



Biosensors Technological Advancement and their Biomedical, Agricultural, Environmental and Food Industrial Applications: A Review

Tariku A^{1*} and Addis S²

¹Microbial Biotechnology Research Program, Ethiopian institute of Agricultural Research, Ethiopia

²Institute of Biotechnology, Addis Ababa University, Ethiopia

***Corresponding author:** Tariku Abena, Microbial Biotechnology Research Program, National Agricultural Biotechnology Research Center (NABRC), Ethiopian institute of Agricultural Research, P.O box 249, Holeta, Ethiopia, Tel: +251909311778; Email: tarikuabena8@gmail.com

Review Article

Volume 8 Issue 3

Received Date: September 01, 2023

Published Date: September 27, 2023

DOI: 10.23880/nnoa-16000263

Abstract

The biosensors are devices that receive the biological message and convert it into a sensible electrical signal. The biosensing involves a combination of biological entities like DNA, RNA, and proteins/enzymes to the electrochemical transducers. Biosensors comprise biorecognition elements including enzymes, antigens, antibodies or nucleic acids that mediate selective biocatalysis or specific binding of analyte and transducers that able to measure the signal. There are several types of biosensors being employed today, such as optical, surface plasmon resonance, enzymes, DNA, Phage, and microbial biosensors. The biosensor technologies have been employed in biomedicine, food safety standards, defense and environmental monitoring. Detection of the lower or higher limits of glucose in the body, microbial invasion in the body and food, heavy metal detection in soil, water and airborne microbes, pesticides in water and soil and various harmful chemicals produced by body, can be easily and timely monitored with high precision using the different types of biosensors. Biosensors can overcome all the limitation of the traditional methods of chemical and microbiological analyses by offering rapid, non-destructive and affordable methods for quality control. Thus, this review paper highlights biosensor and its components, types of biosensors and its application in different disciplines.

Keywords: Bio-recognition Element; Biosensor; Immobilization; Transducers

Abbreviations: SPR: Surface Plasmon Resonance; OPH: Organophosphorus-Hydrolase; OP: Organophosphate Compounds; BOD: Biochemical Oxygen Demand.

Introduction

The biosensors are analytical devices and a hybrid form of physical and chemical sensing technique that converts a biological response into an electrical signal proportional to

the concentration of a specific analytes. In its technical aspect, biosensing is a phenomenon that withholds set of techniques for the production of an accessible detection signal of interaction between biological molecules and another molecule or analyte of interest. Such molecular device that enables the sensing of these molecular interactions is called biosensors [1]. In principle, the biosensors are receptor-transducer based tool which could be used for interpreting the biophysical or biochemical property of the medium. The

presence of biological/organic recognition element which enables the detection of particular biological molecules in the medium distinguishes biosensors apart from other types of sensors [2].

The biosensor was first discovered by Clark, et al. [3] for measuring the amount of glucose in biological samples by electrochemical detection of oxygen or hydrogen peroxide using immobilized glucose oxidase electrode [4]. Since then, biosensor advanced both in technology and applications with innovative approaches involving electrochemistry, nanotechnology to Bioelectronics [4].

The sensor is composed of two parts, i.e., receptor and transducer. Receptor receives the physical/chemical stimulus and transmits this information in the form of electrical energy while the transducer performs the function of transducing this energy into a valuable analytical signal which can further be analyzed and presented in an electronic form [5]. In the utilization of biosensor technology, bio recognition elements include immobilized enzymes, antigens, antibodies and DNA/RNA [6]. Biorecognition elements are tightly bound onto the physical-chemical transducer by physical or chemical immobilization methods. In general, there are five groups of transducers such as: electrochemical, optical, mass-based, thermal and magnetic biosensor [7,8]. Improving technologies allow the development of novel, advanced and new designed transducers [9].

Biosensors have several practical applications in areas including biochemical, medical, environmental, food, industrial, biosecurity or pharmaceutical analysis and personal diagnostics which are based on connection of biological element or molecule with biological activity toward measured analyte onto surface of the used transducer [10,11]. Therefore, this review presents the technological advancement, construction methods, types and application of biosensor.

Components of Biosensors

The components of biosensors are broadly categorized into two such as, analytical devices consisting of biological molecule (biorecognition element) and physico-chemical transducer providing measurable signal working as a physical sensor [12]. Biorecognition element mediates selective biocatalysis or specific binding of analyte. Enzyme, antigen, antibody or nucleic acid usually belongs to one of recognition elements and the specificity of measured system depends on it [6].

Biorecognition Elements: The biorecognition elements can be divided into two categories such as: (a) biocatalytical receptors like enzymes, whole cells, cell organelles, tissues and whole microorganism and (b) bioaffinity receptors like

(antibodies, cell receptors or nucleic acids) [7,13]. Enzymes are very common biorecognition elements mostly for their simple and well-known construction, high sensitivity, availability, satisfactory limits of detection and affordability. For instance glucose oxidase based glucose biosensors belong to the one of the mostly used enzymatic biosensors, and it was also used in the first biosensor construction [7,9,13].

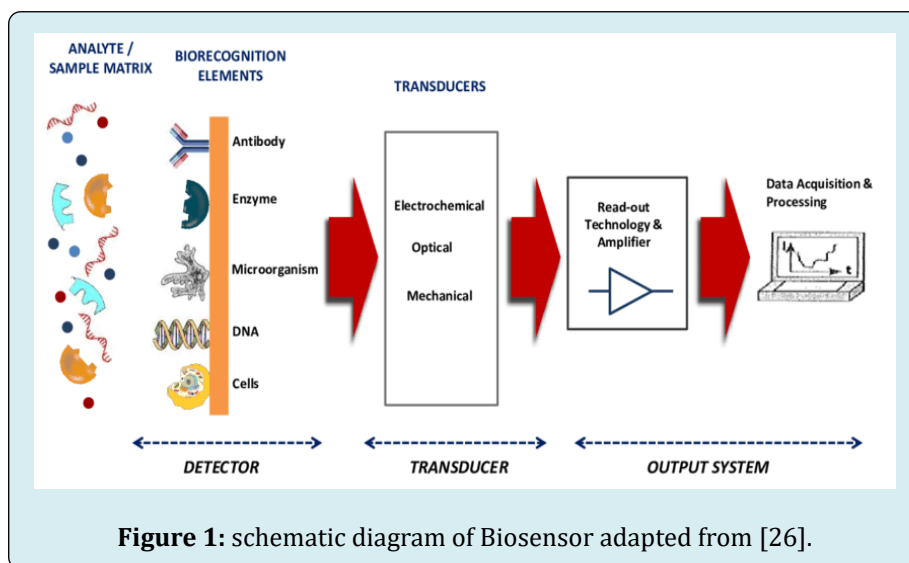
Antibodies have structure of immunoglobulins that consists two polypeptidic heavy and two polypeptidic light chains linked by disulfide bonds. Based on heavy chains differences there are five groups of antibodies: IgG, IgM, IgA, IgD and IgE [14]. Antibodies have been used as biorecognition elements due to their broad spectrum of application such as, high specificity, sensitivity, selectivity and strong antigen-antibody interactions [8]. The biosensors having embedded antibody or working on antibody-antigen interaction are called immunosensors [15]. Polyclonal or recombinant antibodies that secreted by multiple plasma cells, monoclonal antibodies secreted by single clonal lineage and recombinant antibodies produced during recombinant engineering by gene manipulation are usually used in clinical practice and diagnosis [14]. The monoclonal antibodies that produced by fusing an immortal myeloma cells with spleen cells have high specificity of antibody-antigen binding, homogeneity and production in large quantities. Antigens or antibodies can be labeled by enzymes, fluorescent or electrochemical compounds, radionuclides, or avidin-biotin complex because of inability of antibody-antigen complex to generate proper signal for optical and electrochemical transduction [15].

On the other hand, use of mass-based transducers converting mechanical deformation and voltage to measure mass or viscoelastic effects enables direct detection of arising bound without necessity of labeling. The selectivity of measured system is determined by two identical very specific antigen binding sites on the molecule of immunoglobuline [16]. Deoxyribonucleic acid (DNA) contains two antiparallel complementary polynucleotide strands consisted of purine and pyrimidine nucleotides linked by hydrogen bonds used as a biorecognition element is integrated on transducer surface as whole pre-synthesized probe (sequence of polynucleotide chain containing tens of nucleotides) or each base is immobilized on transducer surface individually [17,18]. DNA sensors (also called genosensors) are based on specific nucleic acid-analyte binding process like hybridization between targeting DNA and complementary probe and signal from hybridization is measured [17,19].

Transducers: Transducers vary according to their construction, principle and possibility and frequency of their application. Electrochemical transducers have major role in diagnostic, optical transducers have important influence on research, but thermal, magnetic and mass-based transducers have not gained great clinical impact and nowadays they are

use rarely. The electrochemical transducers are based on monitoring of electric potential or electric current changes caused by electron or ions altering during biochemical reaction of biorecognition element (mostly enzyme) with analyte [20,21]. The enzyme transforms substrate to electroactive product creating measurable signal for electrochemical transducer [22]. Amperometric transducers are based on measuring of current corresponding with amount of electroactive substance produced during chemical reaction in solution. Constant potential is set on electrode so the measured current response to concentration of determined

substance [23]. The potentiometric transducer that measures the change between two electrodes is suitable for detecting of very low concentration or presented mass of analyte due to logarithmic device response on analyte concentration [23]. The benefits of electrochemical transducers includes: Low cost, high sensitivity and measuring turbid samples [21,24]. Electrochemical biosensors have a multiple discipliner applications in biotechnology, food industry, health care, medicine or environmental monitoring because they are high sensitive and selective electroanalytical device [25].



Immobilization Techniques for Biosensors Preparation

There are two types of immobilization techniques in general: physical and chemical immobilization. Selection of a suitable immobilization technique is one of crucial steps of sensor preparation because the possibility of biorecognition element inactivation caused by choosing inappropriate immobilization method is very high and Selection of more appropriate method depends on nature of the chosen biorecognition element, used transducer, physico-chemical conditions and properties of analyte [27,28].

Physical Immobilization Techniques: physical immobilization is based on binding of biological molecules (most often enzymes) to transducer surface without creation of any chemical bonds. Physical immobilization technique includes: physical entrapment, microencapsulation, adsorption and sol-gel techniques [29-31]. Physical entrapment is a method based on embodying biorecognition elements in three-dimensional matrices and it consisted of polydimethylsiloxane, a photopolymer, gelatin, alginate, cellulose acetate phthalate, modified polypropylene and polyacrylamide membranes or a carbon paste can be named as examples of entrapping matrices [30,31].

Electropolymerization: Electropolymerization is immobilization of biorecognition element mostly enzyme on electrode surface under applied current or potential in aqueous solution containing both biomolecule and monomer molecule (such as aniline, pyrrole or thiophene). Conducting polymerized film with precise spatial resolution over surfaces where the bioelement is entrapped inside is created [32,33].

Physical Adsorption: Physical adsorption technique involves attachment of bio recognition element to the inert material by van der Waals forces, hydrophobic interaction as well as by hydrogen bonds [34]. This method has a lot of advantages such as the simplicity, great variety of materials and it does not require chemical modification of biological components. Despite that the clinical application may be limited by the possibility of biomolecule activity loss [7,35].

Microencapsulation: It is based on low temperature forming of solid glass-like transparent film via hydrolysis and condensation of precursor alkoxide where bioelements are encapsulated. Extraordinariness of sol-gel membrane lies in its thermal and chemical stability, simplicity of preparation and possibility of large amount of biomolecule entrapment [27].

Entrapping of Biomolecule into Membrane: It is based

on physical, either hydrophilic or hydrophobic binding of a biomolecule on inert membranes that provide close contact between biomaterials and the transducer. Types of membranes used include cellulose acetate, polycarbonate, collagen, and Teflon [36,37]. Carbon paste consists of graphite powder and pasting liquid and it makes up ideal substance connecting the entrapped biorecognition element to surface of transducer. It is usually used with electrochemical transducer [30].

Chemical Immobilization Techniques

Chemical immobilization is based on creation of chemical bonds between functional group of biorecognition element (side chains unnecessary for its catalytic activity) and surface of the used transducer. Chemical bonds are mostly forming on activated transducer surface carrying out by chemical reagents (such as glutaraldehyde or carbodiimide) or they are created directly because of pre-activated membrane applied on transducer surface. Covalent binding, and covalent cross-linking belongs to the chemical immobilization techniques [18,28]. There are two types of chemical immobilizations such as: Covalent binding and cross-linking

Covalent Binding: Covalent binding is a process where

biorecognition element receives firm bond to either surface or inner cavity of membrane. It is the most widely used type of enzyme immobilization technique [38,39]. The binding process is based on reaction between functional protein groups (usually side chain of amino acids) of biorecognition element and reactive groups of transducer/membrane matrix surface. Covalent binding provides increased lifetime stability and strong and effective bonding and it includes chemical adsorption (also called chemisorptions) and activation of carboxylic or amino groups [38].

Cross-Linking: It is an immobilization process based on covalent binding between biorecognition elements or between biorecognition element and functionally inert protein (for example bovine serum albumin). It leads to formation of three dimensional aggregates bonded via multifunctional linker molecule such as glutaraldehyde, glyoxal and hexamethyldiamine to the transducer surface [27,39]. Process of cross linking requires optimal conditions such as pH, temperature and ionic strength to allow shorter response time, stronger attachment and higher catalytic activity of enzymes [39]. Despite many advantages poor stability and partial denaturation of protein structure may limit application of cross linking immobilization [18,27].

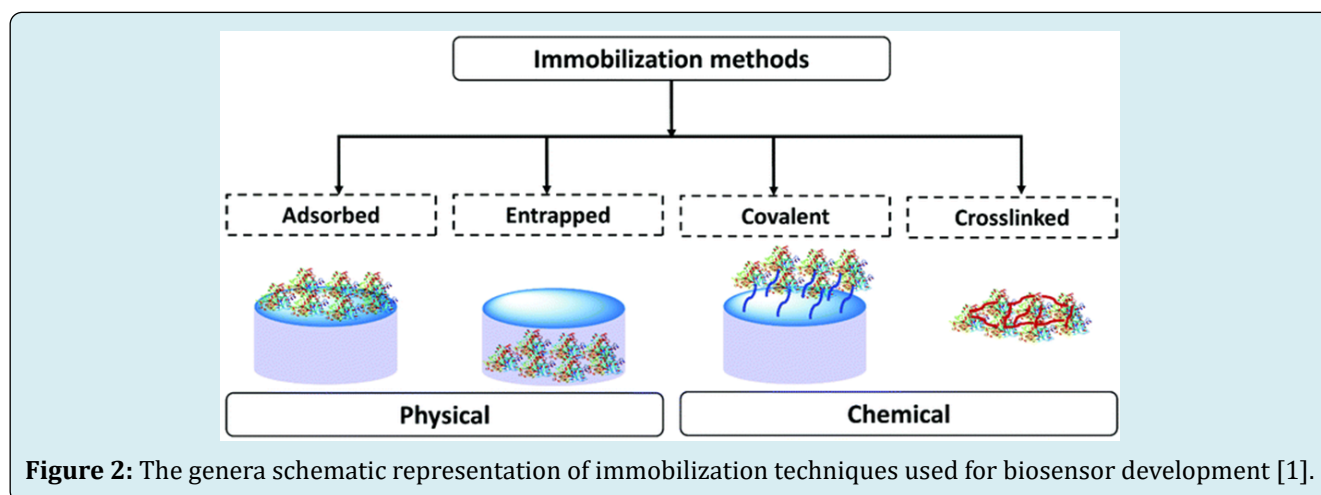


Figure 2: The genera schematic representation of immobilization techniques used for biosensor development [1].

Types of Biosensors

Biosensor devices can be categorized as electrochemical, optical/ visual, polymers, silica and glass, nanomaterials, genetically encoded biosensors and microbial biosensors [40].

Electrochemical Biosensors: Electrochemical biosensors are analytical devices prepared by modifying the surface of metal and carbon electrodes using biomaterials and that transduce biochemical interactions (enzyme-substrate reaction and antigen-antibody) to detectable electrical signals [41]. The electrochemical sensors became extremely desirable for early detection of diseases [42]. Electrochemical

biosensors are employed to detect the amount of antioxidants and reactive oxygen species in physiological systems [4,43]. Major application in this line is the detection of uric acid as primary end product of body fluid purine metabolism, which provide diagnostic tool for various clinical abnormalities or diseases [44]. Even though electrochemical sensor based measurement of uric acid oxidation for glucose quantification seems ideal, similarity of uric acid oxidation with that of ascorbic acid poses major experimental hurdle to develop highly sensitive electrochemical biosensor [4]. In response of this problem, scientists have developed amperometric detection-based biosensor which has the ability to measure both reduction and oxidation potentials

[44]. Electrochemical biosensors have been utilized for hormone detection [4]. Another potential area of technology development in biosensors is that it can be developed from nucleic acids that used for miRNA detection [4,45].

Environmental monitoring is another important aspect wherein biosensor technology is required for rapid identification of pesticidal residues to prevent health hazards [46,47]. Even though traditional methods including: high-performance liquid chromatography, capillary electrophoresis and mass spectrometry are effective for the analysis of pesticides in the environment, they have drawbacks like complexity, time-consuming procedures, requirement of high-end instruments and operational capabilities [4,47]. Some enzymatic biosensors were utilized to understand the physiological effect of pesticides in the environment, food safety, and quality control [4,48]. For example, acetylcholinesterase (AChE) inhibition-based biosensors were developed [48]. It is important to place special emphasis for selection of receptors for biosensor development, the use of different transduction techniques and fast screening strategies for applications of biosensor in food, and environmental safety [4].

Optical/Visual Biosensors: An optical biosensor is a compact analytical device containing a biorecognition element (enzymes, antibodies, antigens, receptors, nucleic acids, whole cells and tissues) integrated with an optical transducer system [49]. The most commonly used optical biosensors include: surface plasmon resonance (SPR)-based biosensors (SPR imaging and localized SPR), evanescent wave fluorescence and bioluminescent optical fibre biosensors, as well as interferometric, ellipsometric and reflectometric interference spectroscopy and surface-enhanced Raman scattering biosensors [49]. Optical biosensors are advantageous than that of traditional analytical techniques since it is highly sensitive, specific and cost effective [50]. Optical biosensors have been utilized in clinical diagnostics, drug discovery, food process control, and environmental monitoring [51]. Optical biosensor also utilized for ethanol determination in fermented beverage samples [52]. Recently, CRISPR-Cas12a powered visual biosensor with a smartphone readout developed for ultrasensitive and selective detection of SARS-CoV-2 [53].

Nanomaterials-Based Biosensors: The nanomaterials involved in the development of biosensor includes: gold and silver nanoparticles, quantum dots, mesoporous silica nanoparticles, carbon nanomaterials, and hybrid nanocomposites [54,55]. They are used either as carriers for immobilizing biorecognition elements, or as labels for signal generation, transduction and amplification [54].

The nanomaterials-based detection technology showed advancement over the conventional methods in selectivity and sensitivity and its physical, chemical, electrical and optical properties makes it suitable for the application in biosensors [56]. The Nanomaterials Like semiconductor quantum dots and iron oxide nanocrystals have been applied to understand the tumor microenvironment for therapeutics and also for the delivery of nano-medicine [57,58]. Recently, rapid and unamplified nanosensing technology has been developed for detection of SARS-CoV-2 RNA in human throat swab specimens [59].

Genetically Encoded Biosensors: Understanding the biological process and different metabolic pathways inside the cell needs to use tagged biosensor that developed by using genetically encoded or synthetic fluorescence [4]. Genetically encoded sensors are engineered fluorescent proteins that have been developed for ions metabolites, redox potential, biophysical processes [60]. The invention of genetically encodable green fluorescent protein has made remarkable progress in terms of optical probe design and efficiency [61]. Förster resonance energy transfer (FRET)-based biosensors have been developed for visualizing cGMP, cAMP, and Ca²⁺ in cells [62]. Now a days, various genetically encoded biosensors have been developed to sense aromatic monomers, including phenolic compounds obtained through direct lignin depolymerisation and other potential intermediates relevant to biomass valorization [63]. According to Lin, et al. [64] genetically encoded fluorescent biosensors used to investigate kinase signaling in cancer cells and tumor tissue sections and enabled visualization of biological processes and events directly in situ. Considering the advent of in vivo imaging with small molecule biosensors, a better understanding of cellular activity and many other molecules ranging from DNA, RNA, and miRNA have been identified [65]. Now the transformation in this field requires whole genome approach using better optical based genetic biosensors.

Microbial Biosensors: Microbial biosensor is an analytical device which developed by immobilization of microorganism(s) onto physical transducer to generate a measurable signal proportional to the concentration of analytes [66]. Microbial biosensors have been applied in numerous fields including medicine, environmental monitoring, defense, food processing and safety [67,68]. Microbial biosensors have been integrated with many recently developed micro/nanotechnologies and applied to a wide range of detection purposes [69]. In future, these microbial biosensors will have wider applications in monitoring environmental metal pollution and sustainable energy production [4,70].

Sl. No.	Type	Principle	Applications	Bibliography (review/original article)
1	Glucose oxidase electrode based biosensor	Electrochemistry using glucose oxidation	Analysis of glucose in biological sample	[3]
3	Uric acid biosensor	Electrochemistry	For detection of clinical abnormalities or diseases	[71]
4	Acetylcholinesterase inhibition-based biosensors	Electrochemistry	Understanding pesticidal impact	[48]
7	Hydrogel (polyacrylamide)- based biosensor	Optical/visual biosensor	for the detection of biological warfare agents (BWAs)	[71]
8	Silicon biosensor	Optical/visual/fluorescence	Bioimaging, biosensing and cancer therapy	[72,73].
10	Nanomaterials-based biosensors	Electrochemical or optical/visual/fluorescence	For multifaceted applications including biomedicine, for example diagnostic tools	[55,74,75]
11	Genetically encoded or fluorescence- tagged biosensor	Fluorescence	For understanding biological process including (monitoring kinase activity, for protein-DNA interaction analyses, real-time assays of motor proteins).	[40,61,76]
12	Microbial fuel cell- based biosensors	Optical	To monitor biochemical oxygen demand and toxicity in the environment and heavy metal and pesticidal toxicity	[70,76]

Table 1: List of some potential biosensors with principle and applications.

Adapted with slight modification of the review paper published by Vigneshvar, et al. [4].

Application of Biosensors

The most recent application of biosensor includes: agriculture, biomedicine, food, environmental and defense fields.

Biomedical Applications: Biomedical application of biosensor includes: measuring of blood glucose level, genetic diagnostics and DNA encoding, tissue/cell engineering and measuring the H_2O_2 amount. Little success is also achieved with few potential molecules for novel therapeutic, antimicrobial, and drug delivery. Invention on this line leads to discovery of electrochemical biosensors as reliable analytical devices for pathogen detection of avian influenza virus in the complex matrices [77]. More recent report revealed potential applications of affinity-based biosensors in sportmedicine and doping control analysis [78].

Measuring of Blood Glucose Level: The demand for use of glucose sensing technologies has grown due to the enhancing number of diabetics patients every year. The enzymes that

are effectively utilized at large scale for glucose detection are glucose oxidases (G-ox) and glucose dehydrogenases [79,80].

Diagnose of Infectious Diseases: Biosensors are being used in the medical field to diagnose infectious diseases. A novel biosensor based on hafnium oxide (HfO_2), has been used for early stage detection of human interleukin (IL)-10 [81]. Interaction between recombinant human IL-10 with corresponding monoclonal antibody is studied for early cytokine detection after device implantation. Fluorescence patterns and electromechanical impedance spectroscopy characterize the interaction between the antibody-antigen and bio-recognition of the protein is achieved by fluorescence pattern. Chen and co-workers applied HfO_2 as a greatly sensitive bio-field-effect transistor [82]. HfO_2 biosensor has been functionalized for antibody deposition with detection of a human antigen by electrochemical impedance spectroscopy.

The biggest dilemma faced today is of heart failure with about one million people suffering from it. Techniques for detection of cardiovascular diseases include immunoaffinity column assay, fluorometric, and enzyme-

linked immunosorbent assay [83,84]. Chen and co-workers synthesized ultra-sensitive sensor based over nuclease mediated highly targeted recycling of DNAzyme for the electrochemical detection of oral cancer from the saliva secretions. With this sensor, they quantified up to the 0.02 fM of the targeted DNA and detected gene mutation up to the single basepair mismatch. Along with the operation and maintenance conveniences and low engineering cost make this biosensor a promising candidate for oral cancer detection at the commercial level [85]. Yang and coworkers devised an altered graphene electrode which possesses the ability to chemically bind with ssDNA and generate Voltametric signal for its counter analogue DNA for detection [86].

Measuring H₂O₂ Content: In humans the H₂O₂ content is a direct indicative of the oxidative stress of cell or hypoxic conditions of tissues. To know or to measure the amount of H₂O₂, various analytic techniques like titration, electrochemistry and photocatalysis could be utilized [87]. The instability of hydrogen peroxide in any biological system makes it highly injurious and cytotoxic for humans, plants, animals as well as bacteria [88]. In the field of tissue engineering, a generally employed method for H₂O₂ quantification are mostly electrochemical in nature and poses several difficulties (poor detection, low sensitivity, less portability and applicability issues on the organic system) to the user [11]. Enzyme based biosensing technique which is the recent finding have quite high stability and accuracy [89,90].

Application in Food Industry

Biosensor applications in food industry can categorized in to two main groups: detection of food borne pathogens and detection of chemical contaminants. To ensure the safety of processed foodstuffs, specific methods have been developed by the food industries to detect and identify chemicals or biological agents that cause food spoilage and responsible for the spread of some serious health related problems [91]. Biosensors being target specific, highly sensitive and quickly responsive and used to determine the chemical activities that lead to the food spoilage. The enzyme substrate interaction or antibody-antigen complex that can be easily detected is the fundamental determinant factors of biosensors [92]. The common types of biosensor employed in food industry are enzyme-based biosensors and immunosensors [92].

Food Borne Pathogens Detection: The microbes that involved in food spoilage mainly include bacteria and fungi that lead to the spread of serious health hazards.

Bacterial Monitoring: Common food spoiling bacterial species that cause health problem are: E. coli strain 0157:H7, Listeria monocytogenes, campylobacter and salmonella. These bacteria are common problems faced by the food industries as they reduce the consumer demands of the food. Food industries had been striving to avoid the problem

relate to food poisoning, by timely detection and removal of this bacterium. For this different type of biosensors like Piezoelectric biosensors for detection of monitoring Salmonella monoclonal antigen-antibody interaction quickly and easily, Fibre-optic biosensors to monitor the presence of Listeria monocytogenes [93]. Amperometric biosensors based on enzymatic system had been successfully used for the detection of E. coli 0157:H7 [94].

Fungal Pathogens Detection: Fungal pathogens are the common food deteriorating microorganisms that cause severe health problems. The common food contaminating fungi includes: Botrytis sp., Aspergillus, Colletotrichum and many other fungal species. Due to the remarkable specificity, reduced costs and easy and quick monitoring through biosensors and there are reports that fungal mycotoxins can be detected using optical SPR biosensors [95,96].

Biosensors for Determination of Chemical Contaminants: Chemical contaminant that causes food spoilage including pesticides, fertilizers, heavy metals, food additives and antibiotics

Biosensors for Contaminant Residues and Pesticides Detection: The presence of pesticide residues and metabolites in food, water and soil currently represents one of the major issues [97]. Due to the limitations in conventional methodologies, the development of biosensors for direct and indirect pesticide detection is of particular interest. The enzymatic biosensors like cholinesterase (AChE, BChE), organophosphorus-hydrolase (OPH), and urease are used for the detection of pesticides, fertilizers, and heavy metals [98]. Analytical devices, based on OPH and cholinesterase inhibition, have been widely used for the detection of carbamates and organophosphate compounds (OP) [99,100]. Immobilized cells of Flavobacterium sp have been used for the detection of methyl parathion. Whole cells of Flavobacterium sp. Flavobacterium sp. have the enzyme organophosphorus hydrolase, which hydrolyzes the methyl parathion into detectable product p-nitrophenol [101]. Immunosensors have great potential for monitoring herbicides in drinking water, the detection of polychlorinated terphenyls and atrazine [102].

Biosensors Used for the Detection of Heavy Metals: Eating foods containing residues of heavy metals cause several health problems like: cardiovascular and respiratory problems, infertility, irritations, inhibition of some hormonal activities, malfunction of the principal organs, and death. Devices have been designed to determine the concentration of heavy metals such as arsenic, cadmium, mercury, and lead, in water and soil samples. These devices incorporate genetically modified microorganisms and enzymes such as urease, cholinesterase, glucose oxidase, alkaline phosphatase, ascorbate oxidase and peroxidase [103]. Immobilized algae inside bovine serum albumin membranes have made a network structure with glutaraldehyde vapors deposited on interdigitated conductometric electrodes and local

conductivity variations caused by algae alkaline phosphatase and acetylcholinesterase activities could be detected [104]. In addition, it is possible to know the presence of cadmium through detection of the inhibition of the urease enzyme by using fiber optics biosensor made from whole cells of *Bacillus badius* with phenol red as an indicator that can sense down to 0.1 g/l of cadmium in milk.

Biosensors as Indicators of Product Acceptability: Food quality involves nutritional and organoleptic characteristics such as freshness, appearance, taste and texture. The food sensory basis is essential for the industry [105]. During storage, compounds that provide aroma and abnormal flavors indicating in most cases microbial growth and insufficient food safety [98].

Biosensors that use whole cells or enzymes have been used for the detection of alcohol [106]. Biosensor with immobilized enzymes: alcohol oxidase, alcohol peroxidase and a chromogen, have been used detect injuries caused by low O₂ in lettuce, cauliflower, broccoli and cabbage lightly processed and packed in a modified atmosphere [107]. This biosensor could also be used to monitor ethanol during the storage of apples in a controlled atmosphere, the decay in potato tubers [108]. Similar research has been done to detect organic acids and sugars as indicators of fruit and vegetables maturity [109]. The co-immobilized biosensor containing alcohol oxidase and glucose oxidase have been used to determine glucose and ethanol [110]. Measurements are based on monitoring decrease in current on reduction potential of tetrathiafulvalene (at 0.1V vs. Ag/AgCl) by using a cyclic voltammetry method and correlations between decreases in biosensor responses and glucose oxidase or alcohol oxidase activity were monitored.

Multiple compounds giving unpleasant flavors and aromas in foods can be potentially detected by biosensors. Varelas and his co-workers developed a biosensor system based on a bioelectric recognition assay for detection of 2, 4, 6-trichloroanisole, a compound that causing considerable losses to the wine industry [111]. Biosensors technology for substance detection significantly reduces analysis time, and improves specificity, reliability and test sensitivity. These properties allow for real time decision making during food processing. A listing of biosensors used to evaluate food composition is presented by Serna, et al. [98,112].

Environmental Applications of Biosensors

The main classes of bioreceptor elements that are applied in environmental analysis are whole cells of microorganisms, enzymes, antibodies and DNA. Additionally, in the most of the biosensors described in the literature for environmental applications electrochemical transducers are used [113]. For environmental applications, the main advantages offered

by biosensors over conventional analytical techniques are the possibility of portability, miniaturization, work on-site, and the ability to measure pollutants in complex matrices with minimal sample preparation. The major pollutant that can be successively detected and removed using biosensors includes heavy metals, polychlorinated biphenyls, pesticides, Biochemical Oxygen Demand (BOD) and nitrogenous compounds.

Heavy Metals Detection and Monitoring: Heavy metals are non-degradable compounds that affect humans' health and their hyper-accumulation leads to various inappropriate health conditions [114]. The metal contaminants most commonly observed in the environment are: lead, chromium, zinc, mercury, cadmium and copper [115]. There are various types of biosensors have been utilized in measuring and monitoring toxic heavy metals.

Bacteria-based cell biosensors require the use of genes that resist certain types of heavy metals like copper, mercury, tin cobalt etc [116]. The bacterial cell biosensors interact with heavy metals by their cytoplasm that is based on the conjugation of some luminescent proteins like luciferin, with those genes that resist heavy metals [117].

Enzyme-based biosensors have also provided promising results in that regard have been used for the detection of the toxic levels of different heavy metals. These biosensors work by inhibition by metal ions on various kinds of enzymes, these inhibitions are then monitored by using different types of biosensors with HIGH specificity. For example, Amperometric biosensors were used for the detection of inhibition of mercury ions (Hg⁺²) by urease enzyme [117]. Inhibition of cobalt, nickel, mercury, gold and lead with same urease enzyme lead to the monitoring of the toxic levels using fibre optic sensors [118].

Biochemical Oxygen Demand (BOD): The BOD is the amount of molecular oxygen (O₂) that consumed by microorganisms during break down of organic compounds in waste water [79]. Nakamura, et al. [75] developed a system for measuring BOD from cells of recombinant *Escherichia coli* with *Vibrio fisheri* genes lux AE. An optical biosensor for parallel multi-sample determination of biochemical oxygen demand in wastewater samples has been developed [117]. The biosensor monitors the dissolved oxygen concentration in artificial wastewater through an oxygen sensing film immobilized on the bottom of glass sample vials.

Biosensors for Pesticides Detection: As defined by the EPA, pesticides are any substance or mixture of substances intended for preventing, destroying, repelling, or lessening the damage of any pest [119]. Pesticides are the most abundant, present in water, atmosphere, soil, plants, and food. The organophosphates that used as insecticides (pesticides) causes different negative impacts like changing soil fertility, damaging beneficial insects and microbes in soil and loss

of biodiversity. To measure toxic levels of these pesticides in soils and in water nanotechnological sensors have been developed. The nanotechnology helped to develop enzymatic biosensors by immobilizing the enzymes on different macromolecules. The common example of enzyme biosensor that has been used as detectors of organophosphates is acetylcholinesterase sensors which work by inhibiting acetylcholinesterase activity [81].

Biosensors in Defense

Advances in science through the 19th century allowed more severe and extensive use of chemical and biological toxins (CBTs) as weapons during World Wars I and II [120]. Episodes of anthrax attacks in the United States (1984 and 2001) and the Sarin attacks in Japan (1995), and in Syria (2013) (The New York Times, Dec. 28, 2013) are some of the tragic events happened in the past that have caused intense fear among civilians.

In response to the destructiveness and the fear associated with such conceivable attacks or accidents, global efforts have focused on developing biosensor technologies to detect environmental CBTs. Development in biosensors for the detection of biological warfare agents includes bacteria, virus, and toxins is often attempted using various devices of biosensors such as: electrochemical, nucleic acid, optical and piezo electric, which will have immense applications in military and health as well as defense and security. There are several biosensors applied in the field of defense now a day. The label-free cell-based electric impedance biosensor technology shows good correlation with standard label-based cytogenetic assays, and is very rapid [121,122]. Optical biosensors that characterize changes in the refractive index (resonant waveguide grating; RWG and surface Plasmon resonance; SPR) and scattering of incident light (Raman spectroscopy; RS), have shown great success as label-free cell-based optical sensor technologies. Biosensor and immunosensor technologies based on SPR and RWG have been used extensively for the rapid detection of CBT [123,124].

Conclusion

Biosensor that composed biorecognition element and transducers is a rapidly growing field encompassing various fields like medicine, agriculture, food industry, environmental science and defense. Now a days various types of biosensors have devised, such as electrochemical, optical, genetic encoded, microbial and nano material based which will have immense applications. Biosensors application in medical field is highly advanced especially in the area of medical diagnostics. In the food industry quality control is a major thrust area, the need for fast methods to

monitor the quality of food like freshness, flavors and aromas is urgent [125-134]. In agriculture and environmental science: Pesticides, fertilizers and heavy metals residues can be quickly detected in small quantities with biosensors, facilitating in situ implementation in pre- and post-harvest processes. Conventional methods are expensive, time consuming and labour intensive. Development of efficient sensors will not only speed up the process but will be also cost effective. The advances in detection techniques have allowed the fabrication of rapid and user-friendly advanced biosensor devices imperative for chemical and biological defense. Advanced biosensors enable the label-free and cell-based detection of toxins and the response of the cell and organism to toxins. Biosensor is an interdisciplinary field involving many areas; research in genetic engineering, material science, microfabrication and nanofabrication will enhance the development of suitable sample preparation steps, such as immobilization, extraction and concentration. Future sensors developments must focus on provide multi-analyte detection combined with signal transmitters for remote sensing and modify these biosensing elements to enhance them to the extent that would be able to detect even most dangerous diseases like the viral diseases (HIV, Ebola, Crimean-Congo Virus, Rabies and COVID-19) and can also be employed for bioremediation of pollutants [135-143].

Authors' Declaration

Authors' contributions
Writing - Original Draft, Conceptualization, Investigation, and text writing and editing: Tariku Abena

Funding

No funding has been obtained for this work.

Consent for Publication

Not applicable.

Competing Interests

Authors declare no competing interests.

References

1. Ibraheem A, Campbell RE (2010) Designs and applications of fluorescent protein-based biosensors. *Current Opinion in Chemical Biology* 14(1): 30-36.
2. Vigneshvar S, Sudhakumari CC, Senthilkumaran B, Prakash H (2016) Recent Advances in Biosensor Technology for Potential Applications – An Overview. *Front Bioeng Biotechnol* 4: 11.

3. Clark LC Jr, Lyons C (1962) Electrode systems for continuous monitoring in cardiovascular surgery. *Ann N Y Acad Sci* 102: 29-45.
4. Varelas V, Sanvicens N, Pilar-Marco M, Kintzios S (2011) Development of a cellular biosensor for the detection of 2,4,6-trichloroanisole (TCA). *Talanta* 84(3): 936-940.
5. Geng T, Morgan MT, Bhunia AK (2004) Detection of low levels of *Listeria monocytogenes* cells by using a fiber-optic immunosensor. *Applied and Environmental Microbiology* 70(10): 6138-6146.
6. D'Orazio P (2003) Biosensors in clinical chemistry. *Clinica Chimica Acta* 334(1-2): 41-69.
7. Ma L, Yin L, Li X, Chen S, Peng L, et al. (2022) A smartphone-based visual biosensor for CRISPR-Cas powered SARS-CoV-2 diagnostics. *Biosensors and Bioelectronics* 195: 113646.
8. Lin W, Mehta S, Zhang J (2019) Genetically encoded fluorescent biosensors illuminate kinase signaling in cancer. *Journal of Biological Chemistry* 294(40): 14814-14822.
9. Tang H (2006) A new amperometric method for rapid detection of *Escherichia coli* density using a self-assembled monolayer-based bienzyme biosensor. *Analytica Chimica Acta* 562(2): 190-196.
10. He L, Toh CS (2006) Recent advances in analytical chemistry—A material approach. *Analytica Chimica Acta* 556(1): 1-15.
11. Oldach L, Zhang J (2014) Genetically encoded fluorescent biosensors for live-cell visualization of protein phosphorylation. *Chem Biol* 21(2): 186-197.
12. Peng F, Su Y, Zhong Y, Fan C, Lee ST, et al. (2014) Silicon nanomaterials platform for bioimaging, biosensing, and cancer therapy. *Acc Chem Res* 47(2): 612-623.
13. Velusamy V, Arshak K, Korostynska O, Oliwa K, Adley C (2010) An overview of foodborne pathogen detection: in the perspective of biosensors. *Biotechnology Advances* 28(2): 232-254.
14. Hanko M, Bruns N, Tiller JC, Heinze J (2006) Optical biochemical sensor for determining hydroperoxides in nonpolar organic liquids as archetype for sensors consisting of amphiphilic conetworks as immobilisation matrices. *Analytical and Bioanalytical Chemistry* 386(5): 1273-1283.
15. Conroy PJ, Hearty S, Leonard P, O'Kennedy RJ (2009) Antibody production, design and use for biosensor-based applications. *Seminars in Cell & Developmental Biology* 20(1): 10-26.
16. Filip J, Tkac J (2014) The pH dependence of the cathodic peak potential of the active sites in bilirubin oxidase. *Bioelectrochemistry* 96: 14-20.
17. Nie S, Xing Y, Kim GJ, Simons JW (2007) Nanotechnology applications in cancer. *Annu Rev Biomed Eng* 9: 257-288.
18. Diculescu VC, Paquim AMC, Brett AMO (2005) Mini-Review: Electrochemical DNA Sensors for Detection of DNA Damage. *Sensors* 5(6): 377-393.
19. Sadoine M, Ishikawa Y, Kleist TJ, Wudick MM, Nakamura M, et al. (2021) Designs, applications, and limitations of genetically encoded fluorescent sensors to explore plant biology. *Plant Physiology* 187(2): 485-503.
20. Sánchez J, Jiménez S, Navarro R, Villarejo M (2009) Patógenos Emergentes en la Swimming *Escherichia coli*. *Journal of Bacteriology* 189: 1756-1764.
21. Das AP, Kumar PS, Swain S (2014) Recent advances in biosensor based endotoxin detection. *Biosensors and Bioelectronics* 51: 62-75.
22. Kuswandi B, Irmawati T, Hidayat MA, Ahmad M (2014) A simple visual ethanol biosensor based on alcohol oxidase immobilized onto polyaniline film for halal verification of fermented beverage samples. *Sensors* 14(2): 2135-2149.
23. Pavlov V, Xiao Y, Willner I (2005) Inhibition of the acetylcholine esterase-stimulated growth of Au nanoparticles: nanotechnology-based sensing of nerve gases. *Nano Letters* 5(4): 649-653.
24. Vadivambal R, Jayas D (2007) Changes in quality of microwave-treated agricultural products: a review. *Biosystems Engineering* 98(1): 1-16.
25. Mello LD, Kisner A, Goulart MO, Kubota LT (2013) Biosensors for antioxidant evaluation in biological systems. *Comb Chem High Throughput Screen* 16(2): 109-120.
26. Dalkiran B, Esra Erden P, Kılıç E (2017) Amperometric biosensors based on carboxylated multiwalled carbon nanotubes-metal oxide nanoparticles-7,7,8,8-tetracyanoquinodimethane composite for the determination of xanthine. *Talanta* 167: 286-295.
27. Mello LD, Kubota LT (2002) Review of the use of biosensors as analytical tools in the food and drink industries. *Food Chemistry* 77(2): 237-256.

28. Pribyl J, Hepel M, Halámek J, Skládal P (2003) Development of piezoelectric immunosensors for competitive and direct determination of atrazine. *Sensors and Actuators B: Chemical* 91(1-3): 333-341.
29. Cirillo G, Nicoletta FP, Curcio M, Spizzirri UG, Picci N, et al. (2014) Enzyme immobilization on smart polymers: Catalysis on demand. *Reactive and Functional Polymers* 83: 62-69.
30. Amine A, Mohammadi H, Bourais I, Palleschi G (2017) Biosensors and Bioelectronics. *Int J Electrochem Sci* 12.
31. Terry LA, White SF, Tigwell LJ (2005) The application of biosensors to fresh produce and the wider food industry. *J Agric Food Chem* 53: 1309-316.
32. Unnikrishnan B, Palanisamy S, Chen SM (2013) A simple electrochemical approach to fabricate a glucose biosensor based on graphene-glucose oxidase biocomposite. *Biosens Bioelectron* 39(1): 70-75.
33. Cosnier S (2007) Recent advances in biological sensors based on electro generated polymers: a review. *Analytical Letters* 40(7): 1260-1279.
34. Cosnier S, Holzinger M (2008) Design of carbon nano polymer frameworks by electro polymerization of SWCNT-pyrrole derivatives. *Electrochimica Acta* 53(11): 3948-3954.
35. Mazzei F, Antiochia R, Botre F, Favero G, Tortolini C (2014) Affinity-based biosensors in sport medicine and doping control analysis. *Bioanalysis* 6(2): 225-245.
36. Ashger M, Shahid M, Kamal S, Iqbal HMN (2014) Recent trends and valorization of immobilization strategies and ligninolytic enzymes by industrial biotechnology. *Journal of Molecular Catalysis B: Enzymatic* 101: 56-66.
37. Johnson BN, Mutharasan R (2014) Biosensor-based microRNA detection: techniques, design, performance, and challenges. *Analyst* 139(7):1576-1588.
38. Hamidi-Asl E, Palchetti I, Hasheminejad E, Mascini M (2013) A review on the electrochemical biosensors for determination of microRNAs. *Talanta* 115: 74-83.
39. Ahuja T, Mir IA, Kumar D, Rajesh (2007) Biomolecular immobilization on conducting polymers for biosensing applications. *Biomaterials* 28(5):791-805.
40. Pohanka M (2015) Determination of acetylcholinesterase and butyrylcholinesterase activity without dilution of biological samples. *Chemical Papers* 69: 4-16.
41. Kaur K, Kaushal P (2019) Enzymes as analytical tools for the assessment of food quality and food safety. *Advances in Enzyme Technology* pp: 273-292.
42. Wang B, Takahashi S, Du X, Anzai J (2014) Electrochemical biosensors based on ferroceneboronic acid and its derivatives: a review. *Biosensors Basel* 4(3): 243-256.
43. Grabowska I, Malecka K, Jarocka U, Radecki J, Radecka H (2014) Electrochemical biosensors for detection of avian influenza virus – current status and future trends. *Acta Biochim Pol* 61(3): 471-478.
44. Maurer M, Burri S, de Marchi S, Hullin R, Martinelli M, et al. (2010) Plasma homocysteine and cardiovascular risk in heart failure with and without cardiorenal syndrome. *Int J Cardiol* 141(1): 32-38.
45. Erden PE, Kili E (2013) A review of enzymatic uric acid biosensors based on amperometric detection. *Talanta* 107: 312-323.
46. Gutierrez JC, Amaro F, Martin-Gonzalez A (2015) Heavy metal whole-cell biosensors using eukaryotic microorganisms: an updated critical review. *Front Microbiol* 6: 48.
47. Lin H, Lu Q, Ge S, Cai Q, Grimes C (2010) Detection of pathogen *Escherichia coli* O157:H7 with a wireless magnetoelastic-sensing device amplified by using chitosan-modified magnetic Fe₃O₄ nanoparticles. *Sensors and Actuators B: Chemical* 147(1): 343-349.
48. Valach M, Katrlík J, Sturdík E, Gemeiner P (2009) Ethanol *Gluconobacter* biosensor designed for flow injection analysis: Application in ethanol fermentation off-line monitoring. *Sensors and Actuators B: Chemical* 138(2): 581-586.
49. Pohanka M, Skladal P (2008) Electrochemical Biosensors-Principles and Applications. *Journal of Applied Biomedicine* 6: 57-64.
50. Damborsky P, Svitel J, Katrlík J (2016) Optical biosensors. *Essays in Biochemistry* Portland Press Ltd *Essays Biochem* 60(1): 91-100.
51. Shen MY, Li BR, Li YK (2014) Silicon nanowire field-effect-transistor based biosensors: from sensitive to ultra-sensitive. *Biosens Bioelectron* 60: 101-111.
52. Chen C, Wang J (2020) Optical biosensors: An exhaustive and comprehensive review. *Analyst* 145: 1605-1628.
53. Kunzelmann S, Solscheid C, Webb MR (2014) Fluorescent bio- sensors: design and application to motor proteins. *EXS* 105: 25-47.

54. Liu J, Deun RV, Kaczmarek AM (2016) Optical thermometry of MoS₂: Eu³⁺ 2D luminescent nanosheets. *Journal of Materials Chemistry* 4: 9937-9941.
55. Wang J (2006) Zinc oxide nanocomb biosensor for glucose detection. *Applied Physics Letters* 88: 3106.
56. Rathnayake IVN, Megharaj M, Bolan N, Naidu R (2009) Tolerance of heavy metals by gram positive soil bacteria. *World Academy of Science Engineering and Technology* 53: 1185-1189.
57. Bisht A, Mishra A, Bisht H, Tripathi RM (2021) Nanomaterial Based Biosensors for Detection of Viruses Including SARS-CoV-2: A Review. *Journal of analysis and testing* 5(4): 327-340.
58. Doctoral dissertation University College Cork (2014) Electrochemical biosensor based on microfabricated electrode arrays for life sciences applications.
59. Imam HT, Marr PC, Marr AC (2021) Enzyme entrapment, biocatalyst immobilization without covalent attachment. *Green Chemistry* 23: 4980-5005.
60. Lei Y, Chen W, Mulchandani A (2006) Microbial biosensors. *Analytica chimica acta* 568(1-2): 200-210.
61. Putzbach W, Ronkainen NJ (2013) Immobilization techniques in the fabrication of nanomaterial-based electrochemical biosensors: a review. *Sensors (Basel)* 13(4): 4811-4840.
62. Monosik R, Stredansky M, Tkac J, Sturdik E (2012) Application of biosensors in analysis of foods and beverages. *Food Analytical Methods* 5: 40-53.
63. Sun JZ, Peter Kingori G, Si RW, Zhai DD, Liao ZH, et al. (2015) Microbial fuel cell-based biosensors for environmental monitoring: a review. *Water Sci Technol* 71(6): 801-809.
64. Gonzalez GA, Dixon N (2019) Genetically encoded biosensors for lignocellulose valorization. *Biotechnology for biofuels* 12: 1-14.
65. Lillehoj PB, Kaplan CW, He J, Shi W, Ho CM (2014) Rapid, electrical impedance detection of bacterial pathogens using immobilized antimicrobial peptides. *Journal of laboratory automation* 19(1): 42-49.
66. Jain RK (2013) Normalizing tumor microenvironment to treat cancer: bench to bedside to biomarkers. *J Clin Oncol* 31(17): 2205-2218.
67. Sheeran TJ (2003) Bioterrorism. *Encyclopedia of environmental microbiology-ENV119*. Department of Defense, Washington DC, pp: 12.
68. Dai C, Choi S (2013) Technology and applications of microbial biosensor. *Open Journal of Applied Biosensor* 2(3): 11.
69. Lazcka O, Del Campo FJ, Munoz FX (2007) Pathogen detection: a perspective of traditional methods and biosensors. *Biosensors and Bioelectronics* 22(7): 1205-1217.
70. Li Y, Liu X, Lin Z (2012) Recent developments and applications of surface plasmon resonance biosensors for the detection of mycotoxins in foodstuffs. *Food Chemistry* 132(3): 1549-1554.
71. Singh S, Singhal R, Malhotra BD (2007) Immobilization of cholesterol esterase and cholesterol oxidase onto sol-gel films for application to cholesterol biosensor. *Analytica Chimica Acta* 582(2): 335-343.
72. Ke N, Wang X, Xu X, Abbasi YA (2011) The xCELLigence system for real-time and label-free monitoring of cell viability. *Methods in Molecular biology* 740: 33-43.
73. Padoa CJ, Crowther NJ (2006) Engineered antibodies: A new tool for use in diabetes research. *Diabetes Research and Clinical Practice* 74(2): 51-62.
74. Sassolas A, Leca Bouvier BD, Blum LJ (2008) DNA biosensors and microarrays. *Chemical Reviews* 108(1): 109-139.
75. Khimji I, Kelly EY, Helwa, Y, Hoang M, Liu, J (2013) Visual optical biosensors based on DNA-functionalized polyacrylamide hydrogels. *Methods* 64(3): 292-298.
76. Kuswandi B (2003) Simple optical fibre biosensor based on immobilised enzyme for monitoring of trace heavy metal ions. *Anal Bioanal Chem* 376(7): 1104-1110.
77. Wang J, Sun XW, Wei A, Lei Y, Cai XP, et al. (2006) Zinc oxide nanocomb biosensor for glucose detection. *Applied Physics Letters* 88: 3106.
78. Gruhl F, Rapp BE, Lange K (2013) Biosensors for diagnostic applications. *Adv Biochem Eng Biotechnol* 133: 115-148.
79. Gordeliy VI (2002) Molecular basis of transmembrane signalling by sensory rhodopsin II-transducer complex. *Nature* 419(6906): 484-487.
80. Luong JH, Male KB, Glennon JD (2008) Biosensor technology: technology push versus market pull. *Biotechnology advances* 26(5): 492-500.

81. Fitzpatrick J, Fanning L, Hearty S, Leonard P, Manning BM, et al. (2000) Applications and recent developments in the use of antibodies for analysis. *Analytical Letters* 33(13): 25-63.
82. Wei D, Oyarzabal OA, Huang TS, Balasubramanian S, Sista S, et al. (2007) Development of a surface plasmon resonance biosensor for the identification of *Campylobacter jejuni*. *Journal of Microbiological Methods* 69(1): 78-85.
83. Lamprecht C, Hinterdorfer P, Ebner A (2014) Applications of biosensing atomic force microscopy in monitoring drug and nanoparticle delivery. *Expert Opin Drug Deliv* 11(8): 1237-1253.
84. Chen YW, Liu M, Kaneko T, McIntyre PC (2010) Atomic layer deposited hafnium oxide gate dielectrics for charge-based biosensors. *Electrochem Solid State Lett* 13(3): 29-32.
85. Mostafa GA (2010) Electrochemical Biosensors for the detection of pesticides. *The Open Electrochemistry Journal* 2: 22-42.
86. Caruso R, Verde A, Cabiati M, Milazzo F, Boroni C, et al. (2012) Association of pre-operative interleukin-6 levels with Interagency Registry for Mechanically Assisted Circulatory Support profiles and intensive care unit stay in left ventricular assist device patients. *The Journal of Heart and Lung Transplantation* 31(6): 625-633.
87. Chen J, Zhang J, Guo Y, Li J, Fu F, et al. (2011) An ultrasensitive electrochemical biosensor for detection of DNA species related to oral cancer based on nuclease-assisted target recycling and amplification of DNAzyme. *Chemical Communications* 47(28): 8004-8006.
88. Bo Y, Wang W, Qi J, Huang S (2011) A DNA biosensor based on graphene paste electrode modified with Prussian blue and chitosan. *Analyst* 136(9): 1946-1951.
89. Pundir CS, Chauhan N (2012) Acetylcholinesterase inhibition-based biosensors for pesticide determination: a review. *Anal Biochem* 429(1): 19-31.
90. Villalonga R, Díez P, Yáñez Sedeño P, Pingarrón JS (2011) Wiring horseradish peroxidase on gold nanoparticles based nanostructured polymeric network for the construction of mediatorless hydrogen peroxide biosensor. *Electrochimica Acta* 56(12): 4672-4677.
91. Schöning M, Arzdorf M, Mulchandani P, Che W, Mulchandani A (2003) Towards a capacitive enzyme sensor for direct determination of organophosphorus pesticides: fundamental studies and aspects of development. *Sensors* 3(6): 119-127.
92. Wang S, Poon GM, Wilson WD (2015) Quantitative investigation of protein-nucleic acid interactions by biosensor surface plasmon resonance. *Methods Mol Biol* 1334: 313-332.
93. Wang X, Lu X, Chen J (2014) Development of biosensor technologies for analysis of environmental contaminants. *Trends in Environmental Analytical Chemistry* 2: 25-32.
94. Rad AS, Mirabi A, Binaian E, Tayebi H (2011) A review on glucose and hydrogen peroxide biosensor based on modified electrode included silver nanoparticles. *Int J Electrochem Sci* 6: 3671-3683.
95. Smyth A, Talasila P, Cameron A (1999) An ethanol biosensor can detect low-oxygen injury in modified atmosphere packages of fresh-cut produce. *Postharvest Biology and Technology* 15(2): 127-134.
96. Amine A, Mohammadi H, Bourais I, Palleschi G (2006) Enzyme inhibition based biosensor for food safety and environmental monitoring. *Biosensors and bioelectronics* 21(8): 1405-1423.
97. Singh P (2017) Surface plasmon resonance: A boon for viral diagnostics. *Reference Module in Life Sciences*.
98. Lee M, Zine N, Baraket A, Zabala M, Campabadal F, et al. (2012) A novel biosensor based on hafnium oxide: Application for early stage detection of human interleukin-10. *Sensors and Actuators B: Chemical* 175: 201-207.
99. Mohamad NR, Marzuki NH, Buang NA, Huyop F, Wahab RA (2015) An overview of technologies for immobilization of enzymes and surface analysis techniques for immobilized enzymes. *Biotechnology, biotechnological equipment* 29(2): 205-220.
100. Wang W, Zhang TJ, Zhang DW, Li HY, Ma YR, et al. (2011) Amperometric hydrogen peroxide biosensor based on the immobilization of heme proteins on gold nanoparticles-bacteria cellulose nanofibers nanocomposite. *Talanta* 84(1): 71-77.
101. Sassolas A, Blum LJ, Leca Bouvier BD (2012) Immobilization strategies to develop enzymatic biosensors. *Biotechnology Advances* 30: 489-511.
102. Wei N, Xin X, Du J, Li J (2011) A novel hydrogen peroxide biosensor based on the immobilization of hemoglobin on three-dimensionally ordered macroporous (3DOM) gold nanoparticle-doped titanium dioxide (GTD) film. *Biosensors and Bioelectronics* 26: 3602-3607.

103. Ooi KGJ, Galatowicz G, Towler HM, Lightman SL, Calder VL (2006) Multiplex cytokine detection versus ELISA for aqueous humor: IL-5, IL-10, and IFN profiles in uveitis. *Investig Ophthalmol Vis Sci* 47: 272-277.
104. Kim J, Imani S, de Araujo WR, Warchall J, Valdes Ramirez G, Paixao TR, et al. (2015) Wearable salivary uric acid mouthguard biosensor with integrated wireless electronics. *Biosens. Bioelectron* 74: 1061-1068.
105. Pérez Elortondo F, Ojeda M, Albisu M, Salmerón J, Etayo I, et al. (2007) Food quality certification: An approach for the development of accredited sensory evaluation methods. *Food Quality and Preference* 18: 425-439.
106. Thévenot DR, Toth K, Durst RA, Wilson GS (1999) Electrochemical biosensors: Recommended definitions and classification. *Pure Appl Chem* 71: 2333-2348.
107. Chouteau C, Dzyadevych S, Durrieu C, Chovelon J (2005) A bi-enzymatic whole cell conductometric biosensor for heavy metal ions and pesticides detection in water samples. *Biosensors and Bioelectronics* 21: 273-281.
108. Tsai H, Doong R, Chiang H, Chen K (2003) Sol-gel derived urease-based optical biosensor for the rapid determination of heavy metals. *Analytica Chimica Acta* 481: 75-84.
109. Caruso R, Trunfio S, Milazzo F, Campolo J, De Maria R, et al. (2010) Early expression of pro- and anti-inflammatory cytokines in left ventricular assist device recipients with multiple organ failure syndrome. *Asaio Journal* 56: 313-318.
110. Turner AP (2000) Biosensors--sense and sensitivity. *Science* 290: 1315-1317.
111. Sheng Q, Wang M, Zheng J (2011) A novel hydrogen peroxide biosensor based on enzymatically induced deposition of polyaniline on the functionalized graphene-carbon nanotube hybrid materials. *Sensors and Actuators B: Chemical* 160: 1070-1077.
112. Serna L, Zetty A, Ayala A (2009) Use of enzymatic biosensors as quality indices: a synopsis of present and future trends in the food industry. *Chilean Journal of Agricultural Research* 69: 270-280.
113. Cañas AN, Cañizares Macías M (2004) Desarrollo de un sistema sensor para la cuantificación de glucosa en jugos de frutas. *Revista de la Sociedad Química de Mexico* 48: 106-110.
114. Asav E, Akyilmaz E (2010) Preparation and optimization of a bienzymic biosensor based on self-assembled monolayer modified gold electrode for alcohol and glucose detection. *Biosensors and Bioelectronics* 25: 1014-1018.
115. Turner AP (2013) Biosensors: sense and sensibility. *Chem Soc Rev* 42(8): 3184-3196.
116. Su L, Jia W, Hou C, Lei Y (2011) Microbial biosensors: a review. *Biosensors and bioelectronics* 26(5): 1788-1799.
117. da Costa Silva LM, Melo AF, Salgado AM (2004) Biosensors for Environmental Applications. *Pure Appl Chem* 76: 723-752.
118. Lim JW, Ha D, Lee J, Lee SK, Kim T (2015) Review of micro/nanotechnologies for microbial biosensors. *Frontiers in bioengineering and biotechnology* 3: 61.
119. Pundir CS, Devi R (2014) Biosensing methods for xanthine determination: a review. *Enzyme and Microbial Technology* 57: 55-62.
120. Domínguez Renedo O, Alonso Lomillo MA, Ferreira Gonçalves L, Arcos Martínez MJ (2009) Development of urease based amperometric biosensors for the inhibitive determination of Hg (II). *Talanta* 79(5): 1306-1310.
121. Kumar J, Kumar S, D'Souza J (2006) Optical microbial biosensor for detection of methyl parathion pesticide using *Flavobacterium* sp. whole cells adsorbed on glass fiber filters as disposable biocomponent. *Biosensors and Bioelectronics* 21(11): 2100-2105.
122. Badihi Mossberg M, Buchner V, Rishpon J (2007) Electrochemical biosensors for pollutants in the environment. *Electroanalysis* 19(19-20): 2015-2028.
123. Kaur H, Bhosale A, Shrivastav S (2018) Biosensors: classification, fundamental characterization and new trends: a review. *Int J Health Sci Res* 8(6): 315-333.
124. Li J, Wu D, Yu Y, Li T, Li K, et al. (2021) Rapid and unamplified identification of COVID-19 with morpholino-modified graphene field-effect transistor nanosensor. *Biosensors and Bioelectronics* 183: 113206.
125. Fang Y (2006) Label-free cell-based assays with optical biosensors in drug discovery. *Assay and Drug Development Technologies* 4(5): 583-595.
126. Chen S, Huang J, Du D, Li J, Tu H, et al. (2011) Methyl parathion hydrolase based nanocomposite biosensors for highly sensitive and selective determination of methyl parathion. *Biosensors and Bioelectronics* 26(11): 4320-4325.

127. Fu G, Yue X, Dai Z (2011) Glucose biosensor based on covalent immobilization of enzyme in sol-gel composite film combined with Prussian blue/carbon nanotubes hybrid. *Biosensors and Bioelectronics* 26(9): 3973-3976.
128. Hasan A, Nurunnabi M, Morshed M, Paul A, Polini A, et al. (2014). Recent Advances in Application of Biosensors in Tissue Engineering. *BioMed Research International* 2014: 307519.
129. Holzinger M, Le Goff A, Cosnier S (2014) Nanomaterials for biosensing applications: a review. *Frontiers in chemistry* 2: 63.
130. Ko P J, Ishikawa R, Sohn H, Sandhu A (2013) Porous silicon platform for optical detection of functionalized magnetic particles biosensing. *J Nanosci Nanotechnol* 13(4): 2451-2460.
131. Mohamad NR, Marzuki NH, Buang NA, Huyop F, Wahab RA (2015) An overview of technologies for immobilization of enzymes and surface analysis techniques for immobilized enzymes. *Biotechnology, Biotechnological Equipment* 29(1): 205-220.
132. Panjan P, Virtanen V, Sesay AM (2017) Determination of stability characteristics for electrochemical biosensors via thermally accelerated ageing. *Talanta* 170: 331-336.
133. Sang S, Wang Y, Feng Q, Wei Y, Ji J, et al. (2015) Progress of new label-free techniques for biosensors: a review. *Crit Rev Biotechnol* 36(3): 465-481.
134. Thunemann M, Schmidt K, de Wit C, Han X, Jain RK, et al. (2014) Correlative intravital imaging of cGMP signals and vasodilation in mice. *Front Physiol* 5: 394.
135. Tothill IE (2001) Biosensors Developments and Potential Applications in the Agricultural Diagnosis Sector. *Computers and Electronics in Agriculture* 30(1-3): 205-218.
136. Trabelsi I, Ayadi D, Bejar W, Bejar S, Chouayekh H, et al. (2014) Effects of *Lactobacillus plantarum* immobilization in alginate coated with chitosan and gelatin on antibacterial activity. *International Journal of Biological Macromolecules* 64: 84-89.
137. Vasileva N, Iotov V, Ivanov Y, Godjevargova T, Kotia N (2012) Immobilization of β -galactosidase on modified polypropylene membranes. *International Journal of Biological Macromolecules* 51(5): 710-719.
138. Verma N, Bhardwaj A (2015) Biosensor technology for pesticides – a review. *Appl Biochem Biotechnol* 175(6): 3093-3119.
139. Wang X, Li F, Guo Y (2020) Recent Trends in Nanomaterial-Based Biosensors for Point-of-Care Testing. *Frontiers in Chemistry* 8.
140. Xie L, Xu Y, Cao X (2013) Hydrogen peroxide biosensor based on hemoglobin immobilized at graphene, flower-like zinc oxide, and gold nanoparticles nanocomposite modified glassy carbon electrode. *Colloids and Surfaces B: Biointerfaces* 107: 245-250.
141. Zhang H, Meng Z, Wang Q, Zheng J (2011) A novel glucose biosensor based on direct electrochemistry of glucose oxidase incorporated in biomediated gold nanoparticles-carbon nanotubes composite film. *Sensors and Actuators B: Chemical* 158(1): 23-27.
142. Zhang W, Ab Asiri AM, Liu D, Du D, Lin Y (2014) Nanomaterial-based biosensors for environmental and biological monitoring of organophosphorus pesticides and nerve agents. *TrAC Trends in Analytical Chemistry* 54: 1-10.
143. Zhang Y, Muench S, Schulze H, Perz R, Yang B, et al. (2005) Disposable biosensor test for organophosphate and carbamate insecticides in milk. *Journal of Agricultural and Food Chemistry*, 53(13): 5110-5115.

