

# Plasmacytoid Dendritic Cells: The Potential of Synergistic Crosstalk between Innate and Adaptive Immunity

**Tania Rahman**<sup>1,2\*</sup>

<sup>1</sup>Department of Biochemistry and Molecular Biology, Bio21 Molecular Science and Biotechnology Institute, University of Dhaka, Bangladesh

<sup>2</sup>Department of Microbiology and Immunology, University of Melbourne at the Peter Doherty Institute for Infection and Immunity, Australia

**\*Corresponding author:** Tania Rahman, Associate Professor, Department of Biochemistry and Molecular Biology, University of Dhaka, Bangladesh, Tel: +88-01917285696; Email: tania.rahman@du.ac.bd

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## Abstract

In rodents, a combination of innate and adaptive immune responses is critical in controlling infection. The adaptive immune components, comprising B cells, CD4 T cells, CD8 T cells, pathogen specific secretory antibodies, and Treg cells have been exhibited to play an indispensable role in comprising and eliminating various infections. Also, there are several innate immune components that help to control infections, such as soluble mediators, lymphotoxin (LT) signaling in innate epithelial cells, TLRs, signaling adaptor MyD88, inflammasomes, CXCL9, P-selectin glycoprotein ligand-1 (PSGL-1). Little is known about the molecules that mediate the crosstalk between innate and adaptive immune response during infection. The work presented here has unravelled the potential of plasmacytoid dendritic cells (pDC) in the immune system in providing the first line of defense during natural infection as well as subsequent adaptive host defense against pathogens. In this work the potential that pDC might have in infection has been investigated in order to understand the basis of the development of protective and pathological responses during infection. These findings would help set up future avenues of research to elucidate a key mechanism of action of these cells and provide new therapeutic insights.

**Keywords:** Type I IFN-producing Cells, CD4 T Cells, CD8 T Cell, Treg Cell, Th17 Response, NK Cell, Soluble Mediators, Secretory Antibodies

**Abbreviations:** LT: Lymphotoxin; PSGL-1: P-Selectin Glycoprotein Ligand-1; pDC: plasmacytoid Dendritic Cells; HIV: Human Immunodeficiency Virus; HSV: Herpes

Simplex Virus; MCMV: Murine Cytomegalo Virus; LCMV: Lymphocytic Choriomeningitis Virus.

## Introduction

pDC are a unique population of bone marrow derived leukocytes that are developmentally and functionally distinct from conventional DCs [1,2]. In the steady state, pDC reside primarily in the lymphoid organs and are recruited by infection and inflammatory mediators [2-4]. For instance, accumulations of pDC are prevalent in the skin in psoriasis [5] and SLE [6], in the mucosal tissues in allergic rhinitis [7], in the lungs in pneumonia [8] and respiratory syncytial virus [9], in the mucosal space in colon and mesenteric LN in IBDs [10], in vagina in genital HSV-2 infection [11]. pDC can differentiate into antigen-presenting dendritic cells as a result of their link to several cellular receptors that promptly detect nucleic acids in pathogens [2,12]. These powerful networks of molecular and cellular events permit pDC to bridge the innate and adaptive immune systems resulting in a concerted pathogen response.

## pDC during Infection

As the major type I IFN-producing cells in the innate immune system, pDC respond to a wide range of viruses, including human immunodeficiency virus type I (HIV-1), influenza virus, sendai virus, herpes simplex virus (HSV), murine cytomegalovirus (MCMV) and lymphocytic choriomeningitis virus (LCMV) etc [13]. However, their role in non-viral diseases was not much appreciated until recently a handful of studies have begun to explore the contribution of pDC to bacterial immune responses and autoimmunity. For instance, pDC produce IFN-I upon recognition of bacterial RNA [14,15] and when exposed to *Borrelia burgdorferi* in vitro [16], they can activate CD4 T cells via antigen presentation and IL12 secretion in response to toxoplasma infection [17]. Also, pDC can suppress induction of CD4 T cells and initiate CD8 T cell responses [18]. pDC can infiltrate the mesenteric LN and spleen of *Listeria*-infected mice [19], although they are not responsible for the elevated systemic IFN-I levels [20] that interfere with the clearance of *Listeria* [21]. pDC have also been shown to drive Th17 responses in patients with GVHD [22] and pDC produce LN homing of NK cells [23]. The potential of pDC to induce Treg has been illustrated in numerous disease models [24-27]. Depletion of pDC results in an upregulation of T cell immune responses, increased production of thymic regulatory T (Treg) cells, fewer Treg cells in the gut, decreased generation of induced Treg cells and an increased frequency Th17 cells [28]. Recently, pDC have been found to be associated with the development of central tolerance in the thymus [29]. This work supports earlier studies that suggest a tolerogenic role for pDC in some circumstances.

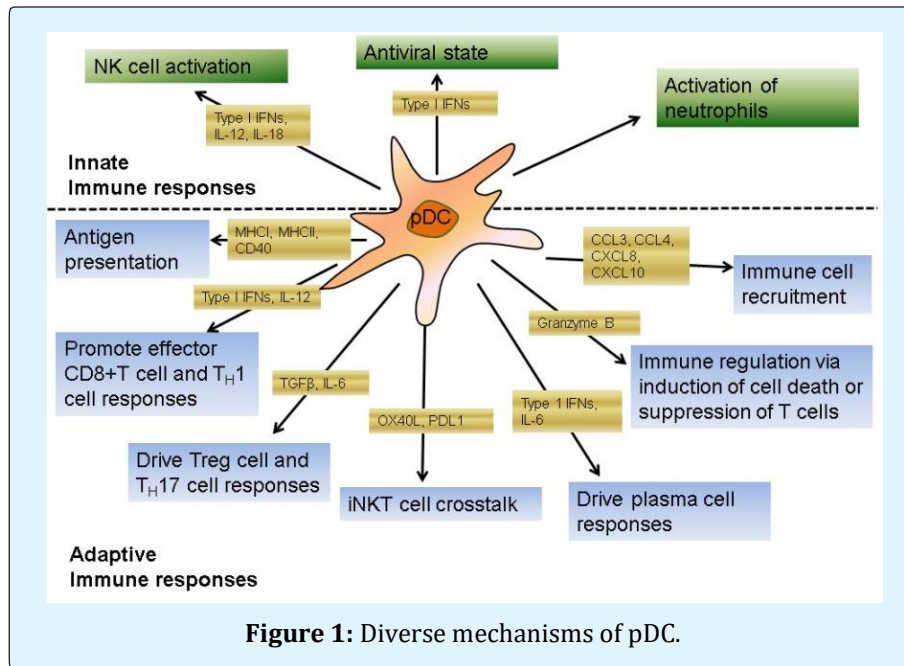
## pDC a Specialized Type I Interferon Secreting Cells

pDC, a distinctive subset of DCs is able to secrete type I IFNs more quickly than other cell types to a large array of viral and non-viral molecules [30-32]. Though comprising only 0.2–0.8% of human blood cells, pDC are able to produce more than 95% of type I IFNs by peripheral blood mononuclear cells following viral insult [33,34]. pDC are proficient in secreting up to 10 pg/cell of type I IFN, producing them 10–100-fold more than other cell types including mDC [35]. Within 6 h of activation, human pDC contribute 60% of their transcriptional activity to secrete type I IFNs, comprising entire transcripts of nineteen diverse subclasses of type I IFN [36]. pDC express a wide range of *ifna* genes compared to other professional antigen-presenting cells (APCs). Human pDC express the entire subtypes of type I IFNs including IFN- $\alpha$ , IFN- $\beta$ , IFN- $\gamma$ , IFN- $\omega$ , and IFN- $\tau$  [1]. Ablation of pDC in vivo abolished the IFN- $\alpha$  secretion in response to cytomegalovirus infection in mice [37,38].

## Role of pDC as a Linker of Innate and Adaptive Immunity

Type I IFNs produced by pDC induce a cellular antiviral response and recruits a network of molecular and cellular events and thus linking the innate with adaptive immunity [39] (Figure 1). Type I IFNs activate NK cell cytolytic activity, however, protect uninfected cells from NK cell-mediated lysis [40,41]. pDC induce production of IFN $\gamma$  in NK cells via IL-12 secretion [42]. Type I interferons released by pDC cause T cells to produce IL-10, IFN $\gamma$  and also induce a Th1 polarization and thus promote clearance of intracellular pathogens [43,44]. Type I interferons can upregulate early activation markers CD69 on T cells [45,46] and generation of long-term antitumor immune response [47]. Through the production of both type I interferons and IL-6, pDC promote human B cells to differentiate into antibody secreting plasma cells facilitating the production of antiviral antibodies [48,49]. Plasma cells induced by pDC predominantly secrete IgG rather than IgM, suggesting that pDC may trigger memory B cells [50,51]. Type I IFNs can increase cross-presentation during viral infection, and induce antigen-specific CD8+ T cells, and the differentiation of naive T cells into Th1 cells using the STAT1, STAT3 activation and T-bet expression [52-54]. Type I IFNs trigger immature cDCs to produce IL-12, IL-15, IL-18, and IL-23, cytokines that are not secreted by human pDC [55,56]. pDC are likewise competent producers of chemokines, mostly CCL3, which can aid

them to trigger additional leukocytes to inflamed tissues [57,58].



pDC are important drivers for both innate and adaptive immune responses. pDC production of MHC class I, Class II and costimulatory molecule CD40 enable pDCs to present antigen to CD4 cells. pDC production of type I IFNs and IL-12 enable pDCs to promote effector CD8 T cells as well as polarization of CD4 cells to Th1 responses. pDC production of TGF $\beta$  and IL-6 drive either Treg or Th17 cell responses. The crosstalk between pDCs and NKT cells occur via OX40L and PDL1 ligand. pDC production of type 1 IFNs and IL-6 drive plasma cell responses. Granzyme B enable pDCs to induce apoptosis or cell death as well as suppression of T cells. pDCs induce chemokines including CCL3, CCL4, CCL8 and CXCL10 which can attract other immune cells to the site of infection. pDC can activate neutrophils. pDC production of type 1 IFNs can produce an antiviral adaptive immune responses. pDC production of type 1 IFNs, IL-12 and IL-18 can activate NK cells.

### Concluding Remarks

The role of pDC has been exhibited in two distinct immune systems, one of which gives rise to the other. pDC, which are typically believed to be as antiviral, enact a central role in host control of bacterial infections. The findings in this study can be extended to other undiagnosed bacterial and parasite infections. Also, additional studies are required on pDC in different type of

infections to hypothesize extended knowledge on the vivid character of these cells and mechanisms contributing to immunity against various pathogens.

### References

1. Lande R, Gilliet M (2010) Plasmacytoid dendritic cells: key players in the initiation and regulation of immune responses. *Ann N Y Acad Sci* 1183: 89-103.
2. Villadangos JA, Young L (2008) Antigen-presentation properties of plasmacytoid dendritic cells. *Immunity* 29(3): 352-361.
3. Asselin-Paturel C, Boonstra A, Dalod M, Durand I, Yessaad N, et al. (2001) Mouse type I IFN-producing cells are immature APCs with plasmacytoid morphology. *Nat Immunol* 2(12): 1144-1150.
4. Sozzani S, Vermi W, Del Prete A, Facchetti F (2010) Trafficking properties of plasmacytoid dendritic cells in health and disease. *Trends Immunol* 31(7): 270-277.
5. Glitzner E, Korosec A, Brunner PM, Drobits B, Amberg N, et al. (2014) Specific roles for dendritic cell subsets during initiation and progression of psoriasis. *EMBO Mol Med* 6(10): 1312-1327.

6. Ronnblom L (2011) The type I interferon system in the etiopathogenesis of autoimmune diseases. *Ups J Med Sci* 116(4): 227-237.
7. Jahnsen FL, Lund-Johansen F, Dunne JF, Farkas L, Haye R, et al. (2000) Experimentally induced recruitment of plasmacytoid (CD123high) dendritic cells in human nasal allergy. *J Immunol* 165(7): 4062-4068.
8. Ang DK, Oates CV, Schuelein R, Kelly M, Sansom FM, et al. (2010) Cutting edge: pulmonary *Legionella pneumophila* is controlled by plasmacytoid dendritic cells but not type I IFN. *J Immunol* 184(10): 5429-5433.
9. Wang H, Peters N, Schwarze J (2006) Plasmacytoid dendritic cells limit viral replication, pulmonary inflammation, and airway hyper responsiveness in respiratory syncytial virus infection. *J Immunol* 177(9): 6263-6270.
10. Baumgart DC, Metzke D, Guckelberger O, Pascher A, Grötzinger C, et al. (2011) Aberrant plasmacytoid dendritic cell distribution and function in patients with Crohn's disease and ulcerative colitis. *Clin Exp Immunol* 166(1): 46-54.
11. Lund JM, Linehan MM, Iijima N, Iwasaki A (2006) Cutting Edge: Plasmacytoid dendritic cells provide innate immune protection against mucosal viral infection in situ. *J Immunol* 177(11): 7510-7514.
12. Lombardi VC, Khaiboullina SF (2014) Plasmacytoid dendritic cells of the gut: relevance to immunity and pathology. *Clin Immunol* 153(1): 165-177.
13. Tang F, Du QM, Liu YJ (2010) Plasmacytoid dendritic cells in antiviral immunity and autoimmunity. *Sci China-Life Sci* 53(2): 172-182.
14. Chiu YH, Macmillan JB, Chen ZJ (2009) RNA polymerase III detects cytosolic DNA and induces type I interferons through the RIG-I pathway. *Cell* 138(3): 576-591.
15. Eberle F, Sirin M, Binder M, Dalpke AH (2009) Bacterial RNA is recognized by different sets of immunoreceptors. *Eur J Immunol* 39(9): 2537-2547.
16. Petzke MM, Brooks A, Krupna MA, Mordue D, Schwartz I (2009) Recognition of *Borrelia burgdorferi*, the Lyme disease spirochete, by TLR7 and TLR9 induces a type I IFN response by human immune cells. *J Immunol* 183(8): 5279-5292.
17. Pepper M, Dzierzinski F, Wilson E, Tait E, Fang Q, et al. (2008) Plasmacytoid dendritic cells are activated by *Toxoplasma gondii* to present antigen and produce cytokines. *J Immunology* 180(9): 6229-6236.
18. Takagi H, Fukaya T, Eizumi K, Sato Y, Sato K, et al. (2011) Plasmacytoid Dendritic Cells Are Crucial for the Initiation of Inflammation and T Cell Immunity in Vivo. *Immunity* 35(6): 958-971.
19. Tam MA, Wick MJ (2006) Differential expansion, activation and effector functions of conventional and plasmacytoid dendritic cells in mouse tissues transiently infected with *Listeria monocytogenes*. *Cell Microbiol* 8(7): 1172-1187.
20. Stockinger S, Kastner R, Kernbauer E, Pilz A, Westermayer S, et al. (2009) Characterization of the interferon-producing cell in mice infected with *Listeria monocytogenes*. *PLoS Pathog* 5(3): 1000355.
21. Rayamajhi M, Humann J, Penheiter K, Andreasen K, Lenz LL (2010) Induction of IFN- $\alpha$  enables *Listeria monocytogenes* to suppress macrophage activation by IFN- $\gamma$ . *J Exp Med* 207(2): 327-337.
22. Bossard C, Malard F, Arbez J, Chevallier P, Guillaume T, et al. (2012) Plasmacytoid dendritic cells and Th17 immune response contribution in gastrointestinal acute graft-versus-host disease. *Leukemia* 26(7): 1471-1474.
23. Romagnani C, Della Chiesa M, Kohler S, Moewes B, Radbruch A, et al. (2005) Activation of human NK cells by plasmacytoid dendritic cells and its modulation by CD4+ T helper cells and CD4+ CD25hi T regulatory cells. *Eur J Immunol* 35(8): 2452-2458.
24. De Heer HJ, Hammad H, Soullié T, Hijdra D, Vos N, et al. (2004) Essential role of lung plasmacytoid dendritic cells in preventing asthmatic reactions to harmless inhaled antigen. *J Exp Med* 200(1): 89-98.
25. Ochando JC, Homma C, Yang Y, Hidalgo A, Garin A, et al. (2006) Alloantigen-presenting plasmacytoid dendritic cells mediate tolerance to vascularized grafts. *Nat Immunol* 7(6): 652-662.
26. Goubier A, Dubois B, Gheit H, Joubert G, Villard-Truc F, et al. (2008) Plasmacytoid dendritic cells mediate oral tolerance. *Immunity* 29(3): 464-475.
27. Adalid-Peralta L, Fragoso G, Fleury A, Sciotto E (2011) Mechanisms underlying the induction of regulatory T cells and its relevance in the adaptive immune



- response in parasitic infections. *Int J Biol Sci* 7(9): 1412-1426.
28. Takagi H, Fukaya T, Eizumi K, Sato Y, Sato K, et al. (2011) Plasmacytoid dendritic cells are crucial for the initiation of inflammation and T cell immunity in vivo. *Immunity* 35(6): 958-971.
  29. Hadeiba H, Lahl K, Edalati A, Oderup C, Habtezion A, et al. (2012) Plasmacytoid dendritic cells transport peripheral antigens to the thymus to promote central tolerance. *Immunity* 36(3): 438-450.
  30. Schettini J, Mukherjee P (2008) Physiological role of plasmacytoid dendritic cells and their potential use in cancer immunity. *Clin Dev Immunol* 2008: 106321.
  31. Liu YJ (2005) IPC: professional type 1 interferon-producing cells and plasmacytoid dendritic cell precursors. *Annu Rev Immunol* 23: 275-306.
  32. Mathan TS, Figdor CG, Buschow SI (2013) Human plasmacytoid dendritic cells: from molecules to intercellular communication network. *Front Immunol* 4: 372.
  33. Fitzgerald-Bocarsly P, Feldman M, Mendelsohn M, Curl S, Lopez C (1988) Human mononuclear cells which produce interferon-alpha during NK(HSV-FS) assays are HLA-DR positive cells distinct from cytolytic natural killer effectors. *J Leukoc Biol* 43(4): 323-334.
  34. Fitzgerald-Bocarsly P, Dai J, Singh S (2008) Plasmacytoid dendritic cells and type I IFN: 50 years of convergent history. *Cytokine Growth Factor Rev* 19(1): 3-19.
  35. Ito T, Kanzler H, Duramad O, Cao W, Liu YJ (2006) Specialization, kinetics, and repertoire of type 1 interferon responses by human plasmacytoidpredendritic cells. *Blood* 107(6): 2423-2431.
  36. Siegal FP, Kadowaki N, Shodell M, Fitzgerald-Bocarsly PA, Shah K, et al. (1999) The nature of the principal type 1 interferon-producing cells in human blood. *Science* 284(5421): 1835-1837.
  37. Barchet W, Cella M, Odermatt B, Asselin-Paturel C, Colonna M, et al. (2002) Virus-induced interferon alpha production by a dendritic cell subset in the absence of feedback signaling in vivo. *J Exp Med* 195(4): 507-516.
  38. Reizis B, Bunin A, Ghosh HS, Lewis KL, Sisirak V, et al. (2011) Plasmacytoid Dendritic Cells: Recent Progress and Open Questions. *Annu Rev Immunol* 29: 163-183.
  39. Kadowaki N, Antonenko S, Lau JY, Liu YJ (2000) Natural interferon alpha/beta-producing cells link innate and adaptive immunity. *J Exp Med* 192(2): 219-226.
  40. Hanabuchi S, Watanabe N, Wang YH, Wang YH, Ito T, et al. (2006) Human plasmacytoid predendritic cells activate NK cells through glucocorticoid-induced tumor necrosis factor receptor-ligand (GITRL). *Blood* 107(9): 3617-3623.
  41. Bandyopadhyay S, Perussia B, Trinchieri G, Miller DS, Starr SE (1986) Requirement for HLA-DR+ accessory cells in natural killing of cytomegalovirus-infected fibroblasts. *J Exp Med* 164(1): 180-195.
  42. Colonna M, Trinchieri G, Liu YJ (2004) Plasmacytoid dendritic cells in immunity. *Nat Immunol* 5(12): 1219-1226.
  43. Cella M, Facchetti F, Lanzavecchia A, Colonna M (2000) Plasmacytoid dendritic cells activated by influenza virus and CD40L drive a potent TH1 polarization. *Nat Immunol* 1(4): 305-310.
  44. Visperas A, Do J, Min B (2014) Cellular factors targeting APCs to modulate adaptive T cell immunity. *J Immunol Res* 2014: 750374.
  45. Agnello D, Lankford CS, Bream J, Morinobu A, Gadina M, et al. (2003) Cytokines and transcription factors that regulate T helper cell differentiation: new players and new insights. *J Clin Immunol* 23(3): 147-161.
  46. Hemmi M, Tachibana M, Tsuzuki S, Shoji M, Sakurai F, et al. (2014) The early activation of CD8+ T cells is dependent on type I IFN signaling following intramuscular vaccination of adenovirus vector. *Biomed Res Int* 2014: 158128.
  47. Belardelli F, Gresser I (1996) The neglected role of type I interferon in the T-cell response: implications for its clinical use. *Immunol Today* 17(8): 369-372.
  48. Jego G, Palucka AK, Blanck JP, Chalouni C, Pascual V, et al. (2003) Plasmacytoid dendritic cells induce plasma cell differentiation through type I interferon and interleukin 6. *Immunity* 19(2): 225-234.

49. Gujer C, Sandgren KJ, Douagi I, Adams WC, Sundling C, et al. (2011) IFN-alpha produced by human plasmacytoid dendritic cells enhances T cell-dependent naive B cell differentiation. *J Leukoc Biol* 89(6): 811-821.
50. Poeck H, Wagner M, Battiany J, Rothenfusser S, Wellisch D, et al. (2004) Plasmacytoid dendritic cells, antigen, and CpG-C license human B cells for plasma cell differentiation and immunoglobulin production in the absence of T-cell help. *Blood* 103(8): 3058-3064.
51. Crampton SP, Voynova E, Bolland S (2010) Innate pathways to B-cell activation and tolerance. *Ann N Y Acad Sci* 1183: 58-68.
52. Kolumam GA, Thomas S, Thompson LJ, Sprent J, Murali-Krishna K (2005) Type I interferons act directly on CD8 T cells to allow clonal expansion and memory formation in response to viral infection. *J Exp Med* 202(5): 637-650.
53. Le Bon A, Etchart N, Rossmann C, Ashton M, Hou S, et al. (2003) Cross-priming of CD8+ T cells stimulated by virus-induced type I interferon. *Nat Immunol* 4(10): 1009-1015.
54. Tang F, Du Q, Liu YJ (2010) Plasmacytoid dendritic cells in antiviral immunity and autoimmunity. *Sci China Life Sci* 53(2): 172-182.
55. Santini SM, Lapenta C, Logozzi M, Parlato S, Spada M, et al. (2000) Type I interferon as a powerful adjuvant for monocyte-derived dendritic cell development and activity in vitro and in Hu-PBL-SCID mice. *J Exp Med* 191(10): 1777-1788.
56. Paquette RL, Hsu NC, Kiertscher SM, Park AN, Tran L, et al. (1998) Interferon-alpha and granulocyte-macrophage colony-stimulating factor differentiate peripheral blood monocytes into potent antigen-presenting cells. *J Leukoc Biol* 64(3): 358-367.
57. Krug A, Uppaluri R, Facchetti F, Dorner BG, Sheehan KC, et al. (2002) IFN-producing cells respond to CXCR3 ligands in the presence of CXCL12 and secrete inflammatory chemokines upon activation. *J Immunol* 169(11): 6079-6083.
58. Megjugorac NJ, Young HA, Amrute SB, Olshalsky SL, Fitzgerald-Bocarsly P (2004) Virally stimulated plasmacytoid dendritic cells produce chemokines and induce migration of T and NK cells. *J Leukoc Biol* 75(3): 504-514.

