

Growth of Different Olive Varieties Influences with Foliar Application of GA₃ under Saline Conditions

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Abstract

Salinity is an important factor affecting plant productivity and constitutes a problem concerning a significant portion of the earth planet. Olives are a good source of iron which helps to transport oxygen in blood. Calcium present in olives is essential for bones and muscles. Gibberellins (GA₃) play a vital role in the detoxification of heavy metals and in tolerance to salt stress by improving plants growth, chlorophyll synthesis and activities of antioxidant enzymes, and by preventing lipid per oxidation. The study was carried out at NARC Islamabad during August, 2018 to October, 2018 to examine olive varieties growth influences with foliar application of GA₃ under saline conditions in tunnel. Soil salinity was developed artificially with the mixture of different salts at 2.0dSm⁻¹. Completely randomized design was applied with three replications. Foliar spray of Gibberellic acid @ 0, 200 and 400 mg l⁻¹ was done. Growth parameters of plant height stem diameter, # of leaves plant⁻¹ and leaf area were recorded at the end of the experiment. Megaron olive variety attained the highest plant height at 200 and 400 mg l⁻¹ GA₃ foliar sprays than other two olive varieties. Stem diameter was the maximum in Coratina olive variety that was higher than control and other two olive varieties. Number of leaves plant⁻¹ and leaf area were gained the highest position by Chetoui olive variety at 200 and 400 mg l⁻¹ GA₃ foliar spray than other two olive varieties under artificially saline conditions.

Keywords: Gibberelic Acid; Growth Hormone; Olea Europaea; Coratina; Chetoui; Megaron

Introduction

Olive tree (*Olea europaea* L.) of the Oleaceae family has a high economic value and many countries such as Iran and Mediterranean countries use its oil and conserved fruits [1]. Olive is very well adapted to the high temperature; tolerate dry weather, high soil salinity levels and infertile soil. The size of the fruit is important, not

only because it is a component of productive yield, but also determines the acceptance by the consumer as conserved fruits. Olive tree (*Olea europaea* L.) of the Oleaceae family has a high economic value and many countries such as Iran and Mediterranean countries use its oil and conserved fruits [1]. Olive is very well adapted to the high temperature; tolerate dry weather, high soil salinity levels and infertile soil. Olives represents a group

of evergreen trees and shrubs distributed in warm temperate and tropical region of the world. Olive is a unique subtropical fruit crop which requires chilling for fruiting [2-4]. Olive is very important as an oil producing plant of the Mediterranean region and in areas having etesian or dry subtropical climate like Cuba, California, New Zealand and South East Australia. Its origin is probably in the eastern regions of the Mediterranean Sea, from where it has spread around the Basin [5,6]. Although the olive trees are moderately tolerant to salinity and significant differences in salt tolerance have been reported among cultivars [7]. The size of the fruit is important, not only because it is a component of productive yield, but also determines the acceptance by the consumer as conserved fruits. Gibberellins are known for their ability to increase cell enlargement, thus enhancing fruit growth in certain species such as citrus, litchi, guava, and pear [8-14]. In all species so far studied, gibberellins had the potential for increasing fruit size. The beneficial effects of Gibberellic acid (GA_3) and nutrient elements sprays specially zinc on yield and fruit quality of different fruit crops were mentioned by many investigators including Swietlik [15]. Olives are also good source of iron which helps to transport oxygen in blood. Calcium present in olives is essential for bones and muscles [16]. Olive reduced risk of heart diseases and even help to fight cancer [17]. Olive oil is the healthy component of our diet. Oleonic acid protects liver and improves blood flow [18]. It release stress and boost up immune system in human body. Its fruit contains all essential elements that are necessary for human health. Regularly consuming olives are good for skin health [19]. Olive oil is also used in cosmetics products and in pharmaceutical industry. 100 milliliter (ml), of one type of olive oil, contains Energy: 800 kcal with Fat: 93.3 g, or which 13.33 g is saturated and 66.6 g is monounsaturated [20]. Olive trees are also persistent, easily sprouting back even when cut to the ground [21].

Salinity is an important factor affecting plant productivity and constitutes a problem concerning a significant portion of the planet, especially in regions with hot, dry climates [22,23]. The availability of fresh water is one of the major limitations for crop production. Therefore, the use of non-conventional water resources, such as saline water and reclaimed sewage effluent, has increased in recent years. The utilization of such water resources accelerates the salinization of the upper layer of the soil, where most root activity takes place, and generally decreases crop production [24].

Increased uptake and accumulation of Na^+ and Cl^- ions decreases the absorption of essential minerals and imposes toxicity to plants [25]. Fruit trees such as citrus and grapevines, accumulation of both Na^+ and Cl^- in the roots and aerial parts is the most damaging to the plants often by inhibiting photosynthesis [26]. Na^+ is the primary cause of ion-specific damage (such as reduction in K^+ activity) [27]. Improving plant resistance to salinity may provide yield stability in subsistence agriculture [28].

Gibberellins (GA_3) play a vital role in the detoxification of heavy metals and in tolerance to salt stress by improving plants growth, chlorophyll synthesis and activities of antioxidant enzymes, and by preventing lipid per oxidation [29,30]. Some researchers have used plant growth regulators (PGRs) for reducing or eradicating the negative effects of salinity [31,32]. For example, the exogenous application of PGRs [auxins, gibberellins, cytokinins] produces some benefit in alleviating the adverse effects of salt stress and also improves germination, growth, development, seed yield, and yield quality [33-36]. Azooz, et al. reported that indole acetic acid (IAA) and gibberellic acid stimulate growth in sorghum under stress conditions. Significant differences in salt tolerance have been reported among cultivars [7,37]. Gibberellins generally stimulate cell division and stem elongation and standardize growth by stretching internodes. GA_3 promotes flowering and fruiting in olive plants. GA_3 promotes stem elongation. Gibberellic acid play important role in germination by breaking seed dormancy. GA_3 encouraged cell elongation [38,39]. GA_3 standardizes flower initiation and development. GA_3 contributes in pollination. GA_3 improves fruit quality and promote generation of female flowers [40]. GA_3 application on olive plants diminishes the chances of alternate bearing in olive plants [41]. GA_3 performs key role in protein synthesis and in regulating nucleic acid. Higher concentration of GA_3 can suppress root initiation [42]. Aids plant to swing at suitable time in reproductive phase [43]. Gibberellins are known for their ability to increase cell enlargement, thus enhancing fruit growth in certain species such as citrus, litchi, guava, and pear [8-14].

The beneficial effects of Gibberellic acid (GA_3) and nutrient elements sprays specially zinc on yield and fruit quality of different fruit crops were mentioned by many investigators including Swietlik [15]. Plant growth regulators are produced by the plants and also produce artificially to regulate growth and development under various physiological actions. They are also called plant hormones. These hormones are applied to plants in

different concentrations in order to control and regulate plant growth in different ways. All functions like normal growth, development, root and shoot growth are control by these hormones [45]. Plant growth is inhibited by abscisic acid result in dormancy and abscission. Auxins promote cell elongation and root initiation in cuttings. Natural auxins like Indole butyric acid (IBA) and Naphthalene acetic acid (NAA) hormones play an important role in root formation of stem cuttings [46]. Cytokinins help in cell division. Gibberellins are the very important plant growth regulators and promote cell elongation and also help in fruit developing. Ethylene helps in fruits ripening [47]. Stem elongation is response of signal transduction pathway with different environmental factors. There is a great importance of gibberellic acid in agriculture. Foliar application of GA₃ increases yield. Also improve the quality of fruits and increase the fruit size [48]. An important accumulation of GAs during the flowering transition was found in the petioles of Arabidopsis by Gocal, et al. [49]. Ben Nissan, et al. observed that GIP1 expression (which is a protein induced by GA₃) coincided with cell elongation in stem and flowers transition in *Petunia hybrida*. Mistra and Datta revealed a significant role of GA in the induction of shoots buds on leaf segments of Marigold and Hall and Camper successfully used GA₃ to develop an in vitro culture protocol for Goldenseal species. During favorable condition and germination permissive environment too, seed germination is poor and erratic due to seed dormancy [50-54]. Stratification under cool and moist condition or gibberellic acid treatment improves germination rates. Treatment with 2500ppm of gibberellic acid gives 31.67% germination [55]. The higher concentration of gibberellic acid of about 2000ppm to 6000ppm for 24 hours shortens the germination period. The use of GA₃ primed seed is believed to increase the seed germination and seedling vigor. Since, the germination of Kiwi seed is low and there were unsatisfactory researches based on single factor it was necessary to combine several factors to increase Kiwi seed germination [56]. The role carried out from carbohydrates during the rooting is controversial, but several reasons can explain their behavior. The levels of total carbohydrates and starch in the cuttings are positively related with the rooting but not through one cause-effect relationship [57]. Keeping in view the slow growing of olive plant, this study was designed with an idea to enhance the plant growth in shorten time period for propagation of olive cutting until its transplantation. Therefore foliar spray of Gibberellic acid (GA₃) is more researchable issue to study the effect of gibberellic acid foliar spray on the olive growth.

Materials and Methods

The study was carried out at NARC Islamabad during August, 2018 to October, 2018 to examine to examine the **growth of three olive varieties** i.e. Coratina, Chetoui and Megaron **influences with foliar application of GA₃ under saline conditions** in tunnel. Soil salinity was developed artificially with the mixture of different salts at 2.0dSm⁻¹. Completely randomized design was applied with three replications. Foliar spray of Gibberellic acid @ 0, 200 and 400 mg l⁻¹ was done. Growth parameters of plant height stem diameter internodal distance, # of leaves plant⁻¹, leaf area, fresh weight and dry weight were recorded at the end of the experiment. Data were statistically analyzed according to completely randomized design and compared treatment means using LSD test with statistical software, Statistix 8.1 [57].

Results and Discussions

Table 1 showed the effects of GA₃ foliar application on olive plant height. Maximum height (21.5 cm) was attained at T₂ (400 mg l⁻¹) in Megaron olive variety. Over all Megaron olive cultivar gained the highest plant height (19.66 cm) and Coratina displayed second highest plant height (18.72cm). Maximum plant height was attained at T₂ (400 mg l⁻¹) i.e., 19.17 cm. GA₃ foliar application on olive plant number of leaves was influenced as indicate in Table 1. Maximum number of leaves were attained at T₀ (0 mg l⁻¹) in Chetoui olive variety followed by T₂ (400 mg l⁻¹) i.e., 26.6 in Coratina. Overall Chetoui olive variety attained the maximum number of leaves (29.36). T₀ (00mg/l) showed the maximum number of leaves i.e., 27.46 followed by 24.66 in T₁ (200 mg l⁻¹). Data in Table 1 depicted the effects of GA₃ foliar application on olive plant stem diameter. Maximum stem diameter (0.32cm) was attained at T₀ (0 mg l⁻¹) in two varieties i.e., Coratina and Chetoui. Over all Coratina and Chetoui olive cultivars gained the highest stem diameter i.e., 0.3 cm and 0.29 cm respectively, and T₀ (0 mg l⁻¹) displayed highest stem diameter (0.30cm). The effects of GA₃ foliar application on olive plant leaf area. Maximum leaf area (4.18cm²) was attained at T₁ (200 mg l⁻¹) by the Chetoui olive cultivar. On the whole chetoui variety attained maximum leaf area of 3.98 cm² followed by Coratina i.e., 3.89 cm². T₀ (control) showed maximum leaf area of 3.96 cm² followed by 3.86 cm² in T₂ (400 mg l⁻¹).

This study demonstrated that T₂ (400 mg l⁻¹) depicted maximum vegetative growth of Olive plant as compare to T₁ (200 mg l⁻¹). GA₃ is responsible for cell elongation rather than cell division. Plant hormones, enzymes and phenolic

compounds play a major role in internal control mechanism of rooting in olives. The endogenous phenolic compounds have different effects especially on rooting cuttings [58-60]. GA₃ foliar application enhances bud sprouting and shoots elongation. GA₃ was successfully used by Grigoriadou, et al. for shoot proliferation of certain olive tree cultivars. From the above mentioned results it was noticed that GA₃ foliar sprays at 400mg/l concentration was more effective in plant height than

spraying GA₃ at other concentrations and control. Highest stem diameter was obtained from cuttings sprayed with T₂ (400 mg l⁻¹) GA₃ while control treatment showed the lowest diameter of olive cuttings. The present results may be attributed to stimulative impact of GA₃ on cell extension and on cell division. GA₃ is an interesting hormone for in vitro shoot elongation of many other species such as Macadamia, Acacia [60-65].

Treatment	Plant height (cm)			Stem diameter(cm)			# of leaves plant ⁻¹			Leaf area (cm ²)		
	V ₁	V ₂	V ₃	V ₁	V ₂	V ₃	V ₁	V ₂	V ₃	V ₁	V ₂	V ₃
T ₀	18.7	19.6	19	0.32	0.32	0.27	21.9	36.2	24.3	4.16	4.14	3.6
T ₁	18	17.5	18.5	0.27	0.3	0.27	25.3	26	22.7	3.49	4.18	3.4
T ₂	19.5	16.5	21.5	0.31	0.27	0.27	26.6	25.9	20.9	4.02	3.63	3.95
Mean	18.7	17.9	19.7	0.3	0.29	0.27	24.6	29.4	22.6	3.89	3.98	3.65

Table1: Olive Varieties Growth in Tunnel Influences with Foliar application of GA₃ under Saline Conditions.

T₀= 0 mg l⁻¹ GA₃, T₁= 200 mg l⁻¹ GA₃, T₂= 400 mg l⁻¹ GA₃, V₁= Coratina, V₂= Chetoui, V₃= Megaron

Conclusion

In three olive varieties studied in this protocol, gibberellins foliar spray had potential for the improvement of growth characteristics.

References

1. Payvandi M, Dadashian A, Ebrahimzadeh H, Madjd A (2001) Embryogenesis and rhizogenesis in mature zygotic embryos of olive (*Olea europaea* L.) cultivars Mission and Kroneiki. *J Sci IR Iran* 12(1): 9-15.
2. Bandelj D, Jakse J, Javornik B (2004) Assessment of genetic variability by microsatellite and AFLP markers. *Euphytica* 136(1): 93-102.
3. Singh A, Babu KD, Patel RK, Barua U (2006) Temperate Fruits: Gaining momentum in subtropical areas of North East. *ENVIS Bull Himalayan Ecol* 14(1): 1-87.
4. Caballero JM, Del C Río (2008) The olive world germplasm bank of Spain. *Acta Horticulturae (ISHS)* 791: 31-38.
5. Barranco D, Rallo L (2000) Olive cultivars in Spain. *Hortech* 10(1): 107-110.
6. Baldoni L, Pellegrini M, Mencuccini M, Angiolillo A, Mulas M (2000) Genetic relationships among cultivated and wild olives revealed by AFLP markers. *Acta Horticulturae* 521: 275 -284.
7. Chartzoulakis K, Loupassaki M, Bertaki M, Androulakis I (2002) Effects of NaCl salinity on growth, ion content and CO₂ assimilation rate of six olive cultivars. *Scientia Hort* 96(1-4): 235-247.
8. Davis PJ (2004) The plant hormones: their nature, occurrence and functions. In: Davis PJ (Eds.), *Plant Hormones*. Kluwer Academic Publishers, Dordrecht, The Netherlands, pp: 1-15.
9. Eman AA, Abd El-moneim MMM, Abd El Migeed O, Ismail MM (2007) GA₃ and Zinc Sprays for Improving Yield and Fruit Quality of Washington Navel Orange Trees Grown under Sandy Soil Conditions. *Res J Agric Biol Sci* 3(5): 498-503.
10. El-Sese AMA (2005) Effect of gibberellic acid (GA₃) on yield and fruit characteristics of Balady mandarin. *Assiut J Agric Sci* 36(1): 23-35.
11. Stern RA, Gazit S (2000) Reducing fruit drop in lychee with PGR sprays. In: Basra A (Eds.), *Plant Growth Regulators in Agriculture and Horticulture*. The Haworth Press Inc, pp: 211-222.
12. Chang JC, Lin TS (2006) GA₃ increases fruit weight in 'Yu Her Pau' litchi. *Sci Horticult* 108(4): 442-443.

13. El-Sharkawy SHMM, Mehaisen SMA (2005) Effect of gibberellin and potassium foliage sprays on productivity and fruit quality of guava trees. *Egypt J Appl Sci* 20(3): 151-162.
14. Zhang C, Tanabe K, Tani H, Nakajima H, Mori M, et al. (2007) Biologically active gibberellins and abscisic acid in fruit of two late-maturing Japanese pear cultivars with contrasting fruit size. *J Am Soc Horticul Sci* 132(4): 452-458.
15. Swietlik D (2002) Zinc Nutrition of Fruit Trees by Foliar Sprays. *International Symposium on Foliar Nutrition of Perennial Fruit Plants. Acta Horti*, pp: 594.
16. Lastra A de la, Barranco C, Motilva MDV, Herrerías JM (2001) Mediterranean diet and health: Biological importance of olive oil. *Current Pharmaceutical Design* 7(10): 933-950.
17. Owen RW, Giacosa A, Hull WE, Haubner R, Wurtele G, et al. (2000b) Olive-oil consumption and health: the possible role of antioxidants. *Lancet Oncology* 1: 107-112.
18. Visioli F, Galli C, Bornet F, Mattei A, Patelli R (2000) Olive oil phenolics are dose-dependently absorbed in humans. *FEBS Lett* 468(2-3): 159-160.
19. Bianco AD, Uccella N (2000) Biophenolic Components of Olives. *Food Res Int* 33(6): 475-485.
20. Moreno JJ, Mitjavila MT (2003) The degree of unsaturation of dietary fatty acids and the development of atherosclerosis (Review). *J Nutr Biochem* 14(4): 182-195.
21. Reiger T (2005) The emergence of words: Attentional learning in form and meaning. *Cognitive Science* 28: 819-865.
22. Chartzoulakis K, Psarras G, Vemmos S, Loupassaki M, Bertaki M (2006) Response of two olive cultivars to salt stress and potassium supplement. *J Plant Nut* 29(11): 2063-2078.
23. Bazakas C, Manioudaki ME, Sarropoulou E, Spano T, Kalaitzis P (2015) 454 pyrosequencing of olive (*Olea europaea* L.) transcriptome in response to salinity. *PloS one* 10(11): 1-22.
24. Demiral MA, Aktaşuygun D, Uygün M, Kasırğa E, Karagozler AA (2011) Biochemical response of *Olea europaea* cv. Gemlik to short term salt stress. *Turk J Biol* 35: 433-442.
25. Tester M, Devenport R (2003) Na⁺ tolerance and Na⁺ transport in higher plants. *Ann Bot* 91(5): 503-527.
26. White PJ, Broadley MR (2001) Chloride in soils and its uptake and movement within the plant: A review. *Ann Bot* 88(6): 967-988.
27. Tabatabaei SJ (2006) Effects of salinity and N on the growth, photosynthesis and N status of olive (*Olea europaea* L.) trees. *Scientia Hort* 108(4): 432-438.
28. Asghari HR (2008) Vesicular-arbuscular (VA) mycorrhizae improve salinity tolerance in pre-inoculation subterranean clover (*Trifolium subterraneum*) seedlings. *Int J Plant Production* 2: 243-256.
29. Saeidi-Sar S, Khavari-Nejad R, Fahimi H, Ghorbanli M, Majd A (2007) Interactive effects of gibberellin GA₃ and ascorbic acid on lipid peroxidation and antioxidant enzyme activities in Glycine max seedlings under nickel stress. *Russ J Plant Physiol* 54(1): 74-79.
30. Maggio A, Barbieri G, Raimondi G, Pascale SD (2010) Contrasting effects of GA₃ treatments on tomato plants exposed to increasing salinity. *J Plant Growth Regul* 29(1): 63-72.
31. Angrish A, Kumar B, Datta KS (2001) Effect of gibberellic acid and kinetin on nitrogen content and nitrate reductase activity in wheat under saline conditions. *Ind J Plant Physiol* 6: 172-177.
32. Chakrabarti N, Mukherji S (2002) Effect of phytohormones pretreatment on metabolic changes in *Vigna radiata* under salt stress. *J Env Biol* 23:295-300.
33. Khan MA, Gul B, Weber DJ (2004) Action of plant growth regulators and salinity on seed germination of *Ceratoides lanata*. *Can J Bot* 82(1): 37-42.
34. Shaheena A, Firoz M, Shamsul H, Manzer H (2005) Exogenous Application of gibberellic Acid counteracts the effect of sodium chloride in mustard. *Turk J Biol* 29: 233-236.
35. Gul B, Khan MA, Weber DJ (2000) Alleviation salinity and dark enforced dormancy in *Allenrolfea*

- occidentalis seeds under various thermo periods. *Aust J Bot* 48(6): 745-752.
36. Egamberdieva D (2009) Alleviation of salt stress by plant growth regulators and IAA producing bacteria in wheat. *Acta Physiol Plant* 31(4): 861-864.
 37. Azooz MM, Shaddad MA, Abdel-Latef AA (2004) Leaf growth and K⁺, Na⁺ ratio as an indication of the salt tolerance of three sorghum cultivars grown under salinity stress and IAA treatment. *Acta Agron Hung* 52: 287-296.
 38. Hannachi H, Breton C, Msallem M, Ben El Hadj S, El Gazzah M, et al. (2008) Are Olive Cultivars Distinguessable from Oleaster Trees Based on Morphology of Drupes and Pits, Oil Composition and Microsatellite Polymorphisms. *Acta Botanica Galica* 155(4): 531-545.
 39. Ismaili H (2016) Study of Some forms of IBA in the Rooting Process of the Olive. *Int J Curr Microbiol App Sci* 5(3): 239-246.
 40. Taiz L, Zeiger E (2004) *Fisiologia Vegetal Porto Alegre*. Artmed, pp: 719.
 41. Afroz A, Chaudhry Z, Khan R, Rashid H, Khan SA (2009) Effect of GA₃ on regeneration response of three tomato cultivars (*Lycopersicon esculentum*). *Pak J Bot* 41(1): 143-151.
 42. Hartmann HT, Kester DE, Davis JFT, Geneve RL (2002) *Plant propagation: principles and practices*. New Jersey: Prentice Hall, pp: 880 .
 43. Desouky IM, Haggag FL, El-Migeed MMMA, Kishk YFM, El-Hady ES (2014) Effect of boron and calcium nutrients sprays on fruit set, oil content and oil quality of some olive oil cultivars. *Sci Lett* 2(2): 48-52.
 44. Jamal Uddin AFM, Hossan MJ, Islam MS, Ahsan MK, Mehraj H (2012) Strawberry growth and yield responses to gibberellic acid concentrations. *J Expt Biosci* 3(2): 51-56.
 45. Agusti J, Herold S, Schwarz M, Sanchez P, Ljung K, et al. (2011) Strigolactone signaling is required for auxin-dependent stimulation of secondary growth in plants. *Proc Natl Acad Sci USA* 108(50): 20242-20247.
 46. Francis D, Sorrell DA (2001) The interface between the cell cycle and plant growth. *Plant Growth Regulation* 33(1): 1-12.
 47. Ramezani S, Shekafandeh A (2009) Roles of gibberellic acid and zinc sulphate in increasing size and weight of Olive fruit. *African Journal of Biotechnology* 8(24): 6791-6794.
 48. Gocal GFW, Sheldon CC, Gubler F, Moritz T, Bagnall DJ, et al. (2001) GAMY B-like Genes, Flowering and Gibberellin Signaling in Arabidopsis. *Plant Physiol* 127(4): 1682-1693.
 49. Ben-Nissan G, Lee JY, Borohov A, Weiss D (2004) GIP, *Petunia hybrida* GA-Induced cysteine-rich protein: A possible role in shoot elongation and transition to flowering. *The Plant J* 37(2): 229-238.
 50. Mistra P, Datta SK (2001) Direct differentiation of shoot buds in leaf segments of white marigold (*Tagetes erecta* L.). *In Vitro Cellular and Development biology-Plant* 37(4): 466-470.
 51. Hall KC, Camper ND (2002) Tissue culture of Goldenseal (*Hydrastis canadensis* L.). *In Vitro Cellular and Development biology-Plant* 38(3): 293-295.
 52. Bradford KJ (2005) Threshold models applied to seed germination ecology. *New Phytologist* 165(2): 338-341.
 53. Finch-Savage, Leubner Metzger G (2006) Seed dormancy and the control of germination. *New Phytol* 171(3): 501-523.
 54. Ozcan M, E Erisgine (2000) The effects of some application on seed germination and seedling growth in kiwi fruit. *Bulletin of Pure and Applied Sciences* 19: 25-31.
 55. Çelik H, Zenginbal H, Özcan M (2006) Enhancing germination of kiwifruit seeds with temperature, medium and gibberellic acid. *Hort Sci (Prague)* 33(1): 39-45.
 56. Rahman N, Awan AA, Nabi G (2002) Root initiation in hard wood cutting of olive cultivar corating using different concentration of IBA. *Asian J Plant Sciences* 1: 563-564.
 57. (2005) Analytical software Statistix 8.1 for Windows. Analytical Software, Tallahassee, Florida.

58. Ercan H, Ozkaya MT (2008) The monthly change of phenolic compounds in some Turkish olive cultivars. *Acta Hort* 791: 321-324.
59. Lee OH, Lee BY, Lee J, Lee HB (2009) Assessment of phenolics- enriched extract and fractions of olive leaves and their antioxidant activities. *Bioresour Technol* 100(23): 6107-6113.
60. Perez-Bnilla M, Salido S, Beek TA, Linares-Palomia PJ, Altarejos J, et al. (2006) Isolation and identification of radical scavengers in olive tree (*Olea europaea*) wood. *J Chromatogr* 1112(1-2): 311-318.
61. Grigoriadou K, Vasilakakis M, Eleftheriou EP (2002) *In vitro* propagation of the Greek olive cultivar 'Chondrolia Chalkidikis'. *Plant Cell Tissue Org Cult* 71(1): 47-54.
62. Mulwa RMS, Bhalla P (2000) *In vitro* shoot multiplication of *Macadamia tetraphylla* L. Johnson. *J Horti Sci Biotech* 75(1): 1-5.
63. Vengadesan G, Ganapathi A, Amutha S, Selvaraj N (2003a) High-frequency plant regeneration from cotyledon callus of *Acacia sinuata* (Lour.) Merr. *In Vitro Cell Dev Biol-Plant* 39(1): 28-33.
64. Barranco D, Trujillo I, Rallo P (2000b) Are 'Oblonga' and 'Fantoio' the same cultivar? *HortScience* 35(7): 1323-1325.
65. Khan AS, Chaudhry NY (2006) GA₃ improves flower yield in some cucurbits treated with lead and mercury. *Afr J Biotechnol* 5(2): 149-153.

