

New Effective Method Preservation of for Red Blood Cells by Means of Nanotechnology

Belousov A*, Malygon E, Yavorskiy V, and Belousova E

Laboratory of Applied Nanotechnology of Belousov, Kharkov Medical Academy of Postgraduate Education, Ukraine

***Corresponding author:** Andrey Belousov, Laboratory of Applied Nanotechnology of Belousov Kharkov Medical Academy of Postgraduate Education, Ukraine Kharkov Regional Center of Blood Service, Ukraine, Tel: 38050-915-18-89; E-mail: an.belousov2012@ukr.net

Research Article

Volume 3 Issue 3

Received Date: August 29, 2018

Published Date: September 17, 2018

DOI: 10.23880/nnoa-16000149

Abstract

This study was devoted to the learning of the use of nanotechnology to correct the functional activity of red blood cells (RBCs) at the storage stages at a positive temperature. It was established that saline NaCl, which had previously been processed by magnetite nanoparticles (ICNB) had a marked membrane-stabilizing effect, inhibits hemolysis and increasing the sedimentation stability of preserved RBCs. The complex analysis of the obtained data allowed to determine the primary mechanisms effect of the saline NaCl, which had previously been processed by ICNB on the preserved RBCs. The proposed method of additive modernization of preserved RBCs was adapted to the production process. The optimisation results were obtained in creating a simple and practical method of additive modernization of preservation solutions that does not violate the compliance requirements, improves the quality, efficiency and safety transfusion of RBCs.

Keywords: Nanotechnology; Preserved Red Blood Cells; Magnetite Nanoparticles (ICNB); RBCs storage; production process

Introduction

Actuality Red blood cells (RBCs) transfusion is a critical, life-saving treatment for severe anemia caused by disease or chemotherapy, or by blood loss due to trauma or major surgery. For several decades RBCs components have been prepared as concentrates suspended in nutrient additive solution, which preserves and extends the shelf-life of the RBCs component, allowing up to 6–7 weeks of

refrigerated storage [1]. Nevertheless, during storage RBCs undergo a complex and progressive accumulation of physicochemical changes, collectively referred to as the RBCs storage lesion [2,3]. Recent clinical studies have identified RBCs transfusion as an independent risk factor for increased morbidities and mortalities in certain groups of patients, including trauma, cardiac surgery and the critically- ill (reviewed in [4-6]). Additionally, some of these studies have identified that older stored RBCs are

more strongly implicated in poorer outcomes compared to fresher RBCs [6]. In order to address these concerns, there is renewed interest to better understand the RBCs storage lesion and to find ways to ameliorate the deleterious effects of storage, thereby improving the quality, efficacy and safety of RBCs components for all transfusion recipients.

While increased research effort is being directed to better understand the effects of storage on RBCs and the potential impact on transfusion outcomes [7], slower progress is being made in finding ways to deter the detrimental effects of the RBCs storage lesion.

Over the past 15–20 years, research into the development of new additive solutions has focused on ways to maintain higher intracellular levels of ATP and 2,3-DPG during storage of RBC components [1].

Despite the RBCs having been a favourite experimental model for cellular biologists and biochemists, RBCs storage research has repeatedly demonstrated that a lot of fundamental biology about RBCs is still not well understood. The complexity of the inter-relationship between RBCs biochemistry, cytoskeletal structure and membrane properties have made it difficult to predict how RBCs will respond to different storage conditions. Exposure of RBCs to non-physiological storage environments has pointed to the existence of previously unknown biochemical mechanisms in RBCs, including apoptotic-like processes, ion and osmotic channels that behave differently than expected, exposure of new or altered receptors possibly due to oxidative and/or protease/glycosidase activities or altered senescence [8–11].

The benefits gained by improved RBCs component quality should more than justify any real or perceived inconvenience to the blood services in implementing adjustments to their processing procedures or additional processing costs of the introduction of new generation RBCs additive solutions. The bigger challenge that has hindered the advancement of this field is the significant financial burden and risk for manufacturers of blood collection systems to obtain licensure and to bring a new RBCs storage system to a market that is inherently based on very low profit margins, such as the blood services sector.

The financial burden to technology developers of new RBC storage systems is largely due to regulatory requirements, particularly those mandated by the FDA. In

addition to in vitro data, the FDA requires in vivo data on the 24 hours post transfusion recovery of transfused autologous RBCs. Recently the FDA has tightened and increased the assessment and acceptance criteria making it potentially more difficult and expensive to bring new RBCs storage systems to market. Although the regulatory agencies are to be commended for focusing on the safety of new therapies and devices for patients, there are concerns that the regulatory requirements for RBC storage systems have become excessive and are hindering progress [12].

Another significant challenge for obtaining licensure of new RBCs storage systems is the inherent donor- related variability in stored RBCs quality. It has long been recognized that RBCs from some donors do not store well, as evidenced by higher levels of haemolysis at RBC component expiry 14 and poorer in vivo 24 hr recovery data [13]. The relationship of specific donors and poorer quality of some stored RBCs components was confirmed in a recent paired cross-over study designed to compare manual and automated whole blood processing methods [14,15]. Technology developers are unwilling to take on the risk that a random poor quality RBCs component could jeopardise the success of licensure tests and clinical trials of their new blood storage systems and their significant financial investment.

In Ukraine, the first standardized and biocompatible magnetite nanoparticles for medical use were manufactured and patented in 1998. These are intracorporal nanobiocorrector of brand ICNB, magnet-controlled sorbent of brand MCS-B, and biologically active nanodevice of brand Micromage-B [16].

It is well established that the magnetite nanoparticles effectively modulate the metabolic processes in leukocytes, regulate activity of the enzyme link of the antioxidant system in erythrocytes in healthy and sick patients [17–19]. Previously the complex investigations that were performed in the study of the influence on metabolism of cells by preparations of nanotechnology show that in whole standardized biocompatibility of magnetite nanoparticles have nonspecific and modulated effect on metabolic processes. Research of ultra structure investigations of the reticuloendothelial system (liver, lungs and kidneys) it was proved that after injection of biocompatibility magnetite nanoparticles into a vein caused nonspecific activation of the metabolic processes, increase adaptive mechanisms and potential of organelle cells, acceleration of reparative processes a level of membranes and macromolecules [18,20,21]. Existing sorption and indirect (magnetic) effects not only allow

selectively absorb the protein of surface membrane cells by magnetite nanoparticles (according to the principle of magnetophoreses), but also to prevent the oxidative modification of proteins by way of stabilizing the active groups, normalizing a state of receptors that are located on the surface membrane of cells, increasing activity of enzymes' membrane-bound [22-24].

Recent scientific work related to use of magnetite nanoparticles (ICNB) in contrast means in an MRI investigation of cancer reliably was shown that nanoparticles cause reversible changes associated with a temporary increase in the mobility of hydrogen protons in the pericellular fluid that inevitably modifies the metabolism in malignant cells [25]. The results of these investigations have not only widened the understanding of the mechanisms of action of nanoparticles on condition outside and intracellular spaces but also have revealed new aspects of the cellular (cells) metabolism, determined the membrane role of cellular enzymes in the regulation processes of metabolism [23,25-28].

Also, it was established that extra corporally processing the blood by nanoparticles of MCS-B reliably reduces activity of Ca, Mg - ATPase of erythrocytes. Currently, studies have shown that magnetite nanoparticles are able to inhibit hemolysis of heparinized blood, increase the activity of ATP and 2.3 DPH in red blood cells, regulate transmembrane metabolism and inhibit eryptosis [23,29,30].

The above was the basis for the choice of the theme of this study, devoted to the learning of the use of nanotechnology to correct the functional activity of red blood cells at the storage stages at a positive temperature.

The main purpose of the first stage of the study is to develop a simple and practical method of additive modernization of preservation solutions that does not violate the compliance requirements, improves the quality, efficiency and safety transfusion of red blood cells.

Materials and methods

Materials: 1. Standardized intra corporeal nano bio corrector of ICNB was taken as nanoparticles. Magnetite nanoparticles synthesized by co-precipitation method. The main physics and chemical properties of ICNB the following data and also in Tables 1-4; Figures 1,2 were presented:

- Concentration of the colloidal solution of magnetite nanoparticles in physiology solution of NaCl is 0.0225%.
- Theoretical osmolality of colloid solution is 500 mosmol/l
- Size of magnetite nanoparticles is 6-12 nm;
- Total area of surface magnetite of nanoparticles $S_s = 800-1200 \text{ m}^2/\text{g}$;
- Magnetization of saturation $I_s = 2.15 \text{ kA/m}$;
- ζ - Potential = - 19 mV.

Phase name	a (Å)	b (Å)	c (Å)	alpha (град)	beta (град)	gamma (град)
magnetite low	8.39	8.39	8.39	90	90	90
magnetite low, syn	5.93	5.93	14.7	90	90	120
Johannsenite	9.89	9.06	5.28	90	105.5	90

Table 1: The calculated lattice parameters of the phases

Compound	Wt%	StdErr	El	Weight%/ O ₂	StdErr	El	Weight%	StdErr
Fe ₃ O ₄	97.37	0.09	Fe	68.4	0.07	Fe	97.62	0.09
CaO	2.26	0.07	Ca	1.71	0.05	Ca	2.3	0.07
P ₂ O ₅	0.28	0.027	Px	0.122	0.012	Px	0.157	0.015
MnO	0.255	0.013	Mn	0.198	0.01	Mn	0.278	0.014
SiO ₂	0.098	0.027	Si	0.046	0.013	Si	0.059	0.016
SO ₃	0.032	0.013	Sx	0.0126	0.0051	Sx	0.0164	0.0066
Cl	0.028	0.009	Cl	0.028	0.009	Cl	0.038	0.012

Table 2: Determination of percent composition of the ICNB by X-ray spectrometer ARL OPTIM'X (semi- quantitative analysis)

Phase	Formula	Space group	Nº Card Database ICDD
magnetite low	$\text{Fe}_2.886\text{O}_4$	227 : Fd-3m, choice-2	10861339 (ICDD)
magnetite low, syn	Fe_3O_4	166 : R-3m, hexagonal	10716766 (ICDD)
Johannsenite	$\text{Ca Mn} + 2\text{Si}_2\text{O}_6$	15 : C12/C1, unique-b, cell-1	380413 (ICDD)

Table 3: X-ray analysis of ICNB in X-ray diffractometer Rigaku Ultima IV ($\text{CuK}\alpha$, $\text{K}\beta$ filter - Ni), one- coordinate DTeX semiconductor detector

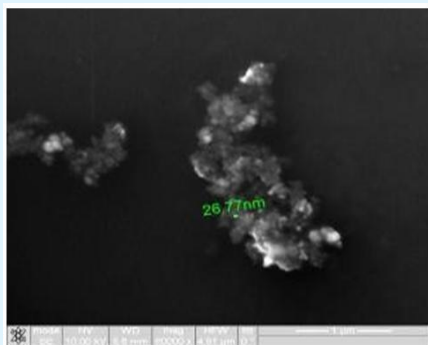


Figure 1: Study of magnetite nanoparticles with use microscope ion-electronic raster-type Quanta 200 3 D

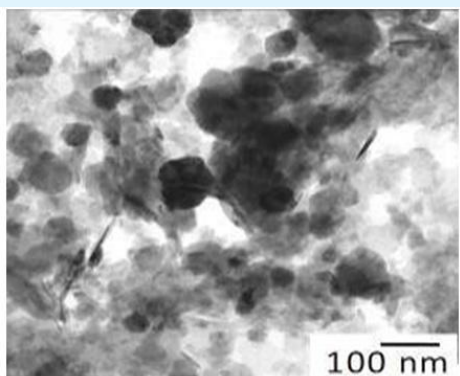


Figure 2: Study of magnetite nanoparticles with use microscope electronic translucent JEM-2100

Phases (method of corundum numbers)	Content, %
magnetite low	71
magnetite low, syn (hexagonal)	29

Table 4: The phases of magnetite of nanoparticles (RIR - method; error $8\pm 3\%$)

1. 0.9% NaCl solution.
2. 0.9% NaCl solution which previously was processed by ICNB in ratio 4:1.

Object of Research

- red blood cells (RBCs) into bags containing anticoagulant citrate, nutrient phosphate and dextrose (CPD);
- Red blood cells (RBCs) into bags containing anticoagulant citrate, nutrient phosphate, dextrose and adenine (CPDA-1).

Methods

Before starting the experience with red blood cells (RBCs) we used visual assessment by comparison of the brightness of the images different variants of solutions. These were: ICNB, 0.9% NaCl solution and 0.9% NaCl solution that had been treated by magnetite nanoparticles (ICNB).

The tests were performed on the Siemens MR-tomography Magnetom Concerto with power magnetic-field 0.2 T.

The axial tomograms were received:

1. T1 - the self-weighted sequences of Echo Spin of TR 50 ms, TE 17 ms the field of review the 250 mm, the thickness cut 2 mm.
2. T2 - the self-weighted sequences Echo Gradient of TR 500 ms, TE 17 ms the field of review a 180 mm, the thickness cut 4 mm.

Of each bag of 3 ml amounts of red blood cells was distributed into 20 sterile glass tubes. Then, into the first 10 tubes of control were added of 2 ml amounts 0.9% NaCl solution. Into the next 10 tubes of test were added of 2 ml amounts 0.9% NaCl solution, which previously was processed by ICNB.

Thus, the distribution of tubes was as the follows:

Tubes of control:

- 3 ml of red blood cells (CPD) + 2 ml 0.9% NaCl solution (n=10);
- 3 ml of red blood cells (CPDA-1) + 2 ml 0.9% NaCl solution (n=10).

Tubes of test:

- 3 ml of red blood cells (CPD) + 2 ml 0.9% NaCl solution that previously was processed by ICNB in ratio 4:1 (n=10);

- 3 ml of red blood cells (CPDA-1) +2 ml 0.9% NaCl solution that previously was processed by ICNB in ratio 4:1 (n=10).

The state of red blood cells was determined visually by the registration of signs of hemolysis. Also, hemolysis was controlled by photometric method by means Plasma / Low Hb and GPHP-01 devices. The centrifuge mark of SM-70M-07 was used to obtain supernatant. Hematocrit (HCT) was calculated by means hematocrit ruler and using the formula:

$$\text{HCT} = \frac{\text{Free Hb} \times (1 - \text{HCT})}{\text{Common Hb}} \times 100\%$$

Morphology of the red blood cells was studied by direct microscopic method. Sedimentation stability of red blood cells was studied by Panchenkov's method. Change

in the acidity of the red blood cells was performed by means of pH metric.

Tests were carried out in six stages: day 1 - I, day 7 - II, day 14 - III, day 21 - IV, day 28 - V, day 35 - VI. The blood after performance of the biochemical investigation was stored in the refrigerating chamber at temperature +4°C.

Statistically processing the obtained results was carried out by parametrical method of variation statistics by Student criterion. Processing the obtained data was carried out by means of Excel.

Results and Discussion

Results of visual assessment of the images brightness used in experiment liquids at the MRI are represented in Figure 3.

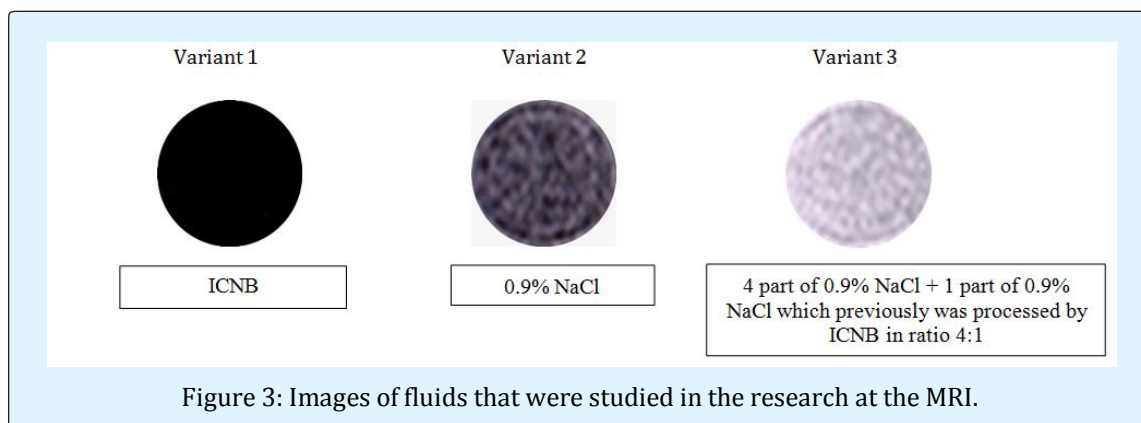


Figure 3 illustrates the difference of image brightness in liquids compared at the MRI. The order of the brightness increase is the following: ICNB, 0.9% NaCl solution, 0.9% NaCl solution that was treated by nanoparticles of ICNB. The difference in brightness of images is explained in the following way:

- Variant 1: Magnetite nanoparticles of ICNB reduce the mobility of hydrogen protons in the liquid medium (0.9% NaCl solution). Therefore, the image brightness was very low in the MRI.
- Variant 2: Rising mobility of hydrogen protons in the intact 0.9% NaCl solution increased brightness in comparison with Variant 1.
- Variant 3: Mobility of hydrogen protons in 0.9% NaCl solution that has previously been processed ICNB nanoparticles is maximized. Therefore, the image brightness is much higher than before (in the previous variants).

Thus, previously conducted research clearly shows that the nanoparticles of ICNB change the mobility and the orientation of the hydrogen atoms in liquids that are registered in the visual evaluation of MRI.

The next set of studies was essential and aimed at studying of functional activity of red blood cells at the storage stages at a positive temperature after by modifying the mobility and spatial orientation of hydrogen protons in the pericellular fluid using magnetite nanopatricles of ICNB.

A study of the sedimentation stability of RBCs showed a highly significant difference between control and test data. Data of sedimentation stability of the RBCs at the stages of a study were presented in Figure 4.

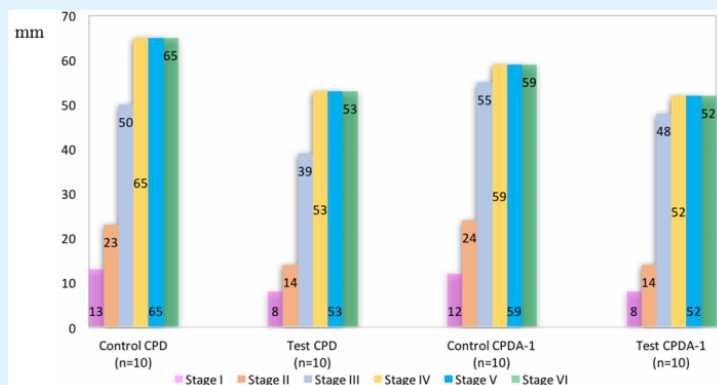


Figure 4: Study of the sedimentation stability of RBCs at the stages ($M \pm m$; $p < 0.001$; $n = 10$).

Figure 4 shows that sedimentation stability of RBC in test tubes are reliably more highly ($p < 0.001$) then in control tubes at stages of research. It should be said that negative surface charge of human RBCs's results primarily from the presence of ionogenic carboxyl groups of sialic acids on the cell surface [31-33]. The value of the charge is determined by the amount of adenosine three phosphates (ATP). ATP is macroergic compounds, the

product of glycolysis. The sedimentation stability of RBC is determined by the amount of ATP.

In this case, the change of mobility and spatial orientation of the hydrogen protons in the extracellular liquid significantly increased the sedimentation stability of RBCs in the test compared to the control. For greater clarity results of RBCs sedimentation is shown in Figure 5.

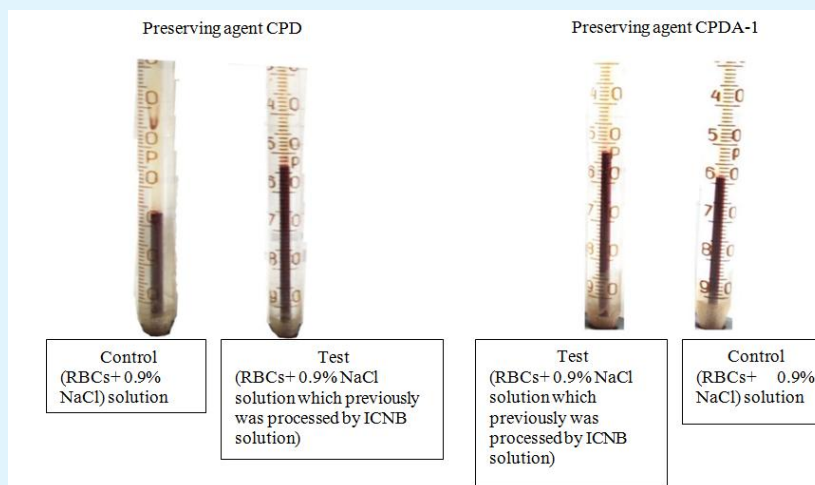


Figure 5: Sedimentation stability of erythrocytes at the stage VI of the study.

Figure 5 is showing that with preserved in the anticoagulant CPD the sedimentation stability in the control was 62 mm; in the test – 53 mm. With preserved in the anticoagulant CPDA-1: control – 58 mm; test – 52 mm.

Thus, following the logic of the above reasoning, if improving sedimentation stability of RBC is associate with

an increase in ATP, then the isotonic solution which previously was processed by ICNB should actively stabilize the membranes of RBCs and inhibit hemolysis. Therefore, the next investigation was to study the hemolysis processes preserved of the RBCs at various stages. Results of the visual assessment hemolysis of erythrocytes in various aspects of exposure are presenting in Figure 6.

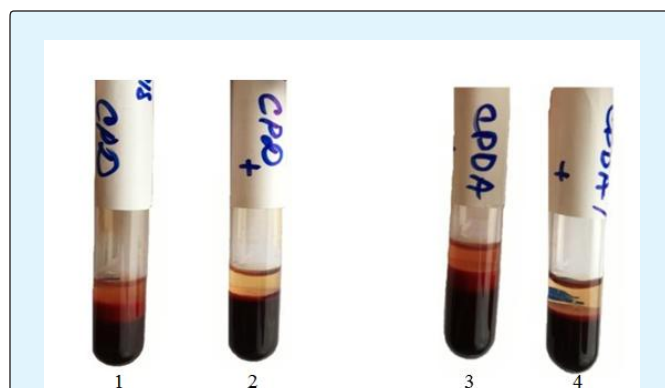


Figure 6: Visual assessment hemolysis of erythrocytes in various aspects of exposure at the stage VI.

Notes: 1- RBC (preserving agent CPD) + 0.9% NaCl solution; 2 - RBCs (preserving agent CPD) + 0.9% NaCl solution which previously was processed by ICNB; 3- RBCs (preserving agent CPDA-1) + 0.9% NaCl solution; 4 - RBCs (preserving agent CPDA-1) + 0.9% NaCl solution which previously was processed by ICNB.

Figure 7 shows that in the control tubes at the stage VI of the study there are pronounced signs of hemolysis. In contrast, hemolysis is not recorded in test tubes. Visual analysis was supplemented by objective data of the photometric method, as well as the method of calculation of hematocrit.

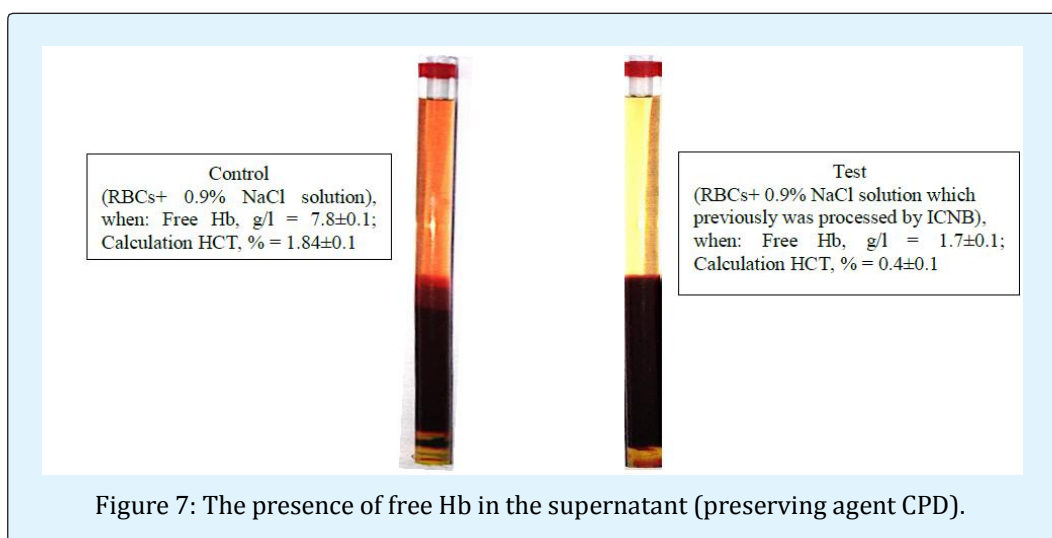


Figure 7: The presence of free Hb in the supernatant (preserving agent CPD).

Objective data of free Hb and calculated HCT at the stage VI of the study are presented in Table 5.

Preservatives	Variants	Stage VI		p
		Free Hb, g/l	Calculation HCT, %	
CPD	Control (n=10)	7.8 ± 0.1	1.84 ± 0.1	<0.001
	Test (n=10)	1.7 ± 0.1	0.4 ± 0.1	<0.001
CPDA-1	Control (n=10)	8.0 ± 0.1	1.9 ± 0.1	<0.001
	Test (n=10)	1.6 ± 0.1	0.5 ± 0.1	<0.001

Table 5: The indicators of free Hb and calculated HCT at the stage VI of the study at baseline values: Hb = 154 g/l; CT = 0.46.

The data in Table 5 indicate that in test tubes the average content of free Hb in the VI stage of the study 6.25 ± 0.1 g/l were less than controls; and the average calculation HCT was lower by 1.42%.

Thus, the objective indicators of Table 5 are highly reliable ($p < 0.001$) and confirm the obtained visual effect

of inhibiting hemolysis of preserved RBCs by physiological solution, which was pretreated by ICNB. Microscopic observation of changes in the morphology of RBCs was a logical continuation of the study of the erythrocyte membranes stabilization effect. The results of microscopic examination of the morphology of

erythrocytes in different variants of preservation and treatment are shown in Figure 8.

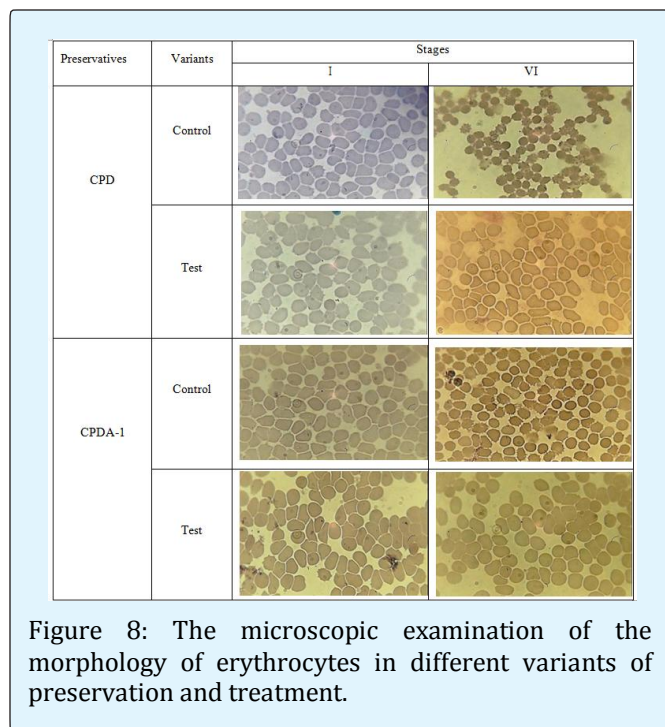


Figure 8: The microscopic examination of the morphology of erythrocytes in different variants of preservation and treatment.

Figure 8 clearly demonstrates that microscopically in the control variant at the VI stage of the study, widespread appearance of spherocytosis is observed.

On the contrary, in the test variants, the shape of red blood cells at the stage VI was unchanged. Pathological changes in the RBC's shape and size in the control variants are most likely associated with inhibition of glycolysis processes [34]. Consequently, the number of ATP and 2,3 DPG decreases, the permeability of the erythrocyte membranes is disturbed, the state of the hemoglobin buffer changes. As a result, the pH of intracellular and extracellular media is changes.

The decrease in the formation of 2,3 DPG leads to the acidulation of intracellular environment of the RBCs. Deoxygenated hemoglobin which was previously formed actively binds the $[H^+]$ that comes from the extracellular environment and alkalizes the extracellular environment. The effect of RBC reduction, the appearance of widespread spherocytosis is observed in microscopy. Subsequently, oxy hemoglobin moves to the extracellular environment as a result of processes intensification destruction of the membranes of RBCs. The accumulation of oxygenated hemoglobin in the extracellular environment causes by shifting towards the acid of the pH.

The above mechanisms have been confirmed in the study of the dynamics of pH changes in the extracellular medium of preserved of the RBCs. The dynamics of pH changes in the extracellular medium of RBCs storage at key stages of the study on the example of preserving agent CPD is shown in Figure 9.

	Preservatives	Stages		
		I	V	VI
Control	CPD			
		pH = 7.3-7.4	pH = 7.2-7.3	pH = 7.1-7.2
Test	CPD			
		pH = 7.3-7.4	pH = 7.3-7.4	pH = 7.4-7.5

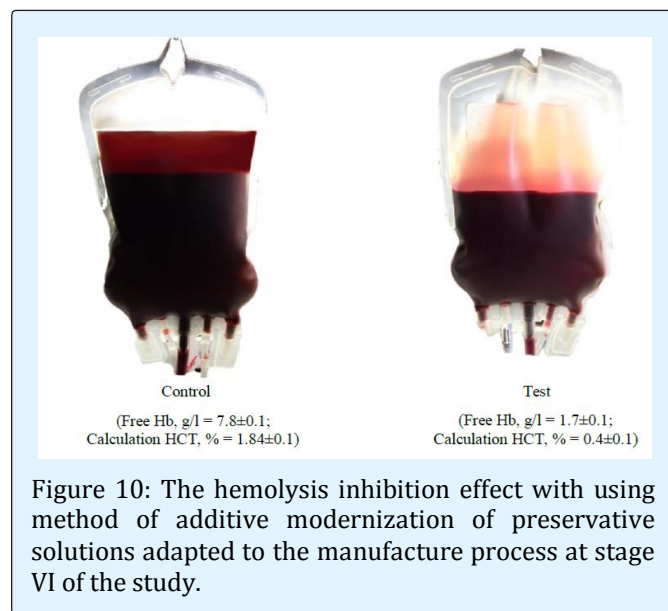
Figure 9: Dynamics of pH changes in the extracellular medium of RBCs storage at key stages of the study on the example of preserving agent CPD.

Figure 9 demonstrates that despite the initial acidic environment of the preservative (pH CPD = 5-6) in the control and test in the extracellular medium at the first stage of the study alkaline pH is registered. The appearance of differences in the dynamics of change in

the color of the pH indicator between the control and the test is clearly observed in the subsequent stages of the study. So, against the background of the appearance of hemolysis signs significant decrease of the pH to 7.1-7.2 in the control at the VI stage of the study is registered. On

the contrary, the pH of the extracellular medium remains relatively stable and corresponds to the parameters 7.4-7.5 in the test at the VI stage of the study.

Thus, obtained result, show that change in cytoplasmic pH is both necessary and sufficient for the shape changes of human erythrocytes [35]. The effect of hemolysis inhibition by the method of additive modernization of preservation solutions that adapted to the manufacture process at the VI stage of the study is shown in Figure 10.



Conclusion

As a result of the studies it was found that physiologic solution NaCl which previously was processed by ICNB and added to the preserved of the red blood cells actively inhibits of hemolysis processes of RBCs at the storage stages at a positive temperature. A comprehensive analysis of data revealed the primary mechanisms of the effect modernized of the saline solution on the preserved RBCs. It was established that saline NaCl, which had previously been processed by magnetite nanoparticles (ICNB) had a marked membrane-stabilizing effect, inhibits hemolysis and increasing the sedimentation stability of preserved RBCs. In General, these effects provide the sustainability of the functional activity of preserved RBCs in during storage.

Thus, the first optimistic results were obtained on the way of creation a simple and practical method of additive modernization of preservation solutions that does not

violate the compliance requirements, improves the quality, efficiency and safety transfusion of red blood cells.

References

1. Hess JR (2006) An update on solutions for red cell storage. *Vox Sang* 91(1): 13-19.
2. Högman CF, Meryman HT (1999) Storage parameters affecting red blood cell survival and function after transfusion. *Transfus Med Rev* 13(4): 275-296.
3. Hess JR (2010) Red cell changes during storage. *Transfus Apher Sci* 43(1): 51-59.
4. Zimrin AB, Hess JR (2009) Current issues relating to the transfusion of stored red blood cells. *Vox Sang* 96(2): 93-103.
5. van de Watering L (2011) Red cell storage and prognosis. *Vox Sang* 100(1): 36-45.
6. Wang D, Sun J, Solomon SB, Klein HG, Natanson C (2012) Transfusion of older stored blood and risk of death: a meta-analysis. *Transfusion* 52(6): 1184-1195.
7. Glynn SA (2010) The red cell storage lesion: a method to the madness. *Transfusion* 50(6): 1164-1169.
8. D'Amici GM, Rinalducci S, Zolla L (2007) Proteomic analysis of RBC membrane protein degradation during blood storage. *J Proteome Res* 6(8): 3242-3255.
9. Verhoeven A, Hilarius P, Dekkers D, Lagerberg JW, de Korte D (2006) Prolonged storage of red blood cells affects aminophospholipid translocase activity. *Vox Sang* 91(3): 244-251.
10. Sparrow RL, Veale MF, Healey G, Payne KA (2007) Red blood cell (RBC) age at collection and storage influence RBC membrane-associated carbohydrates and lectin binding. *Transfusion* 47(6): 966-968.
11. Bosman GJ, Lasonder E, Groenen-Döpp YA, Willekens FL, Werre JM, et al. (2010) Comparative proteomics of erythrocyte aging in vivo and in vitro. *J Proteomics* 73(3): 396-402.
12. Hess JR (2012) Scientific problems in the regulation of red blood cell products. *Transfusion* 52(8): 1827-1835.

13. Hess JR, Sparrow RL, van der Meer PF, Acker JP, Cardigan RA, et al. (2009) Red blood cell hemolysis during blood bank storage: using national quality management data to answer basic scientific questions. *Transfusion* 49(12): 2599-2603.
14. Dumont LJ, AuBuchon JP (2008) Evaluation of proposed FDA criteria for the evaluation of radiolabeled red cell recovery trials. *Transfusion* 48(6): 1053-1060.
15. McAteer MJ, Dumont LJ, Cancelas J, Rugg N, Vassallo R, et al. (2010) Multi-institutional randomised control study of haemolysis in stored red cell units prepared manually or by an automated system. *Vox Sang* 99(1): 34-43.
16. Belousov AN (2011) The use of magnetite nanoparticles in applied medicine. *International Journal of Nano Dimension* 2(1): 25-28.
17. Belousov AN, Belousova E Yu (2002) The first steps in discovery new mechanisms of cellular regulation in means by nanotechnology preparations. *X International Conference New Information Technologies in Medicine and Ecology. - Yalta, Gursuf*, Pp: 420-425.
18. Belousov AN, Belousova E Yu (2012) Mechanisms cell regulation by nanotechnology preparations (MCS-B). *BioNanotech Conference & Expo, Santa Clara, CA, USA*.
19. Belousov AN (2000) Effect of magnet controlled sorbent on parameters of acid-base balance of the blood and the processes of glycolysis in erythrocytes. *Pain, anesthesia and intensive care*: 263-265.
20. Belousov AN, Nevzorov VP (1997) Ultrastructure of cells in the kidneys and lungs of rabbits after administration of magnetite // *International collection of scientific papers IV Scientific and Practical Conference on the Creation and Testing of New Drugs, Moscow*, Pp: 77-87.
21. Belousov AN, Nevzorov VP (1997) Ultrastructure of liver cells after administration of magnetite // *International collection of scientific papers IV Scientific and Practical Conference on the Creation and Testing of New Drugs, Moscow* Pp: 71-77.
22. Belousov AN (2009) Spectrum of Application Magnetite Nanoparticles in Medicine. *Nanotech* 2(3): 154-157.
23. Belousov AN (2012) Effect on hemolysis and transport ATPase activity of erythrocytes by means nanoparticales of magnetit controlled sorbent (MCS-B). *Pain, anesthesia and intensive care*. Kiev: 26-28.
24. Belousov AN (2017) Ultrastructure of Hepatic Cells after Intravenous Injection of Magnetite Nanoparticles ICNB. *J Cell Mol Biol* 2(1): 1-4.
25. Belousov AN (2013) Application Magnetite of Nanoparticles (ICNB Preparation) as Magnetically-Resonant Contrasting Means During Visualization of Tumours. *Clean Technology and Sustainable Industries Organization*: 379-381.
26. Belousov AN (2012) Investigation of the influence nanoparticles of magnetite controlled sorbent (MCS-B) on the functional activity of erythrocytes. *Prospects Medicine and Biology*: 94-97.
27. Belousov AN (2014) Myth and Reality Application of Magnetite Nanoparticles as Selective Contrasting Means of the Malignant Tumors in MRI Investigation. *Journal Biomedical Engineering Research* 2(3): 147-152.
28. Belousov AN (2014) Application of Biocompatible Standardized Magnetite Nanoparticles (ICNB) In MRI Investigation of Malignant Tumour. *Journal Nanomaterials & Molecular Nanotechnology*: 17-21.
29. Belousov AN (2014) Nanotechnology and Discovery of a New Factor Which Influences on Permeability of Erythrocytes and Eryptosis. *Journal of Materials Science and Engineering A* 4(11): 367-372.
30. Belousov AN (2017) Reducing of Erythrocytes Destruction by Means of Medicine Nanotechnology (Magnet- Controlled Sorbent brand of MCS-B). *J Cell Mol Biol* 2: 005.
31. Eylar EH, Madoff MA, Brody OV, Oncley JL (1962) The contribution of sialic acid to the surface charge of the erythrocyte. *J Biol Chem* 237(6): 1993-2000.
32. Cook GMW, Heard DH, Seaman GVF (1961) Sialic acids and the electrokinetic change of the human erythrocyte. *Nature* 191(4783): 44-47.
33. Jan KM, Chien S (1973) Role of Surface Electric Charge in Red Blood Cell Interactions. *J Gen Physiol* 61(5): 638-654.

34. Sakota D, Sakamoto R, Yokoyama N, Kobayashi M, Takatani S (2009) Glucose depletion enhances sensitivity to shear stress-induced mechanical damage in red blood cells by rotary blood pumps. *Artificial Organs* 33(9): 733-739.
35. Gedde MM, Davis DK, Huestis WH (1997) Cytoplasmic pH and human erythrocyte shape. *Biophys J* 72(3): 1234-1246.

