Teeth and Blood Sample of Mother (Case Study-2).

In order to find the maternal lineage, the samples were analyzed with Investigator® Argus X-12 kit. The results are

illustrated in the following Table 4.

Loci	Sample of teeth	Blood sample of mother
DXS10103	16	16
DXS8378	11	11
DXS10101	34	34
DXS10134	36	36
DXS10074	16	16,19
DXS7132	16	13,16
DXS10135	20	20,25
DXS7423	15	14,15
DXS10148	32	27,32
DXS10079	19	18,19

HPRTB	13	13,14
DXS10146	21.1	18,25.1
D21S11	30,31.2	31.2,33.2
Amel	X,Y	X,X

**Table 4:** The Investigator Argus X-12, DNA Profiles for Human Remains Found in the Garden and Reference Sample of Missing Person's Mother (Case Study-2).

X chromosome results further revealed the relationship between missing person with his mother.

The resulting DNA profiles from human remains discovered in the jungle in Case Study-3 are outlined in the following table.

Loci	Sample of bone	Blood sample of father
D8S1179	9,16	14,16
D21S11	30,32.2	30,31
D7S820	7,12	12,12
CSF1PO	12,12	11,12
D3S1358	16,16	14,16
TH01	9,9	9,9
D13S317	8,13	8,14
D16S539	12,12	11,12
D2S1338	16,23	16,25
D19S433	13,14	14,14.2
vWA	17,18	18,18
TPOX	11,13	8,13
D18S51	13,14	14,20
D5S818	10,12	10,13
FGA	21,24.2	23,24.2
Amel	X,Y	X,Y

**Table 5:** The Identifier plus DNA Profiles for Human Remains Found in the Jungle and Reference Sample of Missing Person's Father (Case Study-3).

The profile details of bone sample from missing person indicated each marker shared allele with father's DNA profile. The statistical analysis for paternal relationship further revealed the relationship between missing person

with his father. The table below presents a Summary Matrix for the distribution of corresponding alleles along with the calculated probability value for both samples.

Summary Matrix										
System	Mother		Child		Alleged Father		PI	Mutation Code	Pattern	Rule
D8S1179			9	16	14	16	2.9412		/PQ/QR	1/4Q where Q equals .0850
D21S11			30	32.2	30	31	1.4706		/PQ/PR	1/4P where P equals .1700
D7S820			7	12		12	2.8301		/PQ/Q	1/2Q where Q equals .1767
CSF1PO				12	11	12	1.2605		/P/PQ	1/2P where P equals .3967
D3S1358				16	14	16	1.6129		/P/PQ	1/2P where P equals .3100

TH01			9		9	2.9703		/P/P	1/P where P equals .3367
D13S317		8	13	8	14	1.2		/PQ/PR	1/4P where P equals .2083
D16S539			12	11	12	2.5641		/P/PQ	1/2P where P equals .1950
D2S1338		16	23	16	25	30.012		/PQ/PR	1/4P where P equals .0083
D19S433		13	14	14	14.2	0.25		/PQ/QR	1/4Q where Q equals 1.0000
vWA		17	18		18	2.4793		/PQ/Q	1/2Q where Q equals .2017
TPOX		11	13	8	13	30.012		/PQ/QR	1/4Q where Q equals .0083
D18S51		13	14	14	20	0.8108		/PQ/QR	1/4Q where Q equals .3083
D5S818		10	12	10	13	2.2727		/PQ/PR	1/4P where P equals .1100
FGA		21	24.2	23	24.2	30.012		/PQ/QR	1/4Q where Q equals .0083
CRI Pop = 7,022,840.0759									
Probability = 99.9999%									

**Table 6:** The Summary Matrix for Allele Distribution of Human Remains of Bone and Blood Sample of Father (Case Study-3).

To establish paternity lineage Y-STR was performed and the profile details are illustrated in the following table.

Loci	Sample of bone	Blood sample of father
DYS576	17	17
DYS3891	12	12
DYS448	21	21
DYS389II	30	30
DYS19	17	17
DYS391	10	10
DYS481	25	25
DYS549	13	13
DYS533	10	10
DYS438	10	10
DYS437	14	14
DYS570	20	20
DYS635	21	21
DYS390	24	24
DYS439	12	12
DYS392	12	12
DYS643	11	11
DYS393	13	13
DYS458	18	18
DYS385	15,17	15,17
DYS456	15	15
YGATH4	11	11

**Table 7:** The Powerplex Y23 System, DNA Profiles for Human Remains Found in the Jungle and Reference Sample of Missing Person's Father (Case Study-3).



The Y-chromosome haplotypes of the missing person's bone sample and blood sample of father matched at all of the Y-chromosome STR loci tested, indicating that they are likely to share the same paternal line.

The outcomes of these case studies describe the importance of adapting DNA recovery and amplification strategies to the unique characteristics of each forensic scenario. The advancements employing in these studies not only contribute to the scientific understanding of degraded human remains but also have practical implications for improving the reliability of forensic testing in challenging conditions.

## Conclusion

Forensic genetics encounters significant challenges when identifying human remains and establishing links to missing persons, especially when DNA fingerprinting is the sole viable option. This challenge arises due to the advanced decomposition or degradation of the human remains and other morphological or alternative methods are impractical or unfeasible. In such cases, DNA analysis becomes crucial for providing valuable insights into the identity and potential connections of the deceased individual with missing person's relatives. However when there are no missing person's information and reference samples of relatives, create significant challenges. At present, Government Analyst's Department doesn't have a dedicated database for identifying missing persons. In future, enhancing DNA databases to missing person identification will enhance victim identification, particularly in cases where reference samples are absent.

Conducting DNA analysis on human remains is complex and involving lengthy process. Therefore, dealing with a large volume of skeletal remains collected from a scene can present challenges. Understanding the susceptibility of various skeletal remains to DNA degradation is vital when determining suitable materials for analysis. In cases where DNA preservation is paramount, choosing bones that are less prone to degradation, such as long bones, improve the chances of obtaining viable DNA samples. This strategic selection ensures a higher likelihood of successful DNA analysis and aids in the accurate identification of individuals in decomposed human remains.

The current best practice applying in the DNA laboratory in Government Analyst's Department for identifying human remains involves optimized extraction methods tailored to different tissue types, employing different types of STR

amplification kits depending on quality and quantity of the extracted DNA and utilizing lineage markers for further identification. This approach serves as currently following guideline for effective identification of human remains under challenging conditions. Continuous advancements in forensic technologies, specifically the improvement of DNA analysis techniques are helping us to make more accurate identifications. We are strategically collaborating with international forensic laboratories to enhance our capabilities concentrating on improving forensic technologies to ensure greater precision in DNA analysis of decomposed human remain identification. However, further improvement is imperative and could involve incorporating newly developed technologies in analyzing severely degraded samples and enhancing missing person databases to expedite identification processes, ultimately leading to more precise identifications in human remains cases.

## Ethical Issues

The study adhered to the guidelines set forth by the Government Analyst's Department and was conducted in accordance with principled standards.

## References

1. Belcher WR, Shiroma CY, Chesson LA, Berg GE, Jans MJ (2021) The role of forensic anthropological techniques in identifying America's war dead from past conflicts. *Wires Forensic Science* 4(3): 1-18.
2. De Boer HH, Blau S, Delabarde T, Hackman L (2019) The role of forensic anthropology in disaster victim identification (DVI): recent developments and future prospects. *Forensic Science Research* 4(4): 303-315.
3. Senst A, Caliebe A, Drum M, Cossu C, Zieger M, et al. (2023) Recommendations for the successful identification of altered human remains using standard and emerging technologies: Results of a systematic approach. *Forensic Science International: Genetics* 62: 102790.
4. Ajay K, Dasari H, Amandeep S, Kulbhushan, Sunil KGA (2014) Unknown Dead Bodies: Problems and Solutions. *Journal of Indian Academy of Forensic Medicine* 36(1): 76-80.
5. Senst A, Scheurer E, Gerlach K, Schulz I (2021) Which tissue to take? A retrospective study of the identification success of altered human remains. *Journal of Forensic and Legal Medicine* 84: 102271.

