



# Ganoderma lucidum Modulates Neuronal Distorted Cytoskeletal Proteomics and Protein-Protein Interaction in Alzheimer's disease Model Animals

Rahman MA<sup>1,2\*</sup>, Hossain S<sup>1</sup>, Abdullah N<sup>2</sup> and Aminudin N<sup>2,3</sup>

<sup>1</sup>Department of Biochemistry and Molecular Biology, Jahangirnagar University, Bangladesh

<sup>2</sup>Mushroom Research Centre, Institute of Biological Sciences, Faculty of Science, University of Malaya, Malaysia

<sup>3</sup>University of Malaya Centre for Proteomics Research, Faculty of Medicine, University of Malaya, Malaysia

## Research Article

Volume 6 Issue 1

Received Date: November 20, 2020

Published Date: February 16, 2021

DOI: 10.23880/nnoaj-16000155

**\*Corresponding author:** Mohammad Azizur Rahman, Department of Biochemistry and Molecular Biology, Jahangirnagar University, Savar, Dhaka 1342, Bangladesh, Email: azizbmb@juniiv.edu

## Abstract

**Background:** Alzheimer's disease (AD) is the leading neurodegenerative disorder affecting memory, learning and behavior. Altered expression of proteins involved in neuronal structure and function is a recent observation of AD pathogenesis. Modulation of altered protein expression seems promising in AD therapeutics. In the present experiment, AD ameliorating effect of medicinal mushroom *Ganoderma lucidum* had been evaluated through its effect on neuronal cytoskeletal structure and function related protein expression pattern in AD model rats.

**Methods:** Wistar male rats ( $120 \pm 5\text{gm}$ ) were divided into three groups: control (C), AD (A) and *G. lucidum* hot water extract (HWE) fed AD (AE), each group containing 15 rats. AD model rats were prepared by infusing  $\text{A}\beta 1-42$  (ab120959, abcam, USA) into the cerebral ventricles. Protein extraction from the brain samples was performed following homogenization of the hippocampus (50 mg) with lysis buffer (1ml) using a homogenizer (Polytron PT 1200, Kinematica). Protein separation through SDS-PAGE and protein quantification through LC-chip MS/MS Q-TOF had been performed for label-free relative quantification. For statistical analyses, the data were exported to the Mass Profiler Professional (MPP) software and ANOVA ( $P<0.05$ ) had been performed to overcome the complications of false discovery associated with multiple test analyses. Functional interaction networks of the proteins were identified using the STRING (Search Tool for the Retrieval of Interacting Genes/Proteins) database (version 10.0; <http://string-db.org/>). For further identifying over-representing pathways and biological functions, the ingenuity pathway analysis (IPA), build version: 389077M, content version: 27821452, (Release date: 2016-06-14) was used (<https://www.ingenuity.com/wp-content/themes/ingenuity-quagen>). Datasets of the proteins significantly expressed ( $P<0.05$ ) and having log fold change of 1.5 and higher were uploaded (AD versus C, AD versus AE and C versus AE).

**Results:** Among 2,212 proteins identified in the present study, 819 had been found to be differentially expressed. Of the differentially expressed ones, 9 proteins had been linked with neuronal cytoskeletal structure and function regulation such as tubulin,  $\beta$ -actin, dihydropyrimidinase-related protein 2 (DRP-2), keratin, glial fibrillary acidic protein (GFAP), Rho A proteins, septin, cofilin, gelsolin and dynamin. AD rats manifested altered expression of proteins associated with neuronal structure and function. *G. lucidum* hot water treatment ameliorated the altered expression of those proteins.

**Conclusions:** Altered expression of hippocampal proteins is a hallmark of AD. Neuroproteomics regulatory approach towards AD amelioration seems promising. Inclusion of *G. lucidum* for proteomics based AD therapeutics in regulation of the proteins involved in neuronal structure and function seem apt. Thus, *G. lucidum* could be considered as an AD therapeutic agent.

**Keywords:** Alzheimer's Disease; Neuroproteomics; Ingenuity Pathway Analysis; Protein-protein interaction; STRING

**Abbreviations:** A: Alzheimer's disease model rat; AE: *Ganoderma lucidum* hot water extract fed Alzheimer's disease model rat; C: Control rat; AD: Alzheimer's disease; GL: *Ganoderma lucidum*; HWE: Hot water extract; FC: fold change of protein; KDa: Killo Dalton; M.wt: Molecular weight; PPI: Protein-protein interaction; IPA: Ingenuity pathway analysis; STRING: Search Tool for the Retrieval of Interacting Genes/Proteins.

## Introduction

According to United Nations Education Scientific and Cultural Organization (UNESCO), mushroom had included in the Ben Cao Gang Mu [(本草口目], the first pharmacopoeia in China, compiled during Ming dynasty (1590 AD)] as an aesthetic, spiritual and therapeutic agent [1,2]. Medicinal mushroom, Ling zhi (*Ganoderma lucidum*) has 2000 years-old reputation of usage in the Chinese medicine [3]. The State Pharmacopoeia of the People's Republic of China (2000) has incorporated *G. lucidum* as the potent replenisher of Qi (life force), salient mind soothering and cough relieving as well as anti-asthmatic agent [2]. Modern medicinal sector also acknowledge its inclusion as a therapeutic agent against numerous physiological disorders including, but not limited to, antioxidant, anti-inflammatory, antitumor, anticancer, antimicrobial, immunomodulatory and hepatoprotective agent [3-7]. Content of more than 400 gano-components had accredited *Ganoderma* as the "fungal biofactory", "panacea" and the "elixir of life" [3-7]. Gano-components conferring medicinal values range from polysaccharides to triterpenes, sterols, proteins, peptides, fatty acids and vitamins [3-7]. Recently, utilization of *G. lucidum* in ameliorating neurodegenerative diseases like Alzheimer's disease (AD), have received epoch-making attention [8]. We have also observed promising effect of *G. lucidum* as an AD therapeutic agent [9-15].

AD is a neurodegenerative disorder affecting mostly the elderly people is posing threat to the ever increasing aged humanity of the world. Currently, more than 46 million people over the world have been suffering from AD and this number had been projected to double by 2050 [16,17]. AD patients suffer from progressive loss of memory and learning abilities, behavioral abnormalities, disorientation about time and space, inability to smell and taste, difficulty in performing errands [17]. At severe stage, they become solely dependent on their family members and care-givers that lays extra economic burden on national and global health-care budget. Though the number of AD patients is skyrocketing, the world still awaits successful medication against this age-onset malady. AD occurs due to loss of neurons and synapses associated with memory, learning and behavioral performances [17]. Since its identification in 1901, different

hypotheses relating AD pathogenesis had been put forward. Among them, genetic predisposition, formation of amyloid beta (A $\beta$ ) plaques, neurofibrillary tangles (NFT) and mutation had received attention [14]. Recently, proteomics approaches towards understanding the mechanism and modulation of AD pathogenesis has received high attention [10,18-21]. Differential expression of proteins between AD and normal subjects would aid in formulating therapeutic strategies against AD. Though different approaches of AD therapeutics had been linked with *G. lucidum*, there is scarcity of reports describing AD neuroproteomics modulating effect of *G. lucidum*. Thus, the present study had been designed to evaluate the AD neuroproteomics modulatory effect of *G. lucidum*, especially the expression pattern of proteins involved in maintaining neuronal cytoskeletal structure and function.

## Materials and Methods

### Animals

Wistar male rats (120 ± 5 gm) were divided into three groups: control (C), AD (A) and *G. lucidum* hot water extract (HWE) fed AD (AE), each group containing 15 rats. AD model rats were prepared by infusing A $\beta_{1-42}$  (ab120959, abcam, USA) into the cerebral ventricles following an established method [22]. All the experimental protocols had been approved by the ethical permission committee, University of Malaya Institutional Animal Care and Use Committee (UMIACUC) [Ethics reference no. ISB/25/04/2013/NA (R)].

### Brain Sample Preparation and Protein Quantification

Following our previously established method [10], rat brain samples had been prepared and protein quantified in the hippocampi of C, AD and AE rats. Briefly, protein extraction from the hippocampi had been performed by homogenization (brain sample 50 mg; lysis buffer 1ml; homogenizer Polytron PT 1200, Kinematica); protein separation had been performed through SDS-PAGE [mini-PROTEAN tetra cell (165-8000, BIO-RAD, USA)] and protein quantification through LC-chip MS/MS Q-TOF [10]. To identify the proteins, the acquired MS/MS data were compared against the UniProtKB/Swiss Prot rat (*Rattus norvegicus*) database using the Spectrum Mill and X! Tandem. The differentially expressed proteins in the different groups were identified using their canonical sequence and proteins having fold change of at least 1.5 times were considered as the deregulated proteins. For validation of the identified proteins, the data were exported to the Scaffold database (version 4.5.1, Portland, USA). Proteins were grouped together if they would share at least two peptides and maintained their threshold level at 95.0% and <1% false discovery rate (FDR).

by the Peptide Prophet algorithm with Scaffold delta - mass correction for the matched peptide-spectra.

### Statistical Analysis

Data had been exported to the Mass Profiler Professional (MPP) software that analyzed depending on the MPP entities, the intensity of the total spectra of the proteins. Analysis of variance (ANOVA) had been performed to overcome the complications of false discovery associated with multiple test analyses.

### Bioinformatics and Analysis of Protein-Protein Interaction (PPI)

Most of the proteins do not work singly rather they participate in complex network or scaffold and interact with others. Thus, analysis of the relevant protein-protein networks provides important information in deciphering any bio-molecular system. Functional interaction networks of the proteins were identified using the STRING (Search Tool for the Retrieval of Interacting Genes/Proteins) database (version 10.0; <http://string-db.org/>). STRING displays protein-protein interactions in a large network of connectivity and protein hubs. Active prediction methods that we used were experiments, neighborhood, databases, gene fusions, coexpression, cooccurrence and text mining, using high confidence (0.7).

For further identifying over-representing pathways and biological functions, the ingenuity pathway analysis (IPA), build version: 389077M, content version: 27821452, (Release date: 2016-06-14) was used (<https://www.ingenuity.com/wp-content/themes/ingenuity-qiagen>). Datasets of the proteins significantly expressed ( $P<0.05$ ) and having log fold change of 1.5 and higher were uploaded (AD versus C, AD versus AE and C versus AE).

## Results and Discussion

### Differential Expression of Proteins Involved in Neuronal Cytoskeleton Maintenance

Differential expression of the proteins involved in maintaining neuronal cytoskeletal structure and function had been observed in the present study. Derangement of neuronal cytoskeleton through microtubule disassembly is an important feature of neurodegeneration [23]. STRING analysis revealed strong networks among the microtubule assembling cytoskeletal proteins such as tubulin,  $\beta$ -actin, dihydropyrimidinase-related protein 2 (DRP-2), keratin, glial fibrillary acidic protein (GFAP), Rho A proteins, septin, cofilin, gelsolin and dynamin.

### Tubulin

AD neurons suffer from disrupted microtubule structure and functioning [24]. Tubulin is the main component of microtubule and consists of dimers imparted by the alpha and beta chains. Differential expression of tubulin  $\alpha$  -1c, -4a,  $\beta$  -2a, -2B, -3 and -5 chains were observed in the present study. Molecular function based sub-network analysis showed different tubulin chains to be clustered together and deranged in AD. Both animal and human studies have linked decreased level of  $\alpha$  and  $\beta$  tubulin with human AD [25,26]. In AD brain,  $\beta$  tubulin becomes abnormally hyperphosphorylated and modified tubulin fails to assemble microtubules. Consequently, microtubule disassembly leads towards cytoskeletal vulnerability [27]. Recently, microtubular disassembly has been implicated in causing "mitochondrial traffic jam" in the AD neurons as mitochondrial shifting across the "rail-road of microtubule" becomes impeded in the AD brain [28].

### $\beta$ -Actin

Normally,  $\beta$ -actin is involved in maintenance of cytoskeleton, internal cell motility, neuronal network integrity and aids in memory and learning performances. Its altered expression and oxidized form had been linked with AD pathogenesis [29]. Impaired expression of actin is in agreement with the synaptic dysregulation associated with AD and age-related altered cytoskeletal structure, axonal dystrophy, reduced dendritic spines and impaired transport across membranes [30]. Enhanced accumulation of actin enhances tau-governed neurotoxicity [31,32].

### Dihydropyrimidinase-Related Protein 2 (DRP-2):

Dihydropyrimidinase related protein 2 (DRP2) is involved in regulation of axonal outgrowth and becomes hyperphosphorylated in NFT and its increased level is observed in AD model animals [33]. Compared to the normal neurons, AD neurons possess shortened dendrites which are a characteristic of their lowered communication with neighboring neurons [34]. DRP2 is expressed highly in the developing brains and altered in AD [35,36]. It regulates the activity of collapsin that elongates the dendrites, increases their communicability and repairs the damaged neurons [35,36]. Oxidative modification of DRP2 might cause reduced length of the dendrites and communication leading to lowered cognitive performance [37].

### Glial Fibrillary Acidic Protein (GFAP)

Glial fibrillary acidic protein (GFAP) provides structural support to the astrocytes and its elevated level in AD model

animals and in human subjects [33,38]. Its differential and reduced expression has been reported in AD animal studies [39,40].

### RhoA Proteins

Ras homolog gene family, member A (RhoA) proteins are involved in cytoskeleton regulatory processes such as dendrite development, axonal extension and protrusion [41]. They also stabilize the A $\beta$ -disrupted microtubules [41]. A  $\beta$  increases RhoA-GTPases and decreases neuronal spine production and neural connection both in the cell lines and also in the brains of the transgenic AD models [42,43]. Inhibition of the RhoA/ROCK signaling pathway seems ameliorating towards axonal growth and cognitive decline [41].

### Septin

Septins are microtubule associated filament-forming and GTP-binding proteins that participate in dendritic spine formation and in neurotransmitter release [44,45]. Like that of Musunury, et al. (2014) [46], increased expression of septin -2 and -3 were noted in the AD rats' hippocampi. Its increased expression in the AD rats might be involved in disrupting microtubular filament formation and associated cytoskeletal derangement in the AD rats [47-49].

### Cofilin

Brain cofilin activity reduces with age and in the AD subjects, it goes down aberrantly [50]. Transgenic mice studies also showed its lowered expression [50]. However, unlike other cytoskeletal proteins, its levels have not been found to be up-regulated in AD ameliorating cases [51]. As cofilin is a regulator of actin, increased cofilin expression points towards increased actin turnover and increased depolymerization of actin filament [51].

### Dynamin

Dynamin is a neuronal GTPase capable of free entry into and release from the synaptic vesicles [31]. Current finding of its decreased expression in the AD rats is consistent with those of others [52-55]. A $\beta$ -induced depleted dynamin1 level has been found to impair memory in the AD model rats [53]. Although some other findings found its variable level, altered dynamin induced affected neurotransmitter release has commonly been correlated with AD pathogenesis [32,55].

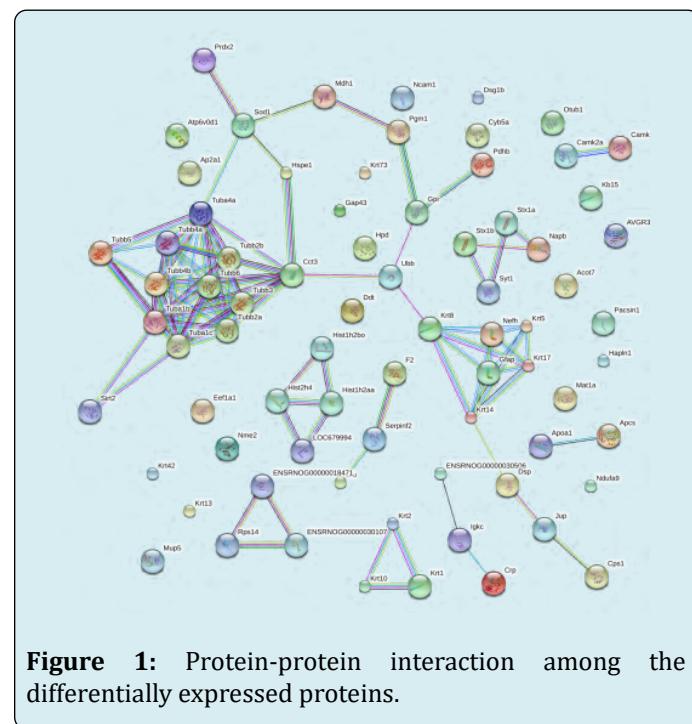
### Gelsolin

Gelsolin is a member of the actin-binding proteins having antioxidative, A $\beta$  binding and fibrillation inhibitory potentiality [56-58]. Its overproduction and/or

administration showed A $\beta$  lowering effect and thus, gelsolin has been regarded as an AD therapeutic agent [59]. More importantly, administration or overexpression of gelsolin results in significant reduction of amyloid load and decrease of A $\beta$  level in AD transgenic mice [56,59]. Down-regulated expression of gelsolin was noted in the AD rats' hippocampus that is in par with those of Manavalan, et al. [60]. However, some studies have reported confounding results regarding the expression of gelsolin in AD brains [61].

### PPI Network of the Upregulated Protein Clusters

The upregulated proteins constituted the interaction network entailing microtubule assembly and cytoskeletal structure and function (Figure 1). The proteins involved in this network are sirtuin2 and tubulin with the corresponding genes *Sirt2*, *Tuba1b*, *Tuba1c*, *Tubb2a*, *Tubb3*, *Tubb2b*, *Tuba4a*, *Tubb4a*, *Tubb4b*, *Tubb5* and *Tubb6* (Figure 1).



identified in the current experiment but not previously been studied, warrants extensive exploration for much conclusive remarks. In this endeavor, western blot analysis of the mostly regulated proteins for validation of the current findings is the immediate future aspect of the present study.

## Conclusion

Present study indicates the Alzheimer's disease modulatory effect of medicinal mushroom *G. lucidum* through differential expression of neuronal cytoskeleton regulatory proteins, restoration of disrupted protein-protein interaction network and maintenance of integrated pathways. Thus, utilization of *G. lucidum* in AD therapeutics seems promising. However, further studies should be carried out to determine therapeutic dosage, toxicity and safety.

## Declarations

**Ethics Approval and Consent to Participate:** All the experimental protocols had been approved by the ethical permission committee, University of Malaya Institutional Animal Care and Use Committee (UMIACUC) [Ethics reference no. ISB/25/04/2013/NA (R)].

**Consent for Publication:** All authors consented to publish.

**Availability of Data and Material:** Not applicable.

**Competing Interests:** Authors declare no competing interest.

**Authors' Contributions:** Mohammad Azizur Rahman designed and conducted the experiments, interpreted outcomes, composed manuscripts. Other authors supervised the experiments.

**Acknowledgement:** Authors gratefully acknowledge the personnel's and institutions that provided laboratory and equipmental support in conducting this research.

## References

- Ben Cao, Gang Mu (Compendium of Materia Medica) (2011) Documentary heritage submitted by China and recommended for inclusion in the Memory of the World Register in 2011. Memory of the World, UNESCO.
- The State Pharmacopoeia Commission of P. R. China (2000) State Pharmacopoeia Commission of the People's Republic of China. People's Medical Publishing House, Chemical Industry Press, Beijing, China.
- Wachtel-Galor S, Yuen J, Buswell JA, Iris F F Benzie (2011) *Ganoderma lucidum* (Lingzhi or Reishi): A Medicinal Mushroom. In: Benzie IFF, Wachtel-Galor S, (Eds.), *Herbal Medicine: Biomolecular and Clinical Aspects*, 2<sup>nd</sup> (Edn.), CRC Press/Taylor & Francis, Boca Raton (FL).
- Upton R (2000) American Herbal Pharmacopeia and Therapeutic Compendium: Reishi Mushroom, *Ganoderma lucidum*. Standards of Analysis, Quality Control, and Therapeutics; Santa Cruz, Canada, USA.
- Rahman MA, Abdullah N, Aminudin N (2018) Evaluation of the antioxidative and hypo-cholesterolemic effects of lingzhi or reishi medicinal mushroom, *Ganoderma lucidum* (Agaricomycetes), in ameliorating cardiovascular disease. *Int J Med Mushrooms*. 20(10): 961-969.
- Paterson RRM (2006) *Ganoderma*-A therapeutic fungal biofactory. *Phytochem* 67(18): 1985-2001.
- Rahman MA, Abdullah N, Aminudin N (2017) Corroborative assessment of mushroom as the graceful ageing and lifespan promoting agent. *Biointerface Research in Applied Chemistry* 7(3): 1-13.
- Qin LH, Wang C, Qin LW, Liang YF, Wang GH (2019) Spore powder of *Ganoderma lucidum* for Alzheimer's disease: A protocol for systematic review. *Medicine (Baltimore)*. 98(5): e14382.
- Rahman MA, Hossain S, Abdullah N, Aminudin N (2020) Lingzhi or Reishi Medicinal Mushroom, *Ganoderma lucidum* (Agaricomycetes) Ameliorates Non-spatial Learning and Memory Deficits in Rats with Hypercholesterolemia and Alzheimer's Disease. *Int J Med Mushrooms* 22(1): 93-103.
- Rahman MA, Hossain S, Abdullah N, Aminudin N (2019) Brain proteomics links oxidative stress with metabolic and cellular stress response proteins in behavioral alteration of Alzheimer's disease model rats. *AIMS Neuroscience* 6(4): 299-315.
- Rahman MA, Hossain S, Abdullah N, Aminudin N (2020) *Ganoderma lucidum* Ameliorates Spatial Memory and Memory-Related Protein Markers in Hypercholesterolemic and Alzheimer's Disease Model Rats. *Archives of Neurology and Neurologica Disorders* 3(2): 117-129.
- Rahman MA, Hossain MS, Abdullah N, Aminudin N (2020) Validation of *Ganoderma lucidum* against hypercholesterolemia and Alzheimer's disease. *Eur J Biol Res* 10(4): 314-325.
- Rahman MA, Hossain S, Abdullah N, Aminudin N (2020) lingzhi or reishi medicinal mushroom, *Ganoderma lucidum* (Agaricomycetes) ameliorates spatial learning and memory deficits in rats with hypercholesterolemia

- and alzheimer's disease. *Int J Med Mushrooms* 22(1): 93-103.
14. Rahman MA, Abdullah N, Aminudin N (2016) Interpretation of mushroom as a common therapeutic agent for Alzheimer's disease and cardiovascular diseases. *Crit Rev Biotechnol* 36(6): 1131-1142.
  15. Rahman MA, Hossain S, Abdullah N, Aminudin N (2020) Restoration of Perturbed  $\text{Ca}^{2+}$  Regulatory Proteins in Alzheimer's disease Model Rats by *Ganoderma lucidum*. *Alzheimer's disease and Dementia*.
  16. WHO (2020) World report on ageing and health. World Health Organization.
  17. Alzheimer's Association (2020) Alzheimer's disease Facts and Figures. *Alzheimers Dement* 16(3): 391.
  18. Rahman MA, Abdullah N, Aminudin N (2020) Hippocampal Proteomics Profiling along with Protein-Protein Interaction Analysis Elucidates Alzheimer's Disease Pathways and Genes. *Proteomics Bioinformatics* 2(2): 68-83.
  19. Bai B, Wang X, Li Y, Chen PC, Yu K, et al. (2020) Deep Multilayer Brain Proteomics Identifies Molecular Networks in Alzheimer's Disease Progression. *Neuron* 105(6): 975-991.e7.
  20. Johnson ECB, Dammer EB, Duong DM, Yin L, Thambisetty M, et al. (2018) Deep proteomic network analysis of Alzheimer's disease brain reveals alterations in RNA binding proteins and RNA splicing associated with disease. *Mol Neurodegener* 13(1): 52.
  21. Butterfield DA, Boyd-Kimball D, Castegna A (2003) Proteomics in Alzheimer's disease: insights into potential mechanisms of neurodegeneration. *J Neurochem* 86(6): 1313-1327.
  22. Fang F, Ning Y, Zhanhui F, Xiangqin L, Yong Y (2013) Alzheimer's disease animal model by aluminum, beta-amyloid and transforming growth factor beta-1, *Aging. Neurodegen* 1: 15-19.
  23. Sudhof TC (2004) The synaptic vesicle cycle. *Annu Rev Neurosci* 27: 509-547.
  24. McMurray C (2000) Neurodegeneration: diseases of the cytoskeleton? *Cell Death and Differentiation* 7(10): 861-865.
  25. Butler D, Bendiske J, Michaelis ML, Karanian DA, Bahr BA (2007) Microtubule-stabilizing agent prevents protein accumulation-induced loss of synaptic markers. *European Journal of Pharmacology* 562(1): 20-27.
  26. Pollak D, Cairns N, Lubec G (2003) Cytoskeleton derangement in brain of patients with Down syndrome, Alzheimer's disease and Pick's disease. *Journal of Neural Transmission Supplementum* 67: 149-158.
  27. Poon H, Castegna A, Farr S, Thongboonkerd V, Lynn B, et al. (2004) Quantitative proteomics analysis of specific protein expression and oxidative modification in aged senescence-accelerated-prone 8 mice brain. *Neuroscience* 126(4): 915-926.
  28. Vijayan S, El-Akkad E, Grundke-Iqbali I, Iqbal K (2001) A pool of  $\beta$ -tubulin is hyperphosphorylated at serine residues in Alzheimer disease brain. *FEBS Letters* 509(3): 375-381.
  29. Correia SC, Perry G, Moreira PI (2016) Mitochondrial traffic jams in Alzheimer's disease-pinpointing the roadblocks. *Biochimica Biophysica Acta (BBA)-Molecular Basis of Disease* 1862(10): 1909-1917.
  30. Castegna A, Thongboonkerd V, Klein JB, Lynn B, Markesberry WR, et al. (2003) Proteomic identification of nitrated proteins in Alzheimer's disease brain. *Journal of Neurochemistry* 85(6): 1394-1401.
  31. Penzes, P, VanLeeuwen JE (2011) Impaired regulation of synaptic actin cytoskeleton in Alzheimer's disease. *Brain Research Reviews* 67(1): 184-192.
  32. Fulga TA, Elson-Schwab I, Khurana V, Steinhilb ML, Spires TL, et al. (2007) Abnormal bundling and accumulation of F-actin mediates tau-induced neuronal degeneration in vivo. *Nature Cell Biology* 9(2): 139-148.
  33. Takano M, Yamashita T, Nagano K, Otani M, Maekura K, et al. (2013) Proteomic analysis of the hippocampus in Alzheimer's disease model mice by using two-dimensional fluorescence difference in gel electrophoresis. *Neuroscience Letters* 534: 85-89.
  34. Sizova D, Charbaut E, Delalande F, Poirier F, High AA, et al. (2007) Proteomic analysis of brain tissue from an Alzheimer's disease mouse model by two-dimensional difference gel electrophoresis. *Neurobiology of Aging* 28(3): 357-370.
  35. Coleman PD, Flood DG (1987) Neuron numbers and dendritic extent in normal aging and Alzheimer's disease. *Neurobiology of Aging* 8(6): 521-545.
  36. Castegna A, Aksakov M, Thongboonkerd V, Klein JB, Pierce WM, et al. (2002) Proteomic identification of oxidatively modified proteins in Alzheimer's disease brain. Part II: dihydropyrimidinase-related protein 2,  $\alpha$ -enolase and heat shock cognate 71. *Journal of Neurochemistry* 82(6):

- 1524-1532.
37. Lubec G, Nonaka M, Krapfenbauer K, Gratzer M, Cairns N, et al. (1999) Expression of the dihydropyrimidinase related protein 2 (DRP-2) in Down syndrome and Alzheimer's disease brain is downregulated at the mRNA and dysregulated at the protein level. *Journal of Neural Transmission and Supplementation* 57: 161-177.
  38. Butterfield DA, Perluigi M, Sultana R (2006b) Oxidative stress in Alzheimer's disease brain: new insights from redox proteomics. *European Journal of Pharmacology* 545(1): 39-50.
  39. Sultana R, Perluigi M, Newman SF, Pierce WM, Cini C, et al. (2010b) Redox proteomic analysis of carbonylated brain proteins in mild cognitive impairment and early Alzheimer's disease. *Antioxidants & Redox Signaling*, 12(3): 327-336.
  40. Korolainen MA, Auriola S, Nyman TA, Alafuzoff I, Pirttilä T (2005) Proteomic analysis of glial fibrillary acidic protein in Alzheimer's disease and aging brain. *Neurobiology of Disease* 20(3): 858-870.
  41. Shiozaki A, Tsuji T, Kohno R, Kawamata J, Uemura K, et al. (2004) Proteome analysis of brain proteins in Alzheimer's disease: subproteomics following sequentially extracted protein preparation. *Journal of Alzheimer's Disease* 6(3): 257-268.
  42. Fujita Y, Yamashita T (2014) Axon growth inhibition by RhoA/ROCK in the central nervous system. *Frontiers in Neuroscience* 8: 338.
  43. Minjarez B, Calderón-González KG, Rustarazo MLV, Herrera-Aguirre ME, Labra-Barrios ML, et al. (2016) Identification of proteins that are differentially expressed in brains with Alzheimer's disease using iTRAQ labeling and tandem mass spectrometry. *Journal of Proteomics* 139: 103-121.
  44. Petratos S, Li QX, George AJ, Hou X, Kerr ML, et al. (2008) The  $\beta$ -amyloid protein of Alzheimer's disease increases neuronal CRMP-2 phosphorylation by a Rho-GTP mechanism. *Brain* 131(1): 90-108.
  45. Ito H, Atsuzawa K, Morishita R, Usuda N, Sudo K, et al. (2009) Sept8 controls the binding of vesicle-associated membrane protein 2 to synaptophysin. *Journal of Neurochemistry* 108(4): 867-880.
  46. Spiliotis ET (2010) Regulation of microtubule organization and functions by septin GTPases. *Cytoskeleton* 67(6): 339-345.
  47. Musunuri S, Wetterhall M, Ingelsson M, Lannfelt L, Artemenko K, et al. (2014) Quantification of the brain proteome in Alzheimer's disease using multiplexed mass spectrometry. *Journal of Proteome Research* 13(4): 2056-2068.
  48. Hanai N, Nagata K, Kawajiri A, Shiromizu T, Saitoh N, et al. (2004) Biochemical and cell biological characterization of a mammalian septin, Sept11. *FEBS Letters* 568(1-3): 83-88.
  49. Marttinen M, Kurkinen KM, Soininen H, Haapasalo A, Hiltunen M (2015) Synaptic dysfunction and septin protein family members in neurodegenerative diseases. *Molecular Neurodegeneration* 10(1): 1.
  50. Tilleman K, Stevens I, Spittaels K, Haute CVD, Clerens S, et al. (2002) Differential expression of brain proteins in glycogen synthase kinase-3 transgenic mice: A proteomics point of view. *Proteomics* 2(1): 94-104.
  51. Barone E, Mosser S, Fraering PC (2014) Inactivation of brain Cofilin-1 by age, Alzheimer's disease and  $\gamma$ -secretase. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease* 1842(12): 2500-2509.
  52. Hwang JY, Shim JS, Song MY, Yim SV, Lee SE, et al. (2015) Proteomic analysis reveals that the protective effects of ginsenoside Rb1 are associated with the actin cytoskeleton in  $\beta$ -amyloid-treated neuronal cells. *Journal of Ginseng Research* 40(3): 278-284.
  53. Cao Y, Xiao Y, Ravid R, Guan ZZ (2010) Changed clathrin regulatory proteins in the brains of Alzheimer's disease patients and animal models. *Journal of Alzheimer's Disease* 22(1): 329-342.
  54. Watanabe T, Iwasaki K, Takasaki K, Yamagata N, Fujino M, et al. (2010) Dynamin 1 depletion and memory deficits in rats treated with A $\beta$  and cerebral ischemia. *Journal of Neuroscience Research* 88(9): 1908-1917.
  55. Zhou K, Yang Y, Gao L, He G, Li W, et al. (2010a) NMDA receptor hypofunction induces dysfunctions of energy metabolism and semaphorin signaling in rats: a synaptic proteome study. *Schizophrenia Bulletin* 38(3): 579-591.
  56. Kelly BL, Vassar R, Ferreira A (2005)  $\beta$ -Amyloid-induced Dynamin 1 Depletion in Hippocampal Neurons a potential mechanism for early cognitive decline in Alzheimer disease. *Journal of Biological Chemistry* 280(36): 31746-31753.
  57. Chauhan V, Ji L, Chauhan A (2008) Anti-amyloidogenic, anti-oxidant and anti-apoptotic role of gelsolin in Alzheimer's disease. *Biogerontology* 9(6): 381-389.
  58. Ji L, Chauhan A, Wegiel J, Essa MM, Chauhan V (2009)

- Gelsolin is proteolytically cleaved in the brains of individuals with Alzheimer's disease. *Journal of Alzheimer's Disease* 18(1): 105-111.
59. Ji L, Zhao X, Hua Z (2015) Potential therapeutic implications of gelsolin in Alzheimer's Disease. *Journal of Alzheimer's Disease* 44(1): 13-25.
60. Carro E (2010) Gelsolin as therapeutic target in Alzheimer's disease. *Expert Opinion on Therapeutic*

Targets 14(6): 585-592.

61. Manavalan A, Mishra M, Sze SK, Heese K (2013) Brain-site-specific proteome changes induced by neuronal P60TRP expression. *Neurosignals* 21(3-4): 129-149.
62. Güntert A, Campbell J, Saleem M, O'Brien DP, Thompson AJ, et al. (2010) Plasma gelsolin is decreased and correlates with rate of decline in Alzheimer's disease. *Journal of Alzheimer's Disease* 21(2): 585-596.

