



Plasmodium's Secret: How a Complex Endomembrane System Drives Malaria's Deadly Efficiency

Moumaris M*

Institute of Medical Sciences, Research and Development Company, France

*Corresponding author: Mohamed Moumaris, Research and Development Company, 14 avenue René Boylesve 75016 Paris, France, Tel: +33762122825; Email: mohamed.moumaris@sciencesettechnologies.com

Editorial

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Abbreviations

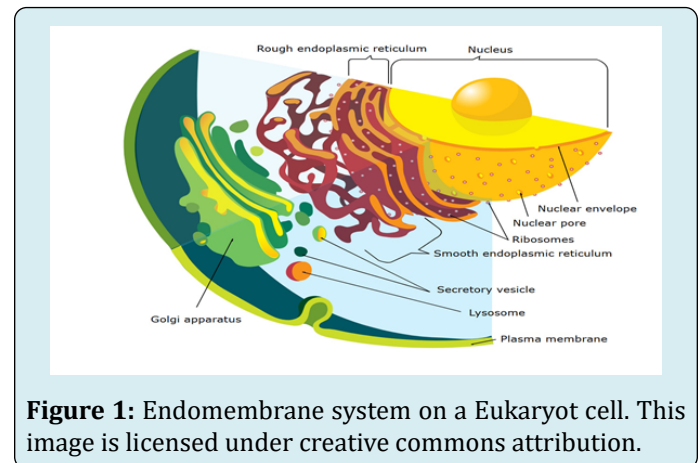
ER: Endoplasmic Reticulum; GRASP: Golgi Reassembly-Stacking Proteins; IMC: Internal Membrane Complex.

Editorial

Plasmodium, the parasite responsible for malaria, has a highly specialized endomembrane system to support its life cycle. Beyond typical organelles like the nucleus, endoplasmic reticulum (ER), and Golgi apparatus, it features unique compartments such as micronemes, rhoptries, dense granules, and the parasitophorous vacuole, crucial for invading host cells and evading the immune system. As the parasite progresses through its stages, it undergoes significant cellular changes, particularly in its ER and Golgi, to meet increasing metabolic demands and export proteins for host cell modification. Specialized organelles like rhoptries and micronemes facilitate host cell entry, while the actomyosin motor system drives its mobility. These adaptations allow *Plasmodium* to survive within red blood cells, making these mechanisms promising targets for antimalarial therapies to disrupt the parasite's life cycle and reduce malaria transmission.

The endomembrane system in eukaryotic cells coordinates essential functions through a network of organelles, including the nucleus, ER, Golgi apparatus, lysosomes, vacuoles, vesicles, and endosomes. In *Plasmodium*, the parasite responsible for malaria, this system

is highly specialized to adapt to its life cycle. Additional compartments- micronemes, rhoptries, dense granules, and the parasitophorous vacuole- aid in host cell invasion, survival, and immune evasion. The parasite's apicoplast and mitochondria also support crucial metabolic functions. As *Plasmodium* transitions between mosquito and human hosts, these organelles undergo structural adaptations to meet changing metabolic needs, illustrating the tight link between cellular structure and parasitic adaptability (Figure 1) [1-4].



The parasite undergoes significant cellular changes throughout its life cycle to adapt to and survive within host erythrocytes. Initially, in the merozoite phase, its ER is underdeveloped, reflecting low protein synthesis needs as it focuses on invading host cells. As it progresses to the ring stage, the ER forms, supporting increased metabolic demands. By the trophozoite stage, the developed ER enables the export of proteins that remodel the host erythrocyte's structure, enhance nutrient uptake, and help the parasite evade immune detection. These exported proteins use targeting sequences to reach specific destinations, ensuring



effective host cell modification. The parasite's nucleus, surrounded by a dual-layered nuclear envelope with ribosome-studded and fibrous membranes, uses nuclear pores to regulate molecular exchange and control gene expression. This gene regulation allows the parasite to adapt to changes within the host cell environment. Overall, the parasite's ability to develop complex organelle structures, remodel host cells, and precisely control protein targeting and gene expression showcases its advanced strategies for survival and replication. These mechanisms highlight potential therapeutic targets for disrupting the parasite's life cycle and reducing its impact on the host (Figure 2) [5-8].

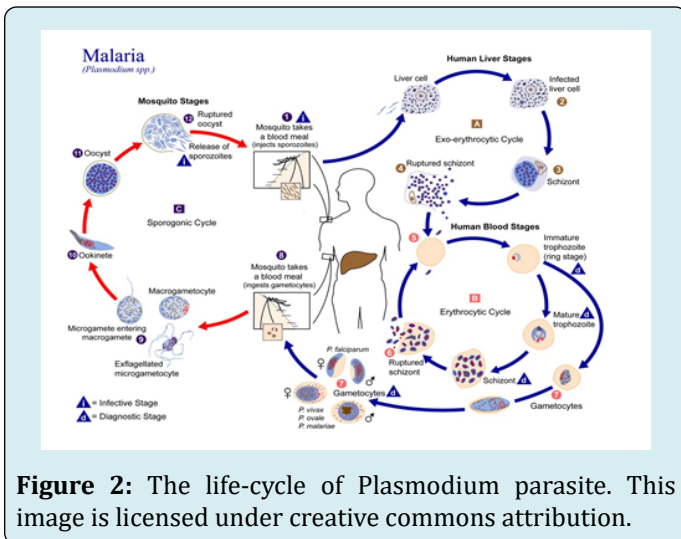


Figure 2: The life-cycle of *Plasmodium* parasite. This image is licensed under creative commons attribution.

The Golgi apparatus in *Plasmodium*, the malaria parasite, is vital for its survival and ability to infect human hosts. It is essential in lipid synthesis, protein modifications, and sorting for the parasite's development and red blood cell invasion. During its life cycle, *Plasmodium* relies on the Golgi to produce glycoproteins, especially in the early stages like the ring and trophozoite, but they are absent in the merozoite stage. The Golgi's structure changes across stages, from tubular in the ring stage to vesicular in the early trophozoite stage and distinct compartments in the schizont stage. These three compartments (Cis, Intermediate, and Trans) receive process and sort proteins for delivery to key destinations, such as the plasma membrane or secretory organelles that help in red blood cell invasion [9-11].

Golgi reassembly-stacking proteins (GRASP) are crucial for maintaining the structure and compartmentalization of *Plasmodium's* Golgi. These proteins, anchored to the membrane, ensure efficient protein processing. The Golgi also plays a vital role in secreting effector proteins that help the parasite manipulate host cells, evade immune responses, and create an environment conducive to replication, contributing to malaria severity. Given its importance in the parasite's lifecycle, the Golgi is a promising target for

antimalarial therapies. Disrupting its function or targeting GRASP proteins could impair the parasite's ability to invade and replicate, offering a potential strategy for malaria treatment [12,13].

Plasmodium merozoites possess a multi-layered pellicle composed of a plasma membrane and an internal membrane complex (IMC) linked to microtubules. This structure enables gliding mobility for invading red blood cells. The apical complex at the merozoite's tip includes secretory organelles like rhoptries, micronemes, and dense granules. Rhoptries secrete enzymes that form a tight junction with the host cells, aiding entry. While micronemes provide adhesion molecules for attachment, and dense granules release proteins to modify the host cell post-invasion. The merozoite's movement relies on the actomyosin motor system, where interactions between actin filaments and myosin generate the force needed for gliding mobility. This system is anchored to the IMC and powered by ATP, allowing the parasite to navigate host tissues. During the schizont stage, the Golgi apparatus processes vesicles to form rhoptries and micronemes. Rhoptries, characterized by a long neck and bulbous base, play a central role in altering host cell membranes to facilitate parasite entry. The coordinated function of structural elements, mobility mechanisms, and secretory systems enables *Plasmodium* to invade host cells. Understanding these mechanisms provides potential targets for antimalarial therapies to disrupt the parasite's life cycle, thereby reducing malaria transmission and infection severity (Figure 3) [14-17].

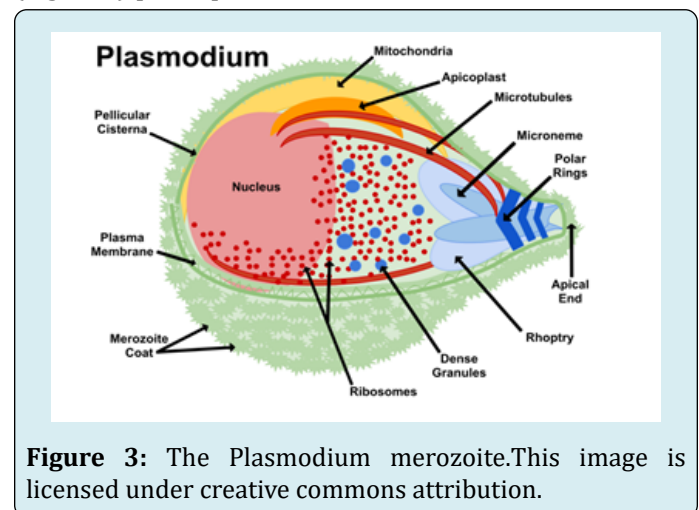


Figure 3: The *Plasmodium* merozoite. This image is licensed under creative commons attribution.

The specialized endomembrane system of *Plasmodium* supports its ability to adapt, survive, and thrive throughout its life cycle. By leveraging organelles like the ER, Golgi apparatus, apicoplast, and secretory compartments such as micronemes and rhoptries, the parasite efficiently invades host cells, evades immune defenses, and meets its metabolic needs. The dynamic structural adaptations of these organelles

tailored to various environmental and host conditions underscore *Plasmodium's* remarkable cellular complexity and resilience. Understanding these adaptative mechanisms deepens our knowledge of the parasite's biology and helps us find new antimalarial therapies. Disrupting key processes like protein export, Golgi function, or host cell remodeling offers promising avenues for reducing malaria's impact and combating drug resistance. As ongoing research unravels the intricate details of *Plasmodium's* cellular machinery, these insights pave the way for innovative strategies to control and eradicate malaria [18-46].

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