

Gross Responses, Haematological Indices and Serum Biochemistry of Growing Cockerels Fed Diets Containing Graded Levels of Rumen Liquor Fermented Sugarcane Scrappings as Replacement for Maize

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Abstract

This study investigated the gross response, Haematological indices and serum biochemistry of growing cockerels fed diets containing 0, 5, 10 and 15 % of rumen liquor fermented sugarcane scrappings (RFSCS) as replacement of maize in diets of growing Ammo cockerels. At the end of 35 d feeding trial, there was no significant ($P>0.05$) differences in final weight (FW), weight gain (WG), feed conversion ratio (FCR), survival percentage (%), mean corpuscular volume (MCV) and protein while feed intake (FI), packed cell volume (PCV), haemoglobin (Hb) concentration, mean corpuscular haemoglobin concentration (MCHC), red blood cells (RBC) count, white blood cells (WBC) count, cholesterol, urea, uric acid, glucose, albumin and globulin were affected ($P<0.05$) by feed treatment. These results showed that RFSCS can replace up to 15% of maize in the diets of growing cockerels without any deleterious effects, 10% replacement of maize with RFSCS is recommended for optimum performance.

Keywords: Cockerels; Performance; Rumen Liquor; Sugarcane Scrappings; Serum

Introduction

The contribution of livestock sector in the achievement of optimum protein supply, economic returns and sustainable environment in most developing countries will require a continuous search for alternative feed

sources that are environmentally friendly, cost effective and in less demand as direct food resource for humans. This is even more important in the production of chickens in spite of their reported potentials of bridging the protein – supply gap in most developing countries, because feeds constitute 70% of production cost [1].

The largest dietary requirements for poultry are energy and protein [2]. These are predominantly supplied by maize grain and soybean meal (SBM) in poultry diets, respectively. The high cost of these feed ingredients resulting from diverse usage in human diets as well as industrial applications makes it necessary to search for alternative replacements that are cost effective. Agro-industrial waste products such as sugarcane scrapings are readily used as non-conventional feeds resource in replacing energy sources such as maize in monogastric diets [2-4]. Sugar cane scrapings are waste produced from scraping the rind of the sugar cane stem with a sharp knife in order to provide easier access to the underlying, soft parenchyma tissue, when the cane is being processed for chewing to extract the cane juice. This scrapings and peels are mostly heaped and sometimes burnt, or left thereby constituting an environmental pollution problem. High energy and crude fiber but low crude protein contents was reported for sugarcane scraping [5,6]. In order to enhance the utilization of agro-waste products in poultry and monogastric diets, the high fiber contents must be broken down and its nutrient composition improved through processing. Various processes have been reported in achieving nutritional improvement, reduction in fiber content of alternative feedstuffs, and these include fermentation, use of exogenous enzymes, mechanical and chemical treatments among others [5,7]. Both *in vivo* and *in vitro* fermentation of sugarcane waste and other agro – industrial waste using rumen liquor have been reported [2,8-10] for ruminants and monogastric animals. This fermentation process has been found to have immune stimulating effects on chickens [11].

Even though cockerels plays important role in community poultry upgrade programmes and are considered as good sources of protein meat, commanding high demand because of their meat flavour and toughness [12,13], their feed cost covering twenty (20) weeks poses great challenge to farmers. The use of fermentation using rumen filtrate will provide the opportunity for improving the nutritive values and utilization efficiency of sugar cane scrapings as a substitute to maize in poultry feeds. The basic assumption of this study was that cockerels' utilization of rumen liquor fermented sugar cane scrapings (RFSCS) will differ with the levels of inclusion. The objective of this study is therefore to evaluate the effect of different replacement levels of maize with RFSCS on the gross performance, haematology and serum biochemistry of cockerels.

Materials and Methods

Study Area

The experiment was carried out at the Poultry Unit of the Teaching and Research Farm, Faculty of Agriculture, Nasarawa State University Keffi, Shabu-Lafia campus, Nasarawa State located along Latitude 08°35'N and Longitude 08°33'E in the Guinea savanna zone of North Central Nigeria.

Agro Waste Products collection and processing

Sugarcane scrapings: Sugarcane scrapings were collected from the local sugarcane marketers who clean up sugar cane sticks for direct chewing by humans in Lafia municipality of Nasarawa State, Nigeria. The scrapings were obtained by scraping the rind of the sugarcane with a sharp knife. The collected scrapings were cleansed and sun dried on a polythene covered concrete slabs for 7d and stored.

Rumen Liquor: The bovine rumen fluid was collected as slaughter waste generated from slaughtering of White Fulani Cows at Lafia Municipal abattoir. The rumen liquor obtained from the slaughtered animals were mixed, homogenized and filtered through 100-mm nylon net. The solid materials were discarded while the fluid (liquor) part of the content was transferred into insulated flasks and stored. The stored rumen liquor was used as the source of inoculums.

Inoculation of Sugarcane scrapping with rumen liquor: The stored rumen liquor was used to inoculate the sun dried and tamped sugarcane scrapings by spraying 6L of rumen liquor per 30 Kg of sugarcane scrapings. The sprayed sugarcane scrapping was thoroughly mixed and transferred into airtight plastic container to initiate fermentation process that lasted for 72 h. Temperature, pH and texture of product were recorded at 24 h intervals. The fermented samples were sundried and analysed for nutrient composition according to [14] methods.

Management of experimental birds

A total of two hundred and eighty- eight (288) – twelve (12) weeks old day - old Ammo cockerels raised on commercial chick mash (1- 30 d) and growers' mash (30 – 84 d) were used for the experiment. The growing cockerels were individually weighed and randomly

assigned in a Completely Randomized design to Four (4) dietary treatments of six (6) replicates each. Each replicate had twelve (12) birds. The birds were reared on deep litter in an open sided well-ventilated poultry house. Routine management practices and routine administration of vaccines against Infectious Bursal

Disease and New Castle Disease were undertaken and the birds were fed *ad libitum* and had access to clean water.

Experimental diets: During the 35 d feeding trial, four (4) experimental diets were formulated to provide approximately 2800 Kcal/ME and 14% CP using Feedwin soft least cost soft ware (table 1).

	**Sugar Cane Scrapings	Fermented Sugar Cane Scrapings
Composition(g/kg)		
Dry Matter	906.7	976.5
Crude Protein	82.5	63.4
Ether Extract	33.6	33
Crude Fiber	364.8	89.5
Total Ash	99.8	31.5
Nitrogen Free Extract	674	759.1
Metabolizable Energy(Kcal/Kg)*	2970.5	3060.2
• Calculated (Pauzenga, 1985) ** Alu (2012) [4,35]		

Table1: Proximate composition of sugarcane scrapings.

The Four (4) dietary treatments are D1, D2, D3 and D4 representing 0, 5, 10 and 15 % replacement of maize with RFSCS.

Data Collection: Data collected during and the conclusion of the experiment include: feed intake, weight gain, feed conversion ratio, and survival percentages as measures of gross response. At the end of the feeding 35 d trial, blood samples were collected from four (4) birds per replicate using 5mls sterile disposable syringes and needles under the jugular veins in two (2) containers each with and without EDTA, blood samples collected were analyzed for haematological indices and serum biochemistry.

Analytical Procedures:

Proximate analysis: Chemical compositions of each of the experimental diets were determined following standard methods [14].

pH and Temperature: pH and temperature values for fermentation of sugarcane scrapings using rumen liquor were determined after every 24 h using Hm Digital Hand Held TDS – 3 tester.

Blood analysis: Blood samples were analysed for haematological parameters including haemoglobin (Hb), packed cell volume (PCV), white blood cell (WBC) count and Red blood cell (RBC) count. The cell counts were carried out by the use of haemocytometer while Hb, PCV and serum chemistry indices were determined using

methods adopted by [15] and the methods described by [16,17].

Statistical analysis: Data collected were subjected to one-way analysis of variance (ANOVA) using SPSS 20, Means were separated where there were significant differences using Duncan's Multiple Range Test [18].

Results and Discussion

The proximate composition of sugar cane scrapings with and without fermentation with rumen liquor is presented in (table 2).

	Temperature (°C)	pH
24h	25.03	6.95
48h	28.05	6.96
72h	27.02	7.65

Table 2: Effects of fermentation time on temperature and pH.

The values of DM, NFE increased with fermentation while CP, EE, CF and total ash were reduced with fermentation activity. The observed variations in the composition of between fermented and unfermented

sugarcane scrapings is accounted for by the effects of fermentation [7,19] through the used up and reconstitution of nutrient resource by fermenting microbes especially during the process of crude fiber degradation [20]. There were variations in fermentation temperature and slightly pH with increased length of fermentation time (24 h to 72 h) which was similarly

observed with *in vivo* fermentation of other agro waste products [10,21]. The composition of the experimental diets (Table 3).

Experimental Diets (%)				
	0	5	10	15
Ingredient composition(g/kg)				
Maize	460	410	360	310
RFSCS*	0	50	100	150
Maize bran	250	250	250	250
GNC	124	124	124	124
Rice offal	100	100	100	100
Bone meal	35	35	35	35
Blood meal	10	10	10	10
Soya bean oil	10	10	10	10
Fish meal	4	4	4	4
Salt	2.5	2.5	2.5	2.5
**Premix	2.5	2.5	2.5	2.5
DL-Methionine	1	1	1	1
L-Lysine	1	1	1	1
	1000	1000	1000	1000
Nutrient composition (g/kg)				
ME (Kcal/Kg)	2836.4	2822.4	2808.4	2794.4
CP	144.4	143.2	142	140.9
L-Lys	6.3	6.2	6.1	6
DL-Meth	3.3	3.2	3.1	3
M+C	5.5	5.3	5.2	5
EE	60.9	59.3	57.6	56
CF	59.1	62.5	65.9	69.3
Ca	8.5	8.7	9.3	10.2
P	5.8	6.1	6.2	6.9

Table 3: Composition of experimental diets.

*Inclusion levels of Rumen Liquor Fermented Sugar cane Scrapings

**Premix contains Vitamin A (8,000,000 I.U);Vitamin D3 (2,000,000 I.U); Vitamin E (5,000mg); Niacin (15,000mg); Vitamin B1 (1,500mg); Vitamin B2 (8,000mg); Vitamin B6 (1,500mg); Vitamin B12 (10mg); Vitamin K3 (2,000mg); Calpan (5,000mg); Biotin (20mg); Folic acid (500mg); Antioxidant (125,000mg); Choline chloride (200,000mg); Cobalt (200mg); Copper (5,000mg); Iodine (1,200mg); Iron (40,000mg); Manganese (80,000mg); Selenium (200mg); Zinc (60,000mg).

showed that the formulated experimental feeds are within the recommended nutritional requirements values for growing cockerels.

The gross responses of the experimental birds are presented in (Table 4).

Experimental Diets (%)					
	0	5	10	15	Pooled SEM
Initial wt (g)	369.41	359.26	359.26	371.41	2.44
Final wt (g)	1514.67	1478.23	1489.68	1449.61	23.61
Feed intake (g)	1073.00a	1066.57a	1071.65a	1041.30b	3.40*
Weight gain(g)	1145.26	1118.97	1130.42	1078.2	23.98
FCR	0.95	0.97	0.96	0.97	0.02
Survival %	98	98	98.67	98.67	0.16

Table 4: Gross Response of cockerels fed experimental diets.

*Inclusion levels of Rumen Liqour Fermented Sugar cane Scrapings

abc Means with different alphabets in the same row were significantly ($P < 0.05$) different

SEM Standard error mean

The experimental birds did not differ ($P < 0.05$) significantly in Final weight (FW), Weight Gain (WG), Feed Conversion Rate (FCR) and Survival percentage while they differ ($P < 0.05$) in feed intake (FI). Complete maize diet however had higher FI, FW and WG compared to other treatments while had the lowest values for the same parameters. These results support the usability of both sugar cane scrapings and the process of its nutritional improvement earlier reported [5,20,22].

The reduction in feed intake for 15% group may be as result of metabolites produced as bye products of fermentation process, thus with increased replacement of maize with RFSCS, these metabolites will increase in the feed and invariably reduce intake by birds. The high survival rate recorded for all groups confirms the report of [15,23] on the hardiness of cockerels and that of [11] who reported immune stimulation ability of sugar cane extracts in diets.

Results of haematological parameters and serum biochemistry are presented in (Table 5).

Experimental Diets (%)		
	15	SEM
Haematology**		
PCV (%)	29.40d	0.35
Hb (g/dl)	8.77c	0.12
MCV(fl)	104.78	4.73
MCH(g/dl)	33.42c	0.23
RBC($\times 10^3$ /ul)	2.45c	0.03
WBC($\times 10^6$ /ul)	3.65b	0.04
Serum Biochemistry		
Cholesterol(mmol/l)	2.85a	0.09
Urea (mmol/l)	2.25c	0.08
Protein(g/dl)	3.65	0.45
Uric Acid(mmol/l)	197.50d	8.06
Glucose(mmol/l)	9.15c	0.26
Albumin(g/dl)	16.50c	0.7
Globulin (g/dl)	1.06c	0.86

Table 5: Haematological and serum biochemistry of experimental cockerels

*Inclusion levels of Rumen Liqour Fermented Sugar cane Scrapings

Abc Means with different alphabets in the same row were significantly ($P < 0.05$) different
SEM Standard error mean

**PCV: Packed Cell Volume; Hb: Haemoglobin; MCV: Mean Corpuscular Volume; MCHC; Mean Corpuscular Haemoglobin Concentration; MCH: Mean Corpuscular Haemoglobin; RBC: Red Blood Cell; WBC: White Blood Cell.

Significant ($P < 0.05$) differences were observed among treatments for packed cell volume (PCV), haemoglobin (Hb) concentration, mean corpuscular haemoglobin concentration (MCHC), red blood cells (RBC) count, and white blood cells (WBC) count, while mean corpuscular volume did not differ ($P < 0.05$) significantly between dietary treatments. Even though the observed pattern of variations does not follow any particular trend in the rate of substitution of maize with RFSCS in the diet, PCV and MVC values decreased with increase in replacement of maize. The Haematological values obtained in this study were all within the reported normal ranges for chickens [15,24,25]. This suggest that the replacement of maize as energy source with up to 15% RFSCS was not deleterious to blood formation activities and its indicative of the nutritional potentials of RFSCS in the diets of cockerels [26,27]. The reduction in the values of haematological indices like PCV, Hb, MCV obtained in this study are supported by the findings of [28] who observed similar trend in broilers when fed rumen liquor fermented feed product which may be associated with poor intake resulting from the presence of metabolites in these diets [29].

The serum biochemical indices of the experimental birds showed significant ($P < 0.05$) effects on the following indices; cholesterol, urea, uric acid, glucose, albumin and globulin while values protein were not affected ($P < 0.05$) by feed treatment. These variations do not follow any pattern, however values for uric acid increased with increased utilization of RFSCS in the diets.

Cholesterol values were highest and lowest ($P < 0.05$) in the 10 and 5% replacement of maize, these values are within range reported for cockerels [15] and that of [30]. The total protein values obtained ranged from (3.05 – 3.65 g/dl) with 0% and 15% replacement diets having the lowest and highest values respectively the total protein of the experimental diets compared with that (3.60 – 4.0g/dl) reported for chickens [15,31]. Uric acid values was particularly high with increased replacement levels

of RFSCS in the diet (116.50 -197.50 mmol/l) for 0 and 15% inclusion levels. This finding is at variance with the report of [32] who reported a significant decrease of uric acid concentration in fasted cockerels, bearing in mind the low feed intake recorded for 15% replacement group. The values are however lower than the one reported. Glucose similarly increased with increased replacement with rumen liquor fermented sugar cane scrapings; this was supported by (El-Abasy et al., 2002) [11] who observed increased role of sugars in blood and immune response of cockerels. The albumin levels obtained in this study (1.65 – 2.15g/dl) for (15 and 0% respectively) compared favorably to the normal range (1.70 – 2.0g/dl) reported [15,33] for chickens while the globulin levels was highest in 10% replacement group (1.73 g/dl) and the values obtained in this study were within similar range (1.35 – 1.75g/dl) observed by [15,34] but were however, lower than the normal range (2.0 – 2.90g/dl) reported by the same workers.

Conclusion

These results obtained from this study showed that RFSCS can serve as up to 15% energy replacement of maize in the diets of growing cockerels without any deleterious effects gross performance and blood parameters of cockerels. For optimum performance, 10% replacement of maize with rumen liquor fermented sugar cane scrapings is recommended.

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References

1. Longe OG (1986) Replacement value of biscuit waste for maize in broiler diets. *Nigerian Journal of Animal Production* 13(1,2): 70-78.
2. Kanyinji F, Moonga M (2014) Effects of replacing maize meal with rumen filtrate-fermented cassava meal on growth and egg production performance in Japanese quails (*Cortunix japonica*). *J Adv Vet Anim Res* 1(3): 100-106.
3. Aletor VA (1986) Some Agro-Industrial by-Products and Waste in Livestock Feeding. A Review of prospects and problems. *World Review of Animal production* 22: 36-41.
4. Alu SE (2012) Nutrient digestibility and serum biochemistry of laying quails (*Cortunix cortunix japonica*) fed sugarcane scrapping meal-based diets supplemented with exogenous enzyme. *IOSR Journal of Agriculture and Veterinary Science (IOSR-JAVS)* 1(5): 29-35.
5. Alu SE, Kaankuka FG, Carew SN, Tuleun CD (2012) Effects Of Maxigrain® Supplementation Of Sugarcane Scrapping Meal-Based Diets On The Growth Rate, Nutrient Digestibility And Cost Implication Of Japanese Quail (*Cortunix Cortunix Japonica*) Chicks. *International Journal of Engineering Research & Technology* 1(9): 1-14.
6. Ayoade JA, Carew SN, Ameh AE (2007) The feed value of sugarcane scrapping meal for weaner rabbits: growth, meat yield and cost of production. *Proc. 32nd Ann. Conf. Nig. Soc. For Anim. Prod. (NSAP) University of Calabar, Nigeria.*
7. Ari MM, Ayanwale BA, Adama TZ, Olatunji EA (2012) Effects of Different Fermentation Methods on the Proximate Composition, Amino Acid Profile and Some Antinutritional Factors (ANFs) In Soyabeans (*Glycine Max*). *Fermentation Technology and Bioengineering* 2: 6-13.
8. Adebisi OA, Sokunbi OA, Ewuola EO (2009) Performance Evaluation and Bone Characteristics of Growing Cockerel Fed Diets Containing Different Levels of Diatomaceous Earth. *Middle-East Journal of Scientific Research* 4(1): 36-39.
9. Ding G, Chang Y, Zhou Z, Ren L, Meng Q (2014) Effect of *Saccharomyces cerevisiae* on Rumen Fermentation Characteristics, Nutrient Degradation and Cellulase Activity of Steers Fed Diets with Different Concentrate to Forage Ratios. *World Journal of Agricultural Research* 2(6): 303-308.
10. Priego A, Wilson A, Sutherland TM (1977) The effect on parameters of rumen fermentation, rumen volume and fluid flow rate of zebu bulls given chopped sugar cane supplemented with rice polishings or cassava root meal. *Trop Anim Prod* 2(3): 292-299.
11. El-Abasy M, Motobu M, Shimura K, Na KJ, Kang C, et al. (2002) Immunostimulating and growth - promoting effects of sugar cane extract (SCE) in chickens. *Journal of Veterinary Medical Science* 64(11): 1061-1063.
12. Azharul IM, Ranvig H, Howliger MAR (2005) Comparison of growth rate and meat yield characteristics of cockerels between Fayoumi and Sonali under village conditions in Bangladesh. *Livestock Research for Rural Development* 17(2).
13. Yakubu A, Ari MM, Ogbe AO, Ogah DM, Adua MM, et al. (2014) Preliminary investigation on community-based intervention through cockerel exchange programme for sustainable improved rural chicken production in Nasarawa State, Nigeria. *Livestock Research for Rural Development* 26(10): 1-6.
14. A.O.A.C. (2000) Association of official analytical chemists. 15th ed. William Tryd Press. Richmond Virginia V.S.A.
15. Kwari ID, Raji AO, Igwebuikue JU, Kibon A (2010) Response of growing cockerels to diets containing differently processed sorrel (*Hibiscus Sabdariffa*) Seed Meal. *International Journal of Science and Nature* 1(2): 183-190.
16. Aletor VA (1986) Some Agro-Industrial by-Products and Waste in Livestock Feeding. A Review of prospects and problems. *World Review of Animal production* 22: 36-41.
17. Green S (1976) Measurement of mean corpuscular volume (MCV) and mean corpuscular haemoglobin concentration. *Nigeria Journal of Animal production* 15: 213-218.

18. Duncan DB (1955) Multiple range and multiple tests. *Biometrics* 11: 1-42.
19. Barde RE, Ari MM (2004) Fermented Cassava Peels as Supplement to West African Dwarf Goats Grazing Natural Forage. Proceedings of the 38th Annual Conference of the Agricultural Society of Nigeria pp 69-624.
20. Fatufe AA, Matanmi IO (2008) The effect of probiotics supplementation on the growth performance of two strains of cockerels. *Journal of Central European Agriculture* 9(3): 405-410.
21. BPM (2016) Rumen pH. *Feeding Ruminants Correctly*.
22. Alikwe PCN (2013) Processing, Chemical, and Amino Acid Determination of Rumen Epithelial Scrapings as Potential Protein Feedstuff for Poultry. *The Pacific Journal of Science and Technology* 14(1): 422-429.
23. Abeke FO, Ogundipe SO, Sekoni AA, Dafwang II, Adeyinka IA, et al. (2008) Response of Shika Brown Cockerels to Graded Levels of Lablab Purpureus Beans. *Asian Journal of Poultry Science* 2(1): 10-16.
24. M Fraser (1986) *Merck Veterinary Manual A Handbook of Diagnosis, Therapy and Disease Prevention and Control for Veterinarians*. (6th edn), Merck and Co, Inc Ranway NJ, USA. 903Pp.
25. Ojabo LD, Adenkola AY (2013) The growth performance and haematology of cockerel chicks fed with sweet orange (*Citrus sinensis*) fruit peel meal. *Annals of Biological Research* 4(10): 11-15.
26. Holmes AD, Pigott MG, Campbell PA (1933) The hemoglobin content of chicken blood. *J Biol Chem* (103): 657-664.
27. Omoikhoje SO, Bamgbose AM, Aruna MB, Oboh SO, Eghiaruwa B (2004) Replacement value of groundnut cake with cooked bambara groundnut on the haematological traits and serum chemistry of broilers. In: Proc 9th Ann Conf Anim Sci Asso Nig (ASAN) Sept 13th-16th Ebonyi State University, Abakaliki – Nigeria Pp 86-89.
28. Adesua AA, Onibi GE (2014) Growth Performance, Haematology and Meat Quality Of Broiler Chickens Fed Rumen Liquor-Fermented Wheat Bran-Based Diets. *Jordan Journal of Agricultural Sciences* 10(4): 725-736.
29. Apata DF (2011) Effect of *Terminalia catappa* fruit meal fermented by *Aspergillus niger* as replacement of me on growth performance, nutrient digestibility and serum biochemical profile of broiler chickens. *Biotechnology Research International*.
30. Alabi OM, Adejumo DO, Aderemi FA, Lawal TE, Oguntunji AO, et al. (2008) Physiological response of broiler chickens to oral supplementation with *Telfaria occidentalis* leaf extract at finisher phase *Anim Sci Asso Nig (ASAN) A.B. U. Zaria*. Pp. 114 – 117.
31. Butler EJ (1971) Plasma Proteins. In: *Physiology and Biochemistry of the Domestic Fowls*. Vol 2 (Bell DJ and Freeman BM Eds), Academy Press, London, Pp. 933-961.
32. Milinković-Tur S, Stojević Z, Pirljin J, Zdelar-TUK M, Poljičak-Milas N, et al. (2007) Effects of fasting and refeeding on the antioxidant system in cockerels and pullets. *Acta Veterinaria Hungarica* 55(2): 181-189.
33. Onifade AA (1995) Comparative utilization of three dietary fibre sources by broiler chickens. Department of Animal Science, University of Ibadan, Ibadan, Nigeria.
34. Zanu HK, Adom SO, Appiah-Adu P (2012) Response of cockerels to diets containing different levels of sheanut cake. *Agricultural Sciences Research Journals* 2(7): 420-423.
35. Pauzenga U (1985) *Feeding Parent Stock*. *Zootech International* 22-25.