



Synthesis of Silica Nanoparticles Doped with Zinc Cation to Remove Bacteria DNA Harboring Antibiotic Resistance Genes from Aqueous Solution

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Abstract

This study investigated removing bacteria DNA from an aqueous solution using silica oxide nanoparticles doped with zinc cation ($\text{SiO}_2@\text{Zn}_2^+$). The authenticity of the adsorbent was confirmed by scanning electron microscopy (SEM) coupled with energy-dispersive x-ray spectroscopy (EDX). Adsorption studies involving two operating conditions (time-concentration profile and adsorbent dose) showed increased removal efficiency during the adsorption of bacteria DNA by $\text{SiO}_2@\text{Zn}_2^+$. Therefore, $\text{SiO}_2@\text{Zn}_2^+$ may be a promising adsorbent that can tackle the consequences of ARGs infected water.

Keywords: Nanoparticles; SEM; EDX

Abbreviations: SEM: Scanning Electron Microscopy; EDX: Energy-Dispersive X-Ray Spectroscopy; ARGs: Antibiotic-Resistance Genes; CTAB: Cetyltrimethylammonium Bromide; NFW: Nuclease-Free Water; AST: Antibiotic Susceptibility Test.

Introduction

The gain in the fight against bacterial infection has been short-lived due to the development and subsequent proliferation of antibiotic-resistance genes (ARGs) [1]. ARGs render antibiotics ineffective in the treatment of bacterial infections. Water bodies are one of the most important routes for the spread of ARGs. Thus, abundant ARGs have been

detected in freshwater, surface water, drinking water, and wastewater [2]. While the presence of naturally occurring minerals enhances the spread of ARGs in fresh and surface water, the hotspot for the enrichment of ARGs has been wastewater, incredibly wastewater rich in antibiotics [3]. In the quest to combat the proliferation of ARGs in water bodies, adsorption technology has been proposed as a cost-effective technique for the removal of ARGs [4], and the application of metallic oxides has been applied for this purpose [5].

In adsorption technology, the specific surface area and the physical and chemical affinities of the adsorbent towards the adsorbate are essential criteria for an adsorbent [6]. Metal oxide nanoparticles possess a large specific surface area [7]

and have been used as an adsorbent to remove organic and inorganic pollutants [8,9]. ZnO is a typical adsorbent used to combat the proliferation of antibiotic-resistant bacteria-harboring ARGs due to their bacteriostatic tendencies [10]. In lieu, in this study, mesoporous silica nanoparticles doped with zinc cation ($\text{SiO}_2@\text{Zn}^{2+}$) were synthesized via the combustion protocol and used to remove bacteria DNA harboring ARGs, free-cell DNA of *Listeria monocytogene* present in aqueous solution.

Materials and Methods

Synthesis and Characterization of $\text{SiO}_2@\text{Zn}^{2+}$

Incorporation of Zn^{2+} cation onto SiO_2 was achieved according to the procedure or method described in the literature with slight modifications [11]. 2 g of cetyltrimethylammonium bromide (CTAB) and 4 mL of NaOH were dissolved into 50 mL deionized water. The solution was stirred on the mantle for one hour at 70 °C. 5 mL of tetraorthosilicate (TEOS) and nitric acid (HNO_3) were after a minute. The solution was further stirred for 30 minutes at 60 °C. Then, 0.02 M of zinc nitrate ($\text{N}_2\text{O}_6\text{Zn}\cdot 6\text{H}_2\text{O}$) was added in the ratio 1:1 of Zn: TEOS. All particles were

further stirred for two hours before being separated and centrifuged. Then, the particles were washed three times with distilled water and dried overnight in an oven at 50 °C. The cation-containing particles were calcined at 550 °C for 18 h to remove the remaining CTAB and nitrite. The surface morphology and elemental compositions were achieved using a scanning electron microscope (SEM) coupled with an energy-dispersive x-ray spectroscope (EDX) (JOEL JSM-6390LVSEM), respectively.

Extraction and Molecular Characterization of Bacterial DNA

The antibiotic-resistant *Listeria monocytogene* bacteria used in this study were obtained from our laboratory archives isolated from food and vegetables. Before the commencement of genomic DNA extraction, the bacteria isolate was subjected to an antibiotic susceptibility test [12]. The extraction was carried out according to the method described in the literature [13]. The test for antibiotic resistance genes was conducted using the primers and PCR conditions of the targeted resistance gene shown in Table 1.

Antibiotic Class	PCR primers	Primer sequences	Product size (bp)	PCR protocols	References
Tetracyclines	tetA	F:GCTACATCCTGCTTGCCCTTC R:CATAGATCGCCGTGAAGAGG	210	94 °C – 5 min; 35[94 °C – 1 min; 55 °C – 1 min; 72 °C 1½ min]; 72 °C – 5 min.	[14]

Table 1: Primer sequence, expected product size, and PCR protocol used for the amplification of resistance genes.

Batch Adsorption Study

Bacterial DNA (2 mL) was used to contaminate nuclease-free water (NFW). 20 mg of $\text{SiO}_2@\text{Zn}^{2+}$ as adsorbent was weighed and added to the prepared contaminated NFW for the batch adsorption study according to the method described in the literature [15]. The effect of contact time (0-80 mins), initial DNA concentrations (7.28 µg/mL), and adsorbent dose (10-30 mg) on DNA removal were investigated. The optimum parameters were determined and adopted during the experimentation. The percentage removal efficiency (%R) in all the solutions was calculated using Equation 1 described by Panahi AH, et al. [16].

$$\%R = \frac{(C_i - C_f)}{C_i} \times 100 \quad (1)$$

C_i and C_f are the initial and final concentrations of DNA measured in µg/mL, and % R is the removal efficiency.

Result and Discussion

Antibiotic Susceptibility Test (AST)/Molecular Characterization of Genomic DNA

The AST test showed that *Listeria monocytogenes* (bacteria isolates) were resistant to sulfamethoxazole, erythromycin, streptomycin, amoxicillin, and tetracycline. The determination of resistance genes was achieved by visualizing the PCR product amplifications containing genomic DNA using gel electrophoresis, indicating that bacteria isolates harbor tetA resistance genes at the molecular weight of 210 bp (Figure 1).

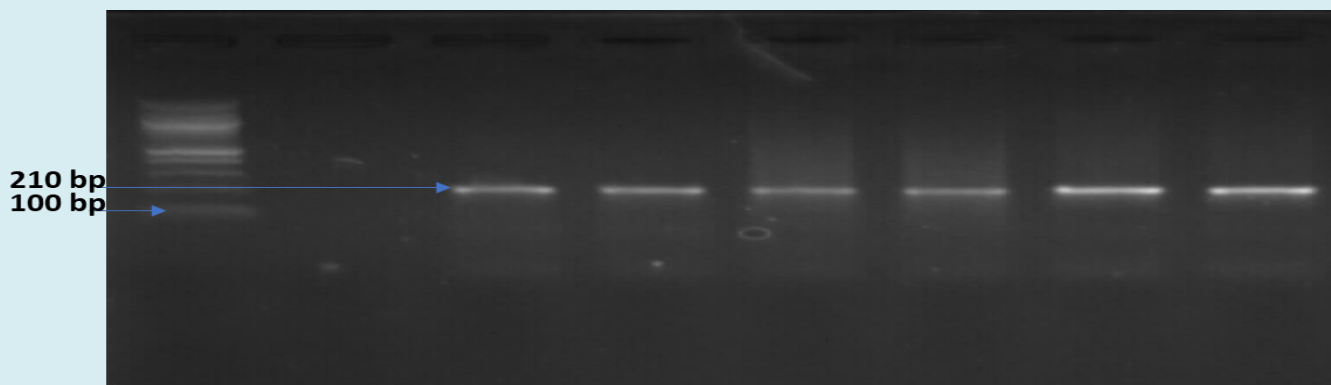


Figure 1: Gel representing tetracycline resistant gene (*tetA*) amplified.

Adsorbent Characterization

SEM and EDX Analysis: The morphology of the adsorbent ($\text{SiO}_2@\text{Zn}^{2+}$) was determined using the SEM instrument captured at 20 μm . The material was irregular and formed some slight aggregates (Figure 2A). The elemental

composition (Figure 2B) showed a strong signal for silica (Si) at 2Kev and a medium peak representing zinc (Zn) at 0.5 Kev, indicating that this material is pure and its synthesis was successful.

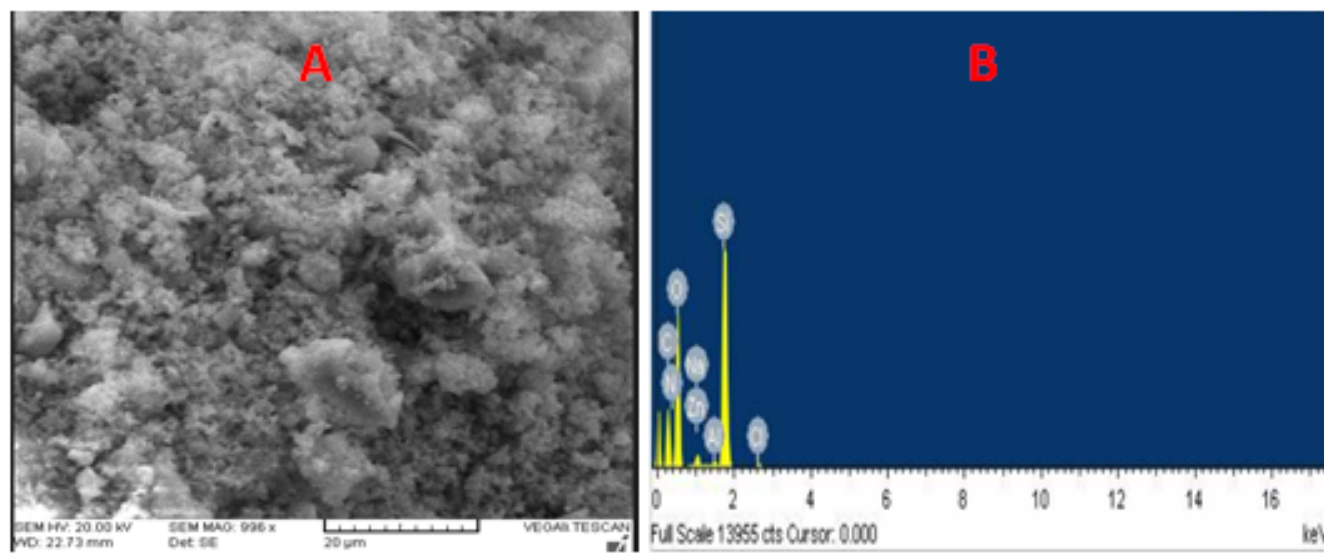


Figure 2: Represent SEM images captured at 20 μm and EDX showing the elemental compositions of $\text{SiO}_2@\text{Zn}^{2+}$.

Bacterial DNA Removal Study

Time-Concentration Profile: The effect of time as a function of initial DNA concentration was conducted at intervals while maintaining the adsorbent dose of 20 mg at pH 7.1. It was observed that when contact time was increased from 0 to 80 minutes, percentage adsorption efficiencies increased from 52.60-71.15% at an initial DNA concentration of 7.28 $\mu\text{g}/\text{mL}$ until removal became stable (Figure 3A). This result

indicates that increased time increases DNA removal from an aqueous solution. The result is similar to a recently published study Wang Y, et al. [17].

Effect of Adsorbent Dose: The adsorbent dose was studied, and it is illustrated in Figure 3B. As expected, the percentage removal of bacteria DNA rapidly increased with an increase in sorbent mass from 10 to 30 mg. At 30 mg, the optimum removal efficiencies reached 81.73%.

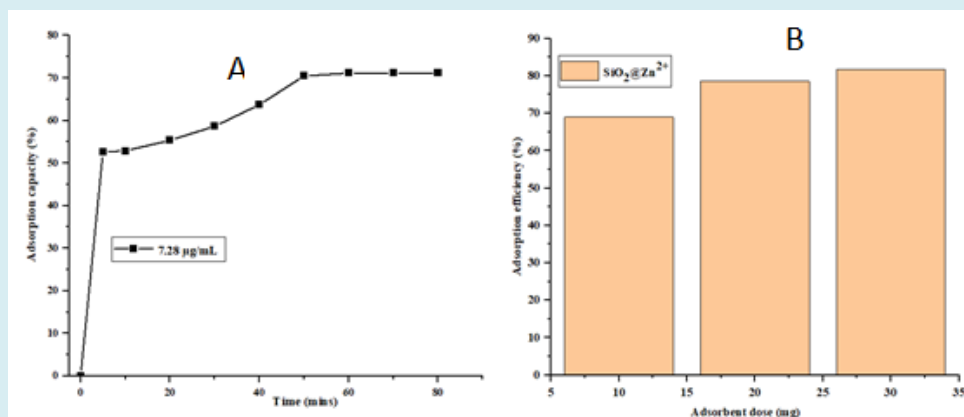


Figure 3: Effect of A) time-concentration profile and B) adsorbent dose on the removal of bacteria DNA onto SiO₂@Zn²⁺.

Conclusion

SiO₂@Zn²⁺ was successfully synthesized and employed for bacteria DNA removal from an aqueous solution. This study showed that different operating parameters (time-concentration profile and adsorbent dose) influenced the adsorption process and enhanced the removal of bacterial DNA conveying ARG onto SiO₂@Zn²⁺ from aqueous solution.

Acknowledgments

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