

Extracts of *Citrus Reticulata* (*Rutaceae*) Fruit Peels Accelerate the onset of Toxicity of *Cerastes Cerastes* venom in Albino Mice

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Research Article

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Abstract

To investigate the effect of aqueous and methanolic extracts of *Citrus reticulata* (*Rutaceae*) Fruit Peels on *Cerastes cerastes* venom. The decline in the mean survival time of the male albino swiss mice were used to deduce the venom property in the presence and absence of aqueous and methanolic extracts of *Citrus reticulata* (*Rutaceae*) fruit peels. The aqueous and methanolic extracts of *Citrus reticulata* (*Rutaceae*) fruit Peels significantly decrease the mean survival time compared to the venom alone. From these results it was evident that the toxicity of *Cerastes cerastes* venomis increased significantly in the presence of *Citrus reticulata* in a dose dependent manner.

Keywords: *Citrus Reticulata*; *Cerastes Cerastes*; Venom; Toxicity

Introduction

Snakebites are severe socio-medical difficulty that lead to morbid and mortal impact on victims, and in Libya and other North African countries [1,2]. Immediate antivenom treatment is crucial and vital to avoid morbidity and mortality [3]. The oxidative stress status, which result from snake bite envenomation is another measurement of kidney impairment and acute renal failure, connected with the antioxidant defense system that might be subject for treatment by antioxidant therapy [4,5]. Reactive oxygen species (ROS) are involved in the inflammatory reactions,

thus affecting the cellular physiology and play an important role in the pathological conditions [6]. These free radical; ROS are involved in harming cellular components, and they play an important function in venom induced toxicity, as reported among envenomed mice [7]. Ascorbic acid is an antioxidant that has been reported to have useful effects on a number of types of cancer and could be concerned in alleviation of ROS cellular damage, produced during metabolism and exposure to toxins and carcinogens, in addition to augmentation of protease inhibitor effects concerned in preventing organ functional damage [8-12]. *Citrus reticulata* (*Rutaceae*) is commonly

known as narangi or santra (orange). It is a small spiny tree with thick top of slim branches, extensively grown in Egypt, Tunisia and Libya [13]. Mandarin is a collection name for this class of orange with thin, loose peel. The name 'tangerine' might be applied as an interchange name to the entire group, but in trade, it is usually limited to the types with red-orange skin. The fruit has aphrodisiac, laxative, tonic properties and astringent [14,15]. It is also used to alleviate vomiting [16,17]. The fruit peel controls the skin moisture, rough and softens hard skin and possess a cleaning effect on oily skin [18]. Chemical composition of the volatile oil of the fruit peels of this species has been reported [19-23]. The effects of the volatile oil of *C. reticulata* has been studied against *Saccharomyces cerevisiae*, pathogenic fungi, *Paenibacillus larvae*, *Schistosoma mansoni*, *Aspergillus flavus*, and other microorganisms [24-30]. The volatile oil of *C. reticulata* also demonstrates the anticancer activity [31-33]. In this present study, our aim was to investigate the effects of *Citrus reticulata* (Rutaceae) fruit Peels extracts on the toxicity of *Cerastes cerastes* venom in albino mice.

Materials and Methods

Collection of Plant Material and Preparation of Aqueous Extract

The oranges were bought from a shop in Tripoli (February 2019). The *Citrus reticulata* was identified and authenticated by a botanist. Orange rinds were peeled off carefully with the help of a sharp razor blade. Each rind sample was cut into smaller pieces and 30g mass of the sample was taken. The sample was initially rinsed with distilled water. The fresh peels (30 g) were added to 30 ml hot distilled water. In addition, another 30 g of the fresh peels were macerated in cold 99% methanol for three hours. After 3 hours of maceration at room temperature (28°C), the mixture was then filtered under vacuum and the filtrate was stored at 4°C and used to treat animals as needed [34].

Experimental Models

Swiss albino mice of either sex weighing about 18–28 g (2–2.6-month-old) used for experimental purpose. They were housed in polypropylene cages in the air-conditioned room with the temperature maintained at $25 \pm 2^\circ\text{C}$, and 12 h alternating light and dark cycles. The mice were provided with a nutritionally adequate diet and drinking water *ad libitum* throughout the study. Approval by the Animal Ethics Committee for the experimental procedures obtained.

Venoms

Snake (*Cerastes cerastes*) venom was extracted by manual stimulation and was obtained in liquid forms, from the Department of Zoology, Faculty of Science, University of Tripoli (Libya) and stored at -20°C until use. An aliquot of $7.5\mu\text{l}$ from the venoms was added to $800\mu\text{l}$ of normal saline. A dose of $100\mu\text{l}$ (100ng) was given to the male Swiss Albino mice.

Acute Toxicity Study

Acute toxicity was generally carried out for the determination of LD_{50} value in experimental animals. The aim of performing acute toxicity study is for establishing therapeutic index of a methanolic and aqueous extracts of *Citrus reticulata* and to ensure safety *in-vivo*. Acute toxicity test was performed in mice. All animals were fasted overnight before treatment and were given food one hour after aqueous and methanolic extracts. General behavior was also observed at 0.5, 1, 8, 12 and 24 h after administration. The number of animals that died after administration was traced daily for 7 days [35,36].

Intoxication of Venom by *Citrus Reticulata* Extracts

Five groups of mice were used in this study. The first group of six mice received only $100\mu\text{l}$ (100 ng of total protein) of the *Cerastes cerastes* venom ($\text{LD}_{99} 5\mu\text{g}/\text{kg}$). Groups 2-4 of six mice each (serving as treatment groups) were given an equivalent amount of the *Cerastes cerastes* venom with $50\mu\text{l}$, $100\mu\text{l}$ and $200\mu\text{l}$ of aqueous *Citrus reticulata* extracts intraperitoneally (30 g/30 ml), respectively. Group 5 of six mice received $100\mu\text{l}$ of the *Cerastes cerastes* venom and ASV. The number of mortality was recorded within 24h. Similar experiments were repeated in the same manner with the methanolic extract.

Statistical Analysis

The difference among various treated groups and control group were analyzed using one-way-ANOVA followed using unpaired Student's test. The results were expressed as the mean \pm SEM of the number of experiments done, with $P < 0.05$ indicating significant difference between groups.

Results and Discussion

Acute Toxicity Study

With the growing amount of research about naringin as a component of the orange and its potential utilize within the pharmacological and food industries, illuminating its toxicological outline becomes increasingly significant. In

the present study, the *Citrus reticulata* extracts were found to be safe up to 200 mg/kg orally. This present study is compared with other previous studies in which an oral single dose of 16 g/kg of naringin did not produce acute oral toxicity in rats [37].

Acute Toxicity of *Cerastes Cerastes* Venom and its Reaction with Aqueous (Methanolic) *Citrus Reticulata* Extracts and Antivenom

The *Cerastes cerastes* venom at the dose 5 µg/kg (LD₉₉) produces 100% mortality in mice. The aqueous (methanolic) *Citrus reticulata* extracts significantly decrease the mean survival times by 3, 5 and 6 times for 50, 100 and 200 µl (30g /30 mL), respectively when compared with the venom alone which was 3.1 ±0.3hours. ASV [polyvalent anti-snake venom by Haffkine Bio-Pharmaceuticals Company (India)] was found to be effective and showing mean survival of two days for five mice and complete survival of one mouse. The toxins of *Cerastes cerastes* venom are composed of neurotoxin, cardiotoxin, enzymes and proteins. The victim might die from respiratory paralysis which is the major cause of death. ASV and assisted ventilation can save life in many cases [38-40].

It has been reported that the Citrus species contain flavonones and glycosides in large amounts, and they play a major role in maintaining a range of pathological conditions. Hesperidin and naringein, are the major constituents of the citrus fruits. Naringin gets converted into naringenin which is an aglycone part by the intestinal microorganism. They established to have metal chelating effect, antioxidant, antidiabetic, antiviral, antiallergic, antiestrogenic, antimicrobial, ischemic heart disease adipolytic activity, anti-inflammatory, antiobesity, hypoxia, anti-cancer and activity hepatoprotective activity. Because of all these pharmacological action, both naringin and naringenin are assumed to be importance food supplement [41-47].

The accelerated death could be related to the interactions of *Citrus reticulata* components (which were mainly polyphenolic components) with snake venom which is not consistent with the previous studies reporting that polyphenolic secondary metabolites are able to inhibit PLA₂ [48]. In the literature, it has been reported that naringin which is a flavonoid that is found in grapefruit and known for its various pharmacological effects and biochemical activities of a secretory phospholipase A (sPLA₂) from *Crotalus durissus cascavella*, an imperative protein involved in the releasing of arachidonic acid in phospholipid membranes [48]. sPLA₂ was incubated with

naringin in a ratio of 1:1 mole at 37°C and a distinct decrease in the UV absorption signal and a changes of the circular dichroism spectra suggesting a significant effect of PLA₂ structure and function [48]. The obtained results are for the whole extract of *Citrus reticulata* and not for naringin or naringenin and this could be explained for the lack of association between pharmacological and enzymatic activities in which the chemical modification of some amino acids induced by naringin, in particular aromatic amino acids and histidines, affected the toxin's ability to interact with the pharmacological receptor, but did not lead to eliminate of this function. Our results and those described by Cardoso et al. [49] expressed that enzymatic activity of sPLA₂ is not crucial for pharmacological activities of this sPLA₂ which was isolated from *C. d. cascavella* venom [49].

Conclusion

The present study demonstrated that the aqueous extract of peeled *Citrus reticulata* possess dose-dependent toxic activity. Further, there is need to isolate, characterize, and screen the active principles that are responsible for its toxic activity. Furthermore, there is need to find out the exact mechanism by which the *Citrus reticulata* extract exerts above effects. Further studies are needed to separate and confirm the active components and its effect as a toxic agent with the venom.

References

1. Tianyi FL, Agbor VN, Tochie JN, Kadia BM, Nkwescheu AS (2018) Community-based audits of snake envenomations in a resource-challenged setting of Cameroon: case series. BMC Res Notes 11(1): 317.
2. Goncalves DV, Martinez-Freiria F, Crochet PA, Geniez P, Carranza S, et al. (2018) The role of climatic cycles and trans-Saharan migration corridors in species diversification: Biogeography of Psammophis schokari group in North Africa. Mol Phylogenet Evol 118: 64-74.
3. Lavonas EJ, Tomaszewski CA, Ford MD, Rouse AM, Kerns WP (2002) Severe puff adder (*Bitis arietans*) envenomation with coagulopathy. J Toxicol Clin Toxicol 40(7): 911-918.
4. Yamasaki SC, Villarroel JS, Barone JM, Zambotti-Villela L, Silveira PF (2008) Aminopeptidase activities, oxidative stress and renal function in *Crotalus durissus terrificus* envenomation in mice. Toxicon 52(3): 445-454.

5. Al Asmari AK, Khan HA, Manthiri RA, Al Yahya KM, Al Otaibi KE (2014) Effects of *Echis pyramidum* snake venom on hepatic and renal antioxidant enzymes and lipid peroxidation in rats. *J Biochem Mol Toxicol* 28(9): 407-412.
6. Carroll IM, Andrus JM, Bruno-Barcena JM, Klaenhammer TR, Hassan HM, et al. (2007) Anti-inflammatory properties of *Lactobacillus gasseri* expressing manganese superoxide dismutase using the interleukin 10-deficient mouse model of colitis. *Am J Physiol Gastrointest Liver Physiol* 293(4): G729-G738.
7. Douset E, Carrega L, Steinberg JG, Clot-Faybesse O, Jouirou B, et al. (2005) Evidence that free radical generation occurs during scorpion envenomation. *Comp Biochem Physiol C Toxicol Pharmacol* 140(2): 221-226.
8. Mirmohammadsadeghi M, Mirmohammadsadeghi A, Mahmoudian M (2018) Preventive Use of Ascorbic Acid For Atrial Fibrillation After Coronary Artery Bypass Graft Surgery. *Heart Surg Forum* 21(5): E415-E417.
9. Kitahata K, Matsuo K, Hara Y, Naganuma T, Oiso N, et al. (2018) Ascorbic acid derivative DDH-1 ameliorates psoriasis-like skin lesions in mice by suppressing inflammatory cytokine expression. *J Pharmacol Sci* 138(4): 284-288.
10. Banerjee P, Bhattacharyya SS, Bhattacharjee N, Pathak S, Boujedaini N, et al. (2009) Ascorbic acid combats arsenic-induced oxidative stress in mice liver. *Ecotoxicol Environ Saf* 72(2): 639-649.
11. Choudhury M, Senthilvadivel V, Velmurugan D (2018) Inhibitory effects of ascorbic acid toward snake venom metalloproteinase (SVMP) from Indian *Echis carinatus* venom: Insights from molecular modeling and binding studies. *J Biochem Mol Toxicol* 32(12): e22224.
12. Shamsi TN, Parveen R, Afreen S, Azam M, Sen P, et al. (2018) Trypsin Inhibitors from *Cajanus cajan* and *Phaseolus limensis* Possess Antioxidant, Anti-Inflammatory, and Antibacterial Activity. *J Diet Suppl* 15(6): 939-950.
13. Sharif SI, Ali BH (1994) Effect of grapefruit juice on drug metabolism in rats. *Food Chem Toxicol* 32(12): 1169-1171.
14. Tseng SH, Lee HH, Chen LG, Wu CH, Wang CC (2006) Effects of three purgative decoctions on inflammatory mediators. *J Ethnopharmacol* 105(1-2): 118-124.
15. McGuire RG, Hagenmaier RD (2001) Shellac formulations to reduce epiphytic survival of coliform bacteria on citrus fruit postharvest. *J Food Prot* 64(11): 1756-1760.
16. Yin OQ, Gallagher N, Li A, Zhou W, Harrell R, et al. (2010) Effect of grapefruit juice on the pharmacokinetics of nilotinib in healthy participants. *J Clin Pharmacol* 50(2): 188-194.
17. Glasscock SG, Friman PC, O'Brien S, Christophersen ER (1986) Varied citrus treatment of ruminant gagging in a teenager with Batten's disease. *J Behav Ther Exp Psychiatry* 17(2): 129-133.
18. Khan MA, Ali M, Alam P (2010) Phytochemical investigation of the fruit peels of *Citrus reticulata* Blanco. *Nat Prod Res* 24(7): 610-620.
19. Fayek NM, Farag MA, Abdel Monem AR, Moussa MY, Abd-Elwahab SM, et al. (2019) Comparative Metabolite Profiling of Four Citrus Peel Cultivars via Ultra-Performance Liquid Chromatography Coupled with Quadrupole-Time-of-Flight-Mass Spectrometry and Multivariate Data Analyses. *J Chromatogr Sci* 57(4): 349-360.
20. Guo Q, Liu K, Deng W, Zhong B, Yang W, et al. (2018) Chemical composition and antimicrobial activity of Gannan navel orange (*Citrus sinensis* Osbeck cv. Newhall) peel essential oils. *Food Sci Nutr* 6(6): 1431-1437.
21. Xu C, Zhang S, Zhang Y, Pu Y, Yin L, et al. (2018) Determination of spirotetramat and its four metabolites in citrus by ultra-high performance liquid chromatography-triple quadrupole-ion trap mass spectrometry. *Se Pu* 36(4): 339-344.
22. Kealey KS, Kinsella JE (1978) Orange juice quality with an emphasis on flavor components. *CRC Crit Rev Food Sci Nutr* 11(1): 1-40.
23. Dunlap WJ, Wender SH (1960) Purification and identification of flavanone glycosides in the peel of the sweet orange. *Arch Biochem Biophys* 87: 228-231.
24. Singh P, Shukla R, Kumar A, Prakash B, Singh S, et al. (2010) Effect of *Citrus reticulata* and *Cymbopogon citratus* essential oils on *Aspergillus flavus* growth and aflatoxin production on *Asparagus racemosus*. *Mycopathologia* 170(3): 195-202.
25. Lemes RS, Alves CCF, Estevam EBB, Santiago MB, Martins CHG, et al. (2018) Chemical composition and

- antibacterial activity of essential oils from *Citrus aurantifolia* leaves and fruit peel against oral pathogenic bacteria. *An Acad Bras Cienc* 90(2): 1285-1292.
26. Uckoo RM, Jayaprakasha GK, Vikram A, Patil BS (2015) Polymethoxy flavones Isolated from the Peel of Miaray Mandarin (*Citrus miaray*) Have Biofilm Inhibitory Activity in *Vibrio harveyi*. *J Agric Food Chem* 63(32): 7180-7189.
 27. Mehmood B, Dar KK, Ali S, Awan UA, Nayyer AQ, et al. (2015) Short communication: in vitro assessment of antioxidant, antibacterial and phytochemical analysis of peel of *Citrus sinensis*. *Pak J Pharm Sci* 28(1): 231-239.
 28. Mahadwar G, Chauhan KR, Bhagavathy GV, Murphy C, Smith AD, et al. (2015) Swarm motility of *Salmonella enterica* serovar Typhimurium is inhibited by compounds from fruit peel extracts. *Lett Appl Microbiol* 60(4): 334-340.
 29. Rakholiya K, Kaneria M, Chanda S (2014) Inhibition of microbial pathogens using fruit and vegetable peel extracts. *Int J Food Sci Nutr* 65(6): 733-739.
 30. Min KY, Kim HJ, Lee KA, Kim KT, Paik HD (2014) Antimicrobial activity of acid-hydrolyzed Citrus unshiu peel extract in milk. *J Dairy Sci* 97(4): 1955-1960.
 31. Nair SA, Sr RK, Nair AS, Baby S (2018) Citrus peels prevent cancer. *Phytomedicine* 50: 231-237.
 32. Arora S, Mohanpuria P, Sidhu GS, Yadav IS, Kumari V (2018) Cloning and Characterization of Limonoid Glucosyltransferase from Kinnow Mandarin (*Citrus reticulata* Blanco). *Food Technol Biotechnol* 56(2): 228-237.
 33. Tahsin T, Wansi JD, Al Groshi A, Evans A, Nahar L, et al. (2017) Cytotoxic Properties of the Stem Bark of *Citrus reticulata* Blanco (Rutaceae). *Phytother Res* 31(8): 1215-1219.
 34. Gray AM, Flatt PR (1999) Insulin-releasing and insulin-like activity of the traditional anti-diabetic plant *Coriandrum sativum* (coriander). *Br J Nutr* 81(3): 203-209.
 35. Ni H, Peng L, Gao X, Ji H, Ma J, et al. (2019) Effects of maduramicin on adult zebrafish (*Danio rerio*): Acute toxicity, tissue damage and oxidative stress. *Ecotoxicol Environ Saf* 168: 249-259.
 36. Wheeler MW (2018) Bayesian additive adaptive basis tensor product models for modeling high dimensional surfaces: an application to high-throughput toxicity testing. *Biometrics*.
 37. Li P, Wang S, Guan X, Cen X, Hu C, et al. (2014) Six months chronic toxicological evaluation of naringin in Sprague-Dawley rats. *Food Chem Toxicol* 66: 65-75.
 38. Abdel-Aty AM, Salama WH, Ali AA, Mohamed SA (2019) A hemorrhagic metalloprotease of Egyptian *Cerastes vipera* venom: Biochemical and immunological properties. *Int J Biol Macromol* 130: 695-704
 39. Ozverel CS, Damm M, Hempel BF, Gocmen B, Sroka R, et al. (2019) Investigating the cytotoxic effects of the venom proteome of two species of the Viperidae family (*Cerastes cerastes* and *Cryptelytrops purpureomaculatus*) from various habitats. *Comp Biochem Physiol C Toxicol Pharmacol* 220: 20-30.
 40. Lin CC, Wang PJ, Liu CC (2019) Venom concentrations in blisters and hemorrhagic bullae in a patient bitten by a Taiwan habu (*Protobothrops mucrosquamatus*). *Rev Soc Bras Med Trop* 52: e20180160.
 41. Abdel-Magied N, Shedid SM (2019) The effect of naringenin on the role of nuclear factor (erythroid-derived 2)-like2 (Nrf2) and haem oxygenase 1 (HO-1) in reducing the risk of oxidative stress-related radiotoxicity in the spleen of rats. *Environ Toxicol*.
 42. Xu C, Chen J, Zhang J, Hu X, Zhou X, et al. (2013) Naringenin inhibits angiotensin II-induced vascular smooth muscle cells proliferation and migration and decreases neointimal hyperplasia in balloon injured rat carotid arteries through suppressing oxidative stress. *Biol Pharm Bull* 36(10): 1549-1555.
 43. Hermenean A, Ardelean A, Stan M, Herman H, Mihali CV, et al. (2013) Protective effects of naringenin on carbon tetrachloride-induced acute nephrotoxicity in mouse kidney. *Chem Biol Interact* 205(2): 138-147.
 44. Annadurai T, Thomas PA, Geraldine P (2013) Ameliorative effect of naringenin on hyperglycemia-mediated inflammation in hepatic and pancreatic tissues of Wistar rats with streptozotocin-nicotinamide-induced experimental diabetes mellitus. *Free Radic Res* 47(10): 793-803.
 45. Tarun EI, Kurchenko VP, Metelitsa DI (2006) Flavonoids as effective protectors of urease from

- ultrasonic inactivation in solutions. *Bioorg Khim* 32(4): 391-398.
46. Kanno S, Tomizawa A, Ohtake T, Koiwai K, Ujibe M, et al. (2006) Naringenin-induced apoptosis via activation of NF-kappaB and necrosis involving the loss of ATP in human promyeloleukemia HL-60 cells. *Toxicol Lett* 166(2): 131-139.
47. Misty R, Martinez R, Ali H, Steimle PA (2006) Naringenin is a novel inhibitor of Dictyostelium cell proliferation and cell migration. *Biochem Biophys Res Commun* 345(1): 516-522.
48. Santos ML, Toyama DO, Oliveira SC, Cotrim CA, Diz-Filho EB, et al. (2011) Modulation of the pharmacological activities of secretory phospholipase A2 from *Crotalus durissus cascavella* induced by naringin. *Molecules* 16(1): 738-761.
49. Cardoso DF, Lopes-Ferreira M, Faquim-Mauro EL, Macedo MS, Farsky SH (2001) Role of crotoxin, a phospholipase A2 isolated from *Crotalus durissus terrificus* snake venom, on inflammatory and immune reactions. *Mediators Inflamm* 10(3): 125-133.

