

The Occurrence and Diversity of Intestinal Digenetic Trematodes in Stray Cats, *Felis Catus*, In Egypt

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Research Article

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Abstract

Stray cats act as good indicators of fish-borne trematodes in the environment. Large populations of stray cats, *Felis catus*, are widespread in Giza, Egypt. Therefore, a total of 47 stray cats from some urban districts in Giza, Egypt were necropsied during the period from December 2018 to April 2019 and their intestines were examined for the detection of digenetic trematodes. Six cats were parasitized with 7 species of trematodes with an overall prevalence of 12.77%. Recovered species were belonging to heterophydes (*Heterophyes heterophyes, Pygidiopsis summa, Procerovum varium, Ascocotyle* sp.), cyathocotylids (*Prohemistomum vivax* and *Mesostephanus appendiculatus*), and echinostomatid (*Echinochasmus liliputanus*) in prevalences of 6.38% (3/47), 2.13% (1/47), and 6.38% (3/47), respectively. Mixed infections with heterophyds were noticed in two infected cats. Meanwhile, a mixed infection with *H. heterophyes* and the echinostomatid was recorded in one cat. Moreover, all trematode-infected cats had concomitant infections with cestode and/or nematode parasites. Therefore, veterinarians and authorized agencies must consider great hygienic measures and sanitary control programs to avoid the potential risk and zoonotic importance of such parasites.

Keywords: Stray Cats; Trematodes; Urban Areas; Giza; Egypt

Introduction

Stray cats, *Felis catus*, are strong predators having the ability to consume a variety of prey hosts including mammals, aves and reptiles. Thus, they act as potential reservoir hosts of a wide range of helminth parasites, particularly digenetic trematodes, reflecting both veterinary and medical importance [1-3]. Populations of stray cats have a random distribution and they live freely in both urban and rural districts, therefore, with a higher opportunity to disseminate infective eggs to the environment [4-9].

Investigations on digeneans-infected cats have been done

globally [10-19]. Moreover, several species of echinostomatids and heterophyids, including *Heterophyopsis continua*, *Pygidiopsis summa* and *Heterophyes nocens Echinostoma revolutum, Echinostoma hortense, Echinochasmus japonicus* were recorded in stray cats in the Korean Peninsula [2,20-22]. In Arabian countries, Schuster, et al. [23] detected *Heterophyes heterophyes* and *Heterophyopsis continua* in Dubai, United Arab Emirates. In Kuwait, for the first time, El-Azazy, et al. [24] revealed that 24.6% of cats infected with 14 species of trematodes, predominantly *H. heterophyes* and *H. dispar, Heterophyes nocens Haplorchis taichui, Echinochasmus japonicus, Mesostephanus appendiculatus, Haplorchis yokogawai*, and *Pygidiopsis genata*. In Egypt, few studies on digenean trematodes infecting cats have been performed previously. Among those, Arafa, et al. [25] reported *Heterophyes heterophyes, Echinochasmus perfoliatus* and *Euparyphium melis* in Giza and Cairo provinces. Other species of echinostomatids and heterophyids were recorded in Egyptian literature during the period from 1981 to 2011 [18,26-28].

The current study aimed to determine the prevalence of intestinal digenean trematodes recovered from stray cats, *Felis catus*, necropsied in randomly selected urban districts in Giza province, Egypt with morphological description of the revealed flukes.

Materials and Methods

Sampling Area and Animals

Forty seven stray cats, of both sexes, were obtained from randomly selected urban areas of Giza province (coordinates: 29° 15′ 36″ N, 29° 40′ 12″ E), Egypt during the period from December 2018 to April 2019. Based on dentition, cats aged either less than 3 years or more than 3 years.

Necropsy of Cats

During necropsy, the abdominal cavity was incised and the entire intestinal tracts of all carcasses were cut, tightly ligated with fine gauze. The small intestine, in particular, was longitudinally opened with a fine scissor and washed in 0.85% saline until the supernatant was cleared. The mucosa was scraped with a scalpel. The epithelial scrapings were passed through 60-80 mesh wire sieves. The contents of the sieves were washed with tap water and sediments were allowed to settle down and carefully examined under a stereomicroscope [3,29,30].

Laboratory Procedure

The recovered trematodes were compressed between two slides/cover slips. The flukes were then fixed in 10% neutral buffered formalin, washed with distilled water, and stained with potassium alum carmine (5 min for smallsized helminths and up to 4 hours for large helminths). Dehydration occurs in ascending grades of ethyl alcohol (70,80,90,95, and 100%). Then, specimens were cleared in xylene, and mounted with Canada balsam [30,31].

Identification of Trematodes

Prepared slides were carefully examined under a light microscopy using different magnifications, and recovered helminths were identified according to keys given by Yamaguti, [32] and Soulsby EJL [29]. Available photographs were taken using a digital microscope (Leica microsystems, CH-9435 Heebrugg, Ec3, and Singapore). Measurements of the recovered digenean trematodes were in micrometers.

Results

In the current study, seven digenetic trematode species were obtained. The recovered trematodes belonged to heterophyids (*Heterophyes heterophyes, Pygidiopsis summa, Procerovum varium, Ascocotyle* sp.), cyathocotylids (*Prohemistomum vivax* and *Mesostephanus appendiculatus*), and echinostomatids (*Echinochasmus liliputanus*). It has been found that six (12.77%) cats were parasitized; 3 cats were infected with heterophyids, one cat was infected with cyathocotylids and 3 cats were parasitized with the revealed echinostomatid, *Echinochasmus liliputanus* (Table 1). Among infected cats, 2 individuals (one male and one female) aged 1-3 years and 4 ones (one male and 3 females) aged more than 3 years. Moreover, 4 (females) out of 6 infected cats coinfected with adult tapeworms.

Digenetic trematode	Infected cats (n= 47)	Prevalence	Worm burden
Heterophyes heterophyes	3 ^{*§}	6.38	1-10
Pygidiopsis summa	1*	2.13	One
Procerovum varium	1*	2.13	One
Ascocotyle longicollis	1*	2.13	2
Prohemistomum vivax	1**	2.13	2
Mesostephanus appendiculatus	1**	2.13	1-12
Echinochasmus liliputanus	3§	6.38	1-5

*One cat had mixed infections of heterophyids.

**One cat had a mixed infection of both *P. vivax* and *M. appendiculatus*.

[§]One cat had a mixed infection of *H. heterophyes* and *E. liliputanus*

Table 1: The prevalence and worm burden of digenetic trematodes recovered from the intestinal tracts of stray cats in Giza, Egypt.

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Species: *Heterophyes heterophyes* Diagnosis (Figures 1 a,b)



cats. a Adult *Heterophyes heterophyes*. Scale bar = 100 μ m. b Gonotyle (ventrogenital complex) (*arrow*) of adult *Heterophyes heterophyes* showing numerous rodlets (more than 70). Scale bar = 50 μ m. c Adult *Pygidiopsis summa*. Scale bar = 100 μ m. d Adult *Procerovum varium*. Scale bar = 100 μ m. e Adult *Ascocotyle longicollis*. Scale bar = 100 μ m.

Short pear-shaped fluke measured 0.8-1.1 x 0.25 mm. It was wider posteriorly. The oral sucker measured approximately 50 μ m in diameter. The oesophagus measured 0.2 mm long. The ventral sucker is situated immediately anterior to the middle. The genital sucker lies directly behind it and to one side and bears an incomplete circle of 70-80 small rods. The testes are oval and horizontal in position measured about 50 μ m.

Species: *Pygidiopsis summa* Diagnosis (Figure 1c)

Very small fluke measured 0.45 x 0.20 mm. The body is more or less pointed at the fore end and broad at the hind end. Adult are ventrally concave, pyriform-shaped with oral and ventral suckers as well as a genital apparatus. The oral sucker measured about 37.5 μ m. The oesophagus measured 57.0 μ m. The whole body surface is covered with numerous tegumental spines, and sensory papillae.

Species: Procerovum varium

Diagnosis (Figure 1d)

The body is small pear-shaped measured 0.35×0.20 mm with the greatest width at the posterior third of the body. The

cuticle is provided with fine spines particularly at the anterior half. The oral sucker is subterminal and measured 50.0 μ m. The prepharynx is very short. Pharynx is subglobular or elliptical-shaped. The oesophagus is short and measured 0.12 mm long. The ventral sucker is very small, located just behind the bifurcation of intestinal ceca, embedded in the ventrogenital sac and masked by several eggs. Intestinal ceca bifurcate at the level of anterior third and terminate at the posterior fourth of the fluke. The ovary is spherical/ovoid, measured 50.0 µm and located sagital to the ventral sucker. Two testes, each is subglobular-shaped, located at the middle of the hind body and measured 95.0 × 99.0 µm. Vitellaria are greatly follicular and located in the posterior testicular field at the posterior extremity. The uterus is filled with small yellowish eggs (measured $22.0 \times 11.0 \mu m$) and occupies the entire area of the hind body.

Species: Ascocotyle (Phagicola) longicollis Diagnosis (Figure 1e)

A small elongate conical-shaped fluke, measured approximately 0.60 x 0.18 mm. The oral sucker measured 37.5 x 50.0 μ m and bears approximately 18-20 hooklets arranged in 2 rows. The pharynx measured 25.0 x 27.5 μ m and leads into the conical prepharynx that is characteristic of this genus. The oesophagus measured 0.18 mm. The genital opening is just anterior to the ventral sucker which measured 30.0 x 37.5 μ m. The majority of the reproductive organs are located posterior to the ventral sucker. Paired testes are symmetrical and located at the posterior end of the body measuring 50.0 x 62.5 μ m. The egg is oval-shaped and measured 12.5 x 25.0 μ m.



Figure 2: Adult cyathocotylids recovered from necropsied cats. a: Adult *Prohemistomum vivax*. Scale bar = $200 \mu m$. b: Adult *Mesostephanus appendiculatus*. Scale bar = $200 \mu m$.

Species: *Prohemistomum vivax* Diagnosis (Figure 2a) The adult fluke measured 1.2 mm x 0.55 mm. The body is oval rather than elongate. The cuticle is covered with minute spines which gradually diminish in size towards the posterior end. A sub terminal and rounded oral sucker measured $70.0 \times 70.0 \mu$ m. The ventral sucker ($50 \times 50 \mu$ m) is weakly developed, oval-shaped and lies at the junction of the anterior third with the middle third of the body. Testes are ovoid-shaped and smooth and they lie posteriorly, obliquely tandem at the posterior third of the body. Ovoid ovary and lies between the two testes. Vitelline follicles are voluminous and filling the posterior half and arranged in an interrupted horse-shoe- shaped manner around the gonads and the upper half of the cirrus pouch. At the posterior end, there is a distinct cirrus pouch ($0.1 \times 0.35 \text{ mm}$) along with a narrow tubular vagina parallel to it.

Species: *Mesostephanus appendiculatus* Diagnosis (Figure 2b)

Linguiform-shaped fluke, measured 1.12×0.35 mm. The anterior part is blunt and the posterior part is cylindrical, wider and provided with protruded caudal appendage (measured 0.12 mm long). The oral sucker is subterminal and measured 50 µm. The pharynx is somewhat large, more or less close to the oral sucker and measured 50×40 µm. The oesophagus is partially covered by the pharynx and measured 0.10 mm. The intestinal bifurcation begins at the anterior fourth and the posterior part of them is completely hidden by vitelline glands. Testes are oval/ovoid, tandem at the posterior fourth of the body and measured 80 µm in diameter. Few large eggs which are yellowish to brown and located at the posterior part. The ovary measured 60 µm in diameter and located in between the 2 testes and masked by vitelline glands.



Figure 3: The revealed echinostomatid, *Echinochasmus liliputanus*. a: The whole adult worm. Scale bar= $100 \mu m$. b: The anterior end of the worm showing a head collar armed with unequally sized collar spines arranged in a single dorsally interrupted row. Scale bar = $50 \mu m$.

Species: *Echinochasmus liliputanus* Diagnosis (Figure 3)

A small, elongate and leaf-shaped fluke, with its extremities narrower than the middle of the body. Sharply pointed spines are confined to the anterior part of the cuticle. The length of the body measured 0.50-0.70 mm (average 0.60 mm). The maximum width (0.20 mm) was at the level of the testes. The head collar measured 112.5 x 50 μ m and armed with 24 unequally-sized collar spines arranged in a single dorsally interrupted row (each spine measured approximately 25.0 µm). The oral sucker is rounded-shaped, subterminal and measured 40.0 µm. The ventral sucker is approximately twice the size of the oral sucker measuring 75.0 µm in diameter. The oesophagus is short measuring 100 µm long. The two testes are tandem closely related in posterior half of the body measuring 60.0 µm wide. The fore testis is more or less larger than the hind one. The ovary is oval-shaped, anterior to the testes and measured 70.0 x 50.0 μm. Vitellaria are large follicles extended from the middle to the posterior extremity. Eggs are few (2-6), golden brown with a distinct knob and measured 30.0 x 14.0 µm.

Discussion

Felids, particularly cats, may harbor several species of digenetic trematodes. The majority of them are zoonotic with subsequent potential harms to humans, especially in low-income countries [25].

In Egypt, helminths, particularly trematodes, of stray cats are considerably existed due to the higher opportunities of the heterogenic life cycle of those trematodes, as cats (the final hosts) have a predatory behavior that permit other reptiles, mammals and birds, containing the infective stages, to be ingested [3,16,23,33].

Currently, the prevalence of digenetic trematodes was 12.77% (6/47) in Giza province. In Egypt, previous literature reported higher infection rates; Fouly EA, [27] found that the prevalence of flukes-induced infection was 42.0%, Thabit HTM, [28] who reported that the prevalence was 61.6% and El-Dakhly, et al. [3] who revealed that the prevalence was 12.9%. Similarly, in Kuwait, El-Azazy, et al. [24] recorded a prevalence of 24.6%. Authors suggest that the lower prevalence in the current study might be referred to the enhancing hygienic measures and strict governmental rules that permit the hygienic disposal of garbage, containing various remnants of infected fish, as well as the periodical elimination of stray cats, the definitive and intermediate hosts of digenetic trematodes.

The present work revealed 7 digenetic trematodes. In Egypt, Kuntz RE, et al. [34] recorded 14 digenetic trematodes, *Heterophyes heterophyes, Heterophyes aequalis, H. pumilio, H.*

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taichui, H. yokogawai, S. falcatus, Pygidiopsis genata, Phagicola longicollis, Phagicola ascolonga, Stictodora sawakinensis, Echinochasmus liliputanus, Stephanoprora denticulatoides, Mesostephanus appendiculatus, and Cynodiplostomum namrui, from domestic cats. They added that the presence of these flukes might be due to ingesting fishes infected with metacercariae. Therefore, stray cats in those areas are potential definitive hosts for various digenetic trematodes.

Heterophyids are often intestinal flukes of fish-eating mammals and their metacercariae encyst in tissues of fishes [35-37]. Herein, the most frequent digenetic trematodes belonged to the family Heterophyidae. Coinciding with such finding, El-Azazy, et al. [24] recorded H. heterophyes and H. dispar, suggesting that stray cats are good biomarkers of fish-borne digenetic trematodes. Moreover, Eom, et al. [20] revealed Heterophyes continua, H. nocens and Pygidiopsis summa from cats purchased at Jungang Market in Seoul, South Korea. Cort WW, et al. [38] mentioned the morphological characters of Egyptian heterophyids. The number of chitinous rodlets in the genital sucker (gonotyl) was 75-85 in Heterophyes heterophyes and 50-60 in H. heterophyes nocens, a characteristic feature for the two heterophyids. The close association between snails and brackish water fish, intermediate hosts for such heterophyids, explain the occurrence of those flukes. In terms of zoonosis, Chai, et al. [39] stated that heterophyids might induce erratic parasitism in humans, and their eggs could be found within inflammatory reactions of affected organs with further complications resulted from the release of eggs into the blood stream via the intestinal wall.

Among heterophyids, *Ascocotyle* (syn. *Phagicola*) species have unique morphological character; the anterior end of adults has a crown of double rows of spines (14-17) and a solid prolongation of the oral sucker [40]. A clear association between the existence of *A. longicollis*, fish and predators was observed [41]. *Pygidiopsis summa* is a heterophyid fluke having a rapid development inside the final hosts [39]. In the present investigation, the occurrence of *P. summa* indicated the existence of definitive hosts, stray cats, and intermediate hosts, fish.

Among cyathocotylids, *Mesostephanus appendiculatus* is a common fluke of major public health significance, thus, it is considered a zoonotic fish-borne digenetic trematode [42]. Such fluke requires more than one host; fish eating mammals/birds (definitive host), gastropods and fish (intermediate hosts) [43]. The feeding habits of stray cats as free living animals evidently suggest the occurrence of *M. appendiculatus*. Meanwhile, *Prohemistomum vivax*, is a cyathocotylid needs catfish (a predominant consumed fish in Egypt) as intermediate host [44]. Currently, adult *Prohemistomum vivax* was revealed from stray cats.

Echinochasmus liliputanus is an intestinal echinostmatid in canids, felids, and humans. Herein, it was detected in 6.45% of infected cats. On the other hand, Thabit, [28] and Fang, et al. [9] revealed a higher prevalence (28.3%). Drinking un boiled pond water infected with cercariae is the often route of human infection [45,46].

To minimize the risk of potential and zoonotic trematode infections, veterinarians, parasitologists and biologists are advised to carefully consider the importance of snails, reptiles and fish as eminent intermediate hosts for digenetic trematodes as well as the hygienic disposal of those hosts to reduce the possibility of completing life cycles of the flukes. Meanwhile, authoritive agencies are strongly recommended to eliminate strays cats and dogs.

Conflict of Interest

Authors declare that there is no conflict of interest.

Ethical Standards

Animals used in the present study were obtained as a result of the cooperation between the Directorate of Veterinary Medicine, Beni-Suef, Egypt and the Faculty of Veterinary Medicine, Beni-Suef. The current work was done according the ethics Animal and Human Research Committee of the Faculty of Veterinary Medicine, Beni-Suef University, Egypt.

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