

Palynostratigraphy of Upper Mesozoic Outcrops near Anjiajia (Mahajanga Basin), and Manamana (Morondava Basin), and It's Bearing on the Development of Cretaceous Angiosperm Floras in Madagascar

Voajanahary R^{1*}, Zavada Michael S² and Toussaint R¹

¹Department of Sedimentary Basins Evolution and Conservation, University of Antananarivo, Madagascar

²Department of Geoscience, University of Texas, USA

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***Corresponding author:** Ranaivosoa Voajanahary, Department of Sedimentary Basins Evolution and Conservation, Faculty of Sciences, University of Antananarivo, BP 906, Antananarivo 101, Madagascar, Email: rnvoaj@gmail.com

Abstract

The two outcrops were investigated palynologically. One outcrop is approximately a 17meter section near Anjiajia in the Mahajanga Basin and the second outcrop is approximately a 5meter section located in the Manamana massif-South Ankazoabo in the Morondava Basin. The section from Anjiajia was dominated by Cryptogrames (56%), Gymnosperms (29.5%) and a minor component of angiosperms (19%). Based on the sedimentology, composition of the palynoflora, the low percentage of angiosperms, and the occurrence of the dinoflagellate *Litosphaeridium arundum* at the Anjiajia sample is estimated to be Albian to Cenomanian in age. The section from Manamana is dominated by Cryptogrames (47%), Angiosperm (30.5%) and Gymnosperms (17.5%). Based on the sedimentology, floral composition, the higher percentage of angiosperms relative to the Anjiajia sample, the occurrence of the Dinoflagellates *Leberidocysta chlamydata* and *Odontochitina operculata* and the location of the section beneath the Coniacian basalts in the Morondava basin, these data suggest a Turonian-Coniacian age for this sample. Based on these two Cretaceous palynofloras and a megafossil flora from the Maevarano Formation (Santonian to Maastrichtian), which has only a few species of gymnosperms and is dominated by the Lower Cretaceous angiosperm *Sapindopsis*, it appears the southern hemisphere floras are lagging behind the northern hemisphere angiosperm floras in diversity and abundance.

Keywords: Angiosperm floras; Palynology; Cretaceous; Mahajanga Basin; Morondava Basin

Introduction

Madagascar separated from the African Continent about 160 million years ago and subsequently from India about 88 mya [1-6]. The isolation of Madagascar in the mid Cretaceous and through the Late Cretaceous and Tertiary has been implicated in the high rate of extant endemism in the fauna and flora of Madagascar. Madagascar is one of the seven biodiversity hotspots in the world [7]. The primate fauna has attracted worldwide interest of scholars and conservationist) [7]. The Late Tertiary fauna and flora has been of particular interest to track and reconstruct the development of

Madagascar's unique biota through the Tertiary and the Holocene [8-11]. Palynological investigations may be used to detect the effects of climatic changes on terrestrial plant vegetation and as additional tool to envisage the past climatic changes [12].

The Mesozoic dinosaurian fauna has been equally significant and has attracted attention [13,14]. Madagascar's flora has also attracted interest because of its diversity and the high number of endemic taxa [15-17], especially the occurrence of basal angiosperms, e.g. the genus Takhtajania perrieri of the Winteraceae [18]. However, interest and recovery of the megafossil flora from the Mesozoic and Tertiary are confined to reports of various species of microfossils [19-26] and just a few reports of megafossil floras [27-29]. Otherwise, the Mesozoic and Tertiary megafossil floras have remain largely unknown.

In the 1980s and in subsequent field seasons our field team began surveys to assess the abundance and distribution of plant fossils in the Mesozoic sediments of the Morondova and Mahajanga Basins. The Mesozoic sequences within these two basins preserve a number of high-energy environments. Environments more conducive to vertebrate preservation than the preservation of plants [13,14]. There have been reports of petrified wood in Madagascar, but few have been studied [30]. During our field surveys, palynological samples were collected from both basins, however, few of these samples produced palynofloras that could provide insight into the floristic composition and abundance of various taxonomic groups. One location in the Mahajanga Basin (Maevarano Formation) produced various species of leaf impressions including angiosperms. This Upper Cretaceous (Campanian -Maastrichtian) age locality is dominated by the primarily early Cretaceous taxon Sapindopsis presumed to be closely related to members of the Platanaceae (see below), however this locality was disturbed by subsequent road work and was unable to be relocated to extract more extensive collections.

This study is to characterize the palynofloras from the Anjiajia site, Mahajanga basin, and the Manamana site, Morondava basin and to assess the age of the sediments based on the microfossil assemblages. This study is to expand our understanding of the fossil flora of Madagascar during the Mesozoic, particularly during the first major diversification of the angiosperms in the world.

Collection Localities and Sedimentology

Anjiajia

Anjiajia is located in the Mahajanga Basin (S 16°22'15", E 46°55'02"). Geographically, the area studied is in the

southwest part of the Mahajanga Basin, 35 km east of the District of Ambato- Boeny on National Route 4. Between the Isalian sandstone reliefs and the northwest coast lay the alluvial plains of Anjiajia, Madirovalo and Marovoay, which were the focus of this field survey [31-33] (Figure 1).



Figure 1: Location of the Anjiajia site, Mahajanga Basin. P1_ANJ is located at S 160 31' 59.9" E 460 56' 42.6", and P2_ANJ is located at S 160 27' 48.7" E 460 57' 42.6".



The 17.5-meter section near Anjiajia is comprised of seven (7) lithological units (Figure 2). The base of the section consists primarily of a fine-grained green clay interbedded occasionally with a cross bedded sandstone (A1) with traces of iron-stained clay and concretions. A2 is weathered grey sandy clay to siltstone, probably a paleosol. A3 is a sandy silt

to claystone becoming variegated grey to red at the top, and grading into the A4 grey clay. The upper portion of this section A5 is characterized by the succession of green interbedded clays with cross-bedded sandstones. A6 is a sandy siltstone to clay and is the unit that produced the playnoflora. A6 grades into a fine-grained sandy silt and sandstone clay (A7) about 3 meters thick (Figure 2).

The entire outcropping section was sampled and processed for microfossil. The A6 level was the only lithological unit to produce a palynoflora.

Manamana

The 5-meter Cretaceous portion of the section at Manamana is located in the Morondava basin southwest of the District of Ankazoabo, and located 3 km from the fokontany of Ambatomainty whose geographical coordinates are: S 22°30'00", E 44° 27' 00". This site has been characterized as Upper Jurassic sediments (Callovien-Oxfordian) grading into Cretaceous sediments [34] (Figure 3).



This section at Manamana has seven (7) distinctive levels (Figure 4). The Cretaceous part of the section has a thickness of about 5 m. The basal bed (M1) is a fine grained, friable white sandstone overlain by a fine to medium grained brown cross-bedded sandstone (M2). The M1 – M2 levels comprise the Jurassic portion of the section. The Cretaceous M3 is a medium to course grained greenish-grey sandstone and is overlain by the M4 consolidated sandstone. M5 is cross-bedded fine-grained sandstone interbedded with silt to claystone with invertebrate fossils and is the lithological unit that produced the palynoflora. M5 is capped off by consolidated blocky sandstone (M6). The overall section is overlain by a basalt (Figure 4).



Figure 4: Stratigraphic section that was sampled near Manamana. Note the section is just below the basalts of Coniacian Age [35-40].

Sample Processing Methods

All of the samples were processed according to the method of Barss and Williams [41] modified by Zavada [42] for palynological analysis. Slides were examined to determine the taxonomic affinities of the palynomorphs and the relative abundance of each taxon in the samples. These data along with an analysis of the sedimentological data were used for the elucidating the paleoenvironments of the two localities (after Chateauneuf [43] and Reyre; Demchuk, et al. [44].

The Pollen and Spore Flora, Age and Depositional Environment

The palynoflora from Anjiajia has approximately 29 taxa and the flora from Manamana has approximately 32 taxa and both are considered to be Cretaceous. Another feature of these palynoforas is that the two localities share 80 percent of the same taxa suggesting that the two palynofloras may be close in age (Table 1).

The major difference between the two localities is the abundance of angiosperms. Anjiajia is comprised of 19% angiosperms and the Manamana has 30.5% angiosperms (Table 1).

The difference in the percentage of angiosperm pollen indicates that the Anjiajia palynoflora is of Lower Cretaceous age and the Manamana percentage is more indicative of floras from the Lower-Upper Cretaceous [45]. The increase in the percentage of angiosperms may represent the Lower and Upper Cretaceous angiosperm diversification that was occurring in many regions of the world [45,46]. Fifteen of the twenty-one taxa of spore and pollen in the Anjiajia sample are common Lower Cretaceous palynofloras, but not necessarily restricted to Lower Cretaceous palynofloras [47-50].

Taxon	Anjiajia (A6)	Manamana (M5)
Aequitriradites spp.	4	1
Appendicisporites baconicus	2	6
Appendicisporites tricomitus	6	7
Appendicisporites dorogensis	2	2.5
Appendicisporites potomaensis	5	1.5
Cicatricosporites australiensis	4	10
Cicatricosporites ludbrookiae	6	4
Cingulatisporites sp.	4	1
Concavisporites sp.	5	1
Cyathidites spp.	NP	2.5
Gleicheniidites spp.	2	NP
Matonia pectinata	2	2.5
Marattiopsis	1	1
Osmundacidites	3	NP
Paxillitrilites dakotaensis	NP	1
Polypodiasporites sp.	4	2
Retitriletes sp.	4	NP
Verrucosisporites rarus	2	2
TOTAL CRYPTOGAMS	56	45
Alisporites grandis	1	1.5
Araucariacites australis	3	2
Balmeiopsis imbatus	4	NP
Callialasporites segmentatus	NP	1.5
Callialasporites trilobatus	1.5	NP
Classopollis classoides	NP	2
Cycadopites spp.	NP	2
Microcachrydites antarcticus	12	2
Podocarpidites spp.	3	4
Trichotomonosulcites subgranulosus complex	5	2
TOTAL GYMNOSPERMS	29.5	17
<i>Clavatipollenites</i> Figure 5A	NP	5
Ericipites spp.	5	2.5
Liliacidites giganteus	NP	4
Monocolpopollenites sp.	4	2
Monosulcites Figure 5C	4	4
Proteacidites polymorphus	2	2
Retimonocolpites Figure 5B	2	7
Tricolpites spp. Figures D,E,F	2	4
TOTAL ANGIOSPERMS	19	30.5
UNDETERMINED	5.5	5
REWORKED POLLEN	0	2.5

Table 1: The percentages of the taxa observed from Level A6(Anjiajia) and M5 (Manamana). NP = Not Present.

All of the angiosperm taxa recovered from the Anjiajia locality occur frequently in lower Cretaceous palynofloras in other regions of the world [51]. The common occurrence and abundance of Lower Cretaceous spores and pollen, coupled with the low percentage of angiosperms indicates a Lower Cretaceous to Lower - Upper Cretaceous age for the Anjiajia and suggest this locality belongs to the Ankazomihaboka Beds Rogers, et al. [33] or the Serie d'Ankazomihaboka of Besairie [31] (Table 1). The presence of the dinoflagellate cyst *Litosphaeridium arundum*, further refines the age to Albian [52-56] (Table 2).

Dinoflagellate / Location	Anjiajia (A6)	Manamana (M5)	Age range
Litosphaeridium arundum	Х	NP	Albian
Leberidocysta chlamydata	NP	Х	Albian- Cenomanian
Odontochitina operculata	NP	Х	Senonian

Table 2: Distribution and age range of identifiabledinoflagellates. NP = Not Present.

Although the rock units from these two sites indicate similar depositional environments, they two localities may be of two different ages. The sediments at Manamana are considered to be Upper Cretaceous possibly Coniacian. This palynoflora does have some common Upper cretaceous taxa that are not found in the Anjiajia palynoflora, e.g. Liliacidites giganteus, Paxillitriletes dakotaensis, Cyathidites, and *Cycadopites* (Table1). The occurrence of the dinoflagellates Leberidocysta chlamydata and Odontochitina operculata suggest a Turonian-Coniacian (Senonian) age for the beds, but no older than the Cenomanian [57] (Table 2). A basalt overlays the entire section (Figure 4). During the Cretaceous, Madagascar and India underwent a southward lateral displacement, which resulted in the initial separation of Madagascar and India causing magmatism during and after the rifting, these basalts in the Morondava Basin are a result of this rifting. The basalts have been dated Coniacian [35-40] and support the interpretation that the Cretaceous section, which lies beneath these basalts is of Senonian age (Turonian- Coniacian).

Discussion

Rifting between east and west Gondwanaland began about 176 million years ago [58]. This initiated a breakup of Africa (west Gondwanaland) and Madagascar which occurred about 165 million years ago [5,6,58]. Madagascar drifted southeast along the trend of the Davie Ridge. Spreading ceased approximately 130 my ago. Since that

time Madagascar has not changed its position significantly [59]. This event first isolated Madagascar from the African continent (western Gondwanaland), and subsequently in the Coniacian, India separated from Madagascar and began to drift northward, reducing the land area that can act as a target for migrating organisms to Madagascar and further isolated the island through the remainder of the Cretaceous and Tertiary. The high rate of extant endemism in Madagascar is attributed to this early isolation. Concurrent with these tectonic events during the Upper Jurassic to Coniacian, is the Lower Cretaceous and Upper Cretaceous worldwide distribution and diversification of angiosperms.



Figure 5: Representative Palynomorphs from the Manamana Locality in the Morondova Basin, similar taxa also occur at Anjiajia. A) *Clavatipollenites* sp., Scale Bar = 5 um. B) *Retimonocolpites* sp. Scale Bar = 10 um C) Monosulcites Scale Bar = 8 um D) *Tricolpites* sp. Scale Bar = 8 um. E) Striate – Tricolpate, Scale Bar = 3 um F) *Tricolpites* sp. cf. Rousea, a tricoplate type. Scale Bar = 8 um

Angiosperms comprised a relatively small percent of palynofloras during the Lower Cretaceous (no more than 15-20% of a flora, Lidgard and Crane,) [45]. During the Upper Cretaceous, however, palynofloras steadily increased

in diversity and increased as a percent of the palynoflora reaching about 55% in the Northern Hemisphere palynofloras in the Maastrichtian [45,60]. The two palynofloras in this study share about 80 % of the same or similar taxa (Figure 5), however angiosperm taxa through this time interval in Madagascar continue to be a relatively minor component of the palynofloras, approximately 19% in the lower Cretaceous and 30% by the Coniacian. This lags behind the increase in diversity and dominance observed in other parts of the world, notably in the Northern Hemisphere.

Megafossil floras also track the increase in angiosperm diversity and dominance that the palynofloras exhibit during this time interval. In addition to the two palynofloras in Madagascar our team also found a megafossil locality 25 km north of the turn off to Marovoay in the Maevarano Formation just off of the RN4 [33]. The Maevarano Formation is primarily Upper Cretaceous (Santonian to Maastrichtian) in age, and has produced a rich dinosaurian fauna. The megafossil flora produce an assemblage of leaves, seeds and insect remains (mostly Beetle elytra). An unusual feature of this flora is it is dominated by Sapindopsis leaves with only 2-3 other gymnosperm species present. Sapindopsis is a taxon more indicative of Lower Cretaceous and Lower Upper Cretaceous floras than the more diverse Late Cretaceous megafossil floras. Sapindopsis leaves are simple, margins are entire, leaves are alternate, linear to elliptic in outline. The terminal portion of the leaf is acute, and basal portion is cuneate (Figure 6, A, B,C). The leaf has a prominent midrib and the secondary venation appears to be alternate to subopposite. The leaves are net veined (Figure 6 C).

Beetle elytra are common at this Upper Cretaceous megafossil locality (Figure 6, D,E), in addition to seed and fruit- like structures (Figure 6, F). The flora is dominated by a single species of *Sapindopsis*, i.e. a monculture of this early angiosperm. Beetles may be predators or pollinatiors of this species of angiosperm. The pattern visible on the elytra is found in the Curculionidae, Tenebrionidae, and in Boganiidae, Oedemeridae. The latter two families are known from the Cretaceous and have been implicated as angiosperm pollinators [61].

Much like the palynofloras, the megafossil flora was not as diverse as fossil floras found elsewhere in the world in the same time interval. This suggest that the initial angiosperm diversitification and migration may have been dampened by the isloation of Madagascar due to tectonic events in the southern hemisphere.

The paleoenvironmental setting of the two localities based on the palynomorph assemblages and the sedimentology represent near shore environments, i.e., lower delta plain [43,44].

Figure 6: Examples of the megafosssil flora recovered from the Maevarano Formation (Campanian – Maastrictian) 25 km north of the turn off to Maroavay on the NR 4. A) The terminal portion of a *Sapindopsis* leaf showing the acut apex (arrow). Note the prominent midrib. Scale Bar = 4 mm. B) A portion of a *Sapindopsis* leaf showing the cuneate basal portion of the leaf and the petiole (arrow). Note that the

leaf venation appears to be alternate to sub-opposite. Scale Bar= 4 mm C) Terminal branch of *Sapindopsis* showing the alterate arrangement of the leaves. Scale Bar = 4 mm D-E) Beetle elytron, possibly of the families Curculionidae (e.g. *Hylastinus* sp), Tenebrionidae (e.g. *Tribolium*), Boganiidae, or Oedemeridae. Scale Bar = 1 mm F) Possibly a fourloculate capsule (arrows) Scale Bar= 1 mm.

Conclusion

The timing of the breakup of Madagascar from the African Continent in the Upper Jurassic and the subsequent separation of Madagascar from India in the coniacian severed Madagascar's connection to continuous land masses during the initial diversification and migration of the angiosperms throughout the world [6]. It is also interesting that angiosperm diversification in southern Africa [62] and Australia [63] exhibit a similar pattern, i.e., a lag in the diversification and dominance of angiosperms through the Lower and Upper Cretaceous of these continents. The timing of these tectonic events may have prevented the spread of angiosperms not only to Madagascar, but to other areas in the southern hemisphere. The approximately 90 million years of isolation have significantly contributed to the high rate of endemism in the fauna and flora of Madagascar.

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References

- 1. Heirtzler JR, Burroughs RH (1971) Madagascar's paleoposition: New data from the Mozambique channel. Science 174(4008): 488-490.
- Embleton BJJ, McElhinny MW (1975) The paleoposition of Madagascar: Palaeomagnetic evidence from the Isalo Group. Earth and Planetary Science Letters 27(2): 329-341.
- 3. McElhinny MW, Embleton BJJ, Daly L, Pozzi JP (1976) Paleomagnetic evidence for the location of Madagascar in Gondwanaland. Geology 4(8): 455-457.
- 4. Leroy JF (1978) Composition, Origin, and Affinities of the Madagascan Vascular Flora. Ann Missouri Bot Gard 65(2): 535-589.
- 5. Rabinowitz PD, Coffin MF, Falvey DF (1983) The Separation of Madagascar and Africa. Science 220(4592): 67-69.
- 6. Reeves C (2014) The position of Madagascar within Gondwana and its movements during Gondwana dispersal. J Afr Earth Science 94: 45-57.
- 7. Myers N, Mittermeier RA, Mittermeier CG, da Fonseca GAB, Kent J (2000) Biodiversity hotspots for conservation priorities. Nature 403: 853-858.
- 8. MacPheeRDE, BurneyDA, WellsNA(1985) Early Holocene Chronology and Environment of Ampasambazimba, a Malagasy Subfossil Lemur Site. International Journal of Primatology 6: 463-489.
- Burney DA (1987a) Late Holocene vegetational change in Central Madagascar. Quaternary Research 28(1): 130-143.
- 10. Burney DA (1987b) Pre-Settlement vegetation changes at Lake Tritrivakely, Madagascar. In: Coetzee JA, (Ed.), Palaeoecology of Afriac and the Surrounding Islands, A A Balkema, Rotterdam, pp: 357-381.

- 11. Gasse F, Van Campo E (2001) Late Quaternary environmental changes from a pollen and diatom record in the southern tropics (Lake Tritrivakely, Madagascar). Palaeogeography, Palaeoclimatology, Palaeoecology 167(3-4): 287-308.
- 12. Azzazy MF, Marco AVC (2020) Contribution to the Eco-Palynological Studies of Wadi El Natron, Egypt. Int J Paleobiol Paleontol 3(1): 000111.
- Flynn JJ, Parrish JM, Rakotosamimanana B, Simpson WF, Whatley RL, et al. (1999) A Triassic Fauna from Madagascar, Including early dinosaurs. Science 286(5440): 763-765.
- 14. Rogers RR (2005) Fine-grained debris flowsand extraordinary vertebrate burials in the Late Cretaceous of Madagascar. Geological Society of America 33(4): 297-300.
- 15. Schatz GE (2001) Generic Tree Flora of Madagascar. Royal Botanic Gardens, Kew and Missouri Botanical Gardens. The Cromwell Press, pp: 477.
- 16. Phillipson PB, Schatz GE, Lowry II PP, Labat JN (2006) A catalogue of the vascular plants of Madagascar. In: Ghazanfar SA, Beentje HJ (Eds.), Taxonomy and ecology of African plants, their conservation and sustainable use, Royal Botanic Gardens, Kew, pp: 613-627.
- Callmander MW, Phillipson PB, Schatz GE, Andriambololonera S, Rabarimanarivo M, et al. (2011) The endemic and non-endemic vascular flora of Madagascar updated. Plant Ecology and Evolution 144(2): 121-125.
- 18. Schatz GE, Lowry II PP, Ramisamihantanirina A (1998) Takhtajania perrieri rediscovered. Nature 391: 133-134.
- 19. De Jekhowsky B, Goubin N (1962) Sub surface palynology in Madagascar. Permian, Triassic and Jurassic of Morodava Basin. Bull Am Assoc Pet Geol 46(2): 269-270.
- 20. De Jekhowsky B, Goubin N (1963) Esquisse palynologique du Permian, Tras, Jurassique du bassin de Morondava a Madagascar. Ann Geol Madagascar 33: 163-165.
- De Jekhowsky B, Goubin N (1964) Subsurface palynology in Madagascar: a stratigraphic sketch of the Permian, Triassic and Jurassic of the Morondava Basin. In: Cross AT, (Ed.), Palynology in Oil Exploration, Soc Econ Paleontol Mineral, pp: 116-130.
- 22. Goubin N (1965) Description and distribution of the main Permian Triassic and Jurassic pollenites from the soundings of the Morondava Basin (Madagascar).

Journal of the French Petroleum Institute and Annales des Combustibles Liquides 20(10): 1415.

- 23. Rakotoarivelo HJ (1970) Comparative palynostratigraphy of the Gondwanan coal basin of Sakoa-Sakamena, Madagascar, PhD thesis, Faculty of Sciences of Paris, pp: 404.
- 24. Herngreen GFW, Randrianasolo A, Verbeek JW (1982) Micropaleontology of Abian to Danian strata in Madagascar. Micropaleontology 28(1): 97-109.
- Hankel O (1993) Early Triassic plant microfossils from Sakamena sediments of the Majunga Basin. Madagascar Rev Paleobot Palynol 77: 213-233.
- 26. Zavada MS (2003) The ultrastructure of angiosperm pollen from the Lower Cenomanian of the Morondava Basin, Madagascar. Grana 42(1): 20-32.
- Carpentier A (1936) L'Etude de la Flore du Groupe de la Sakamena. Annales Geologiques du Service des Mines 6: 35-40.
- Appert O (1973) Die Pteridophyten aus dem Oberen Jura des Manamana in Sudwest-Madagaskar, Schweiz Abt B 94: 1-58.
- 29. Appert O (1977) Die Glossopterisflora der Sakoa in Sudwest-Madagaskar. Paleontographica, Schweiz Abt B 162(1-3): 1-50.
- Yoon CJ, Kim KW (2008) Anatomical descriptions of silicified woods from Madagascar and Indonesia by scanning electron microscopy. Micron 39(7): 825-831.
- 31. Besairie H (1972) Géologie de Madagascar-1, Les terrains sédimentaires. Ann Géol Mada Fasc 35(463): 89.
- 32. Besairie H (1973) Précis de Géologie malgache. Ann Géol Mada Fasc 141(36): 3.
- Rogers RR, Hartman JH, Krause DW (2000) Stratigraphic Analysis of Upper Cretaceous Rocks in the Mahajanga Basin, Northwestern Madagascar: Implications for Ancient and Modern Faunas. Journal of Geology 108(3): 275-301.
- 34. Razafimbelo E (1987) Le bassin de morondava (madagascar): synthese geologique et structural. Thesis, pp: 241.
- 35. Blant G (1973) Structure and Paleogeography of the Southern and Eastern Coast of Africa. Association of African Geological Surveys, Paris, pp: 15e.
- 36. Storey M, Mahoney JJ, Saunders AD, Duncan RA, Kelly

SP, et al. (1995) Timing of hot spot-related volcanism and the breakup of Madagascar and India. Science 267(5199): 852-855.

- 37. Storey M, Mahoney JJ, Saunders AD (1997) Cretaceous basalts in Madagascar and the transition between plume and continental lithosphere mantle sources. Large Igneous provinces: continental, oceanic, and planetary flood volcanism. American Geophysical Union, Geophys. Monographs 100: 95-122.
- Gioan P, Rasendrasoa J, Bardintzeff JM, Rasamimanana G (1996) Nouvelles données pétrographiques et structurales sur le magmatisme du Sud du bassin de Morondava (Sud-Ouest de Madagascar). J Afr Earth Sci 22(4): 597-608.
- 39. Rasamimanana G, Bardintzeff JM, Rasendrasoa J, Bellon H, Thouin C, et al. (1998) Les episodes magmatiques du Sud-Ouest de Madagascar (bassin de Morondava), marqueurs des phenomenes de rifting cretace et neogene. Comptes Rendus de l'Academie des Sciences, Series IIA, Earth and Planetary Science 326(10): 685-691.
- 40. Bardintzeff JM, Bonin B, Rasamimanana G (2001) The Cretaceous Morondava volcanic province (west Madagascar): mineralogical, petrological and geochemical aspects. J Afr Earth Sci 32(2): 299-316.
- 41. Barss MS, Williams GL (1973) Palynology and Nannofossil processing techniques. Geol Survey Canada, pp: 73-26.
- 42. Zavada MS (2007) Botanical Methods. Bent Tree Press, Reno, NV, pp: 116.
- 43. Chateauneuf JJ, Reyre YY (1974) Elements of palynology: geological applications: third cycle course in Earth Sciences. Laboratory of paleontology of the University of Geneva, Geneva 1: 345.
- 44. Demchuk TD, Dolby G, McIntyre DJ, Suter JR (2008) The utility of palynofloral assemblages for the interpretation of depositional paleoenvironments and sequence-stratigraphic systems tracts in the McMurray Formation at Surmont, Alberta. AAPG Search and Discovery.
- Lidgard S, Crane PR (1990) Angiosperm diversification and Cretaceous Floristic Trends: A Comparison of Palynofloras and Leaf Macrofloras. Paleobiology 16(1): 77-93.
- 46. Heimhofer U, Hochuli PA, Burla S, Dinis JML, Weissert H (2005) Timing of Early Cretaceous angiosperm diversification and possible links to major

paleoenvironmental change. Geology 33(2): 141-144.

- 47. Thiergart F (1949) The stratigraphic value of Mesozoic pollens and spores. Pal Abt B Allen Bd 89(13): 1-34.
- Samoilovich SR (1967) Tentative botanico-geographical subdivision of northern Asia in Late Cretaceous time. In Review of Palaeobotany and Palynology 2(1-4): 127-139.
- 49. Singh HP, Venkatachala BS (1988) Upper Jurassic-Lower Cretaceous spore-pollen assemblages in the peninsular India. Palaeobotanist 36: 168-196.
- 50. Rafamantanantsoa JG (1991) Contribution to the micropaleontological study (spores, pollens and diatoms) of four peatlands in the Highlands of Madagascar. Energy recovery, Paleoecology, Dynamics of environments, 3rd cycle thesis, University of Antananarivo, Madagascar, pp: 223.
- 51. Doyle JA (1969) Cretaceous angiosperm pollen of the Atlantic Coastal Plain and its evolutionary significance. Journal of the Arnold Arboretum 50(1): 1-35.
- 52. Foucher JC (1980) Dinoflagelles and Acritarches. In: Robaszynski F, (Ed.), Biostratigraphic synthesis from the Aptian to the Santonian of the Boulonnais from seven groups Paleontological-foraminiferes, Nannoplankton, dinoflagella and Macrofauna. Journal of Micropaleontology 22: 228-290.
- 53. Fauconnier D (1984) Optical methods of assessing the degree of thermal evolution of palynofacies development of a reference scale. Geological and Mining Research Office.
- 54. Helby R, Morgan R, Partridge AD (1987) A palynological zonation of the Australian Mesozoic. Mem Assoc Australas Paleontol 4: 1-94.
- 55. Tocher BA (1995) Dinocyst distributions and stratigraphy of two Cenomanian-Turonian boundary (Upper Cretaceous) sections from western Anglo-Paris Basin. J Microplaeontology 14(2): 97-105.
- 56. Digbehi ZB, Ouffouet KB, Sombo BC, Yao NJP (2011) Biostratigraphical Analysis and Palaeoenvironmental Reconstruction of Cenomanian-Turonian Boundary in Côte d'Ivoire Onshore Sedimentary Basin, West Africa. African Research Review 5(6): 361-380.
- 57. Stover LE, Evitt WR (1978) Analyses of pre-Pleistocene organic-walled dinoflagellates. Stanford University Publications, Geological Sciences 15: 300.
- 58. Storey BC, Vaughan APM, Millar IL (1996) Geodynamic

evolution of the Antarctic Peninsula during Mesozoic times and its bearing on Weddell Sea history. Geological Society, London, Special Publications 108(1): 87-103.

- 59. Coffin MF, Rabinowitz PD (1988) Evolution of the conjugate East African- Madagascan margins and the western Somali Basin. Geol Soc Am Spec 225: 1-78.
- 60. McLoughlin S (2001) The breakup history of Gondwana and its impact on pre-Cenozoic floristic provincialism. Aust J Bot 49(3): 271-300.
- 61. Peris D, Labandeira CC, Barro E, Delclo X, Rust J, et al. (2020) Generalist Pollen-Feeding Beetles d u r i n g the Mid-Cretaceous. iScience 23(3): 100913.
- 62. Zavada MS (2004) The earliest occurrence of angiosperms in Southern Africa. South-African Journal of Botany 70(4): 646-653.
- 63. Jansonius J, McGregor DC (2002) Palynology: Principles and Applications. American Association of Stratigraphic Palynologists Foundation, Dallas 2: 463-910.

