

Autologus Barriers, Fillers and Growing Factors: Using the Secondary Pathway of Haemostasis in Dental Clinics-Report of Four Clinical Cases

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Abstract

Haemostasis is an important system that maintain the hydrolytic balance of the body. The secondary pathway of this system produce a fibrin network to improve coagulation and regeneration. Platelet rich in fibrin, use the secondary pathway of haemostasis to isolate large quantities of fibrin in vitro, for clinical uses. In dental clinic, for improve bone regeneration, the PRF protocol it has become the principal toll for these purposes, leading a predictable bone regeneration and rapid osseointegration.

Keywords: Residual root extraction; Wisdom molar extraction; Osteointegration; Platelet plug

Introduction

Haemostasis is a unique and important system of physiology to maintain the hydrolytic balance in the body, stop bleeding [1]. When an injury occurs, a vasoconstriction lead the beginning of the primary haemostasis reducing the amount of blood flow through the area and limits the amount of blood loss where the platelets are activated to form a platelet plug [1,2]. To address the tissue regeneration a secondary path of haemostasis must be activated, where a blood clotting must be done [3]. This blood clotting needs fibrin threads that act as a glue for the sticky platelets [4,5]. As the fibrin mesh begins to form, the blood is also transformed from a liquid to a gel like substance through involvement of clotting and growing factors and pro-coagulants [1,3,5]. The coagulation process is useful in closing up and maintaining the platelet plug on larger wounds to tissue regeneration [6], in this fact, formation of a stable clot that is capable of withstanding the pressure in a blood vessel is essential to prevent bleeding and to promote wound healing [6,7]. Therapies using fibrin has been proposed to increase the tissue regeneration, and has been used for many conditions, like oral tissue regeneration [8-11], and plastic surgeries mainly [12,13]. Platelet rich in fibrin belongs to a new generation of platelet concentrates characterized by a simple preparation and, the most important item, the conservation of all biological components of blood [14]. Choukroun first developed the PRF in France for use in oral surgery [14]. This technique avoids any chemical anticoagulant or coagulant in the preparation, simplifying the technique, letting the natural polymerization of blood, producing a 3-dimensional organization of a fibrin

Case Report

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network [15].

The PRF process leads to engage the accumulation of many biological healing factors like cytokines, glycanic chains and structural glycoproteins (fibronectin) and immunity promoters trapped in the colloidal suspension between the fibrin network during centrifugation; these biological factor are available in situ for a convenient period, to lead the cells star the cicatricial matrix remodeling having well known synergetic effects on healing processes [16,17]. Platelet derived growth factors (PDGFs) are the first growing factors to be present in a wound initiating the connective tissue healing, bone regeneration and repair [18]. This factor is crucial in the migration, proliferation and survival of mesenchymatous cells linages, where perform well mitogenesis, angiogenesis and macrophage activation [19,20]. Moreover, PRF, contain IL-4, a healing cytokine, who during inflammatory phenomena supports healing process by modulation of inflammation and immune control [21]. Other important biological factor is the vascular endothelial growth factor (VEGF) considered as a master regulatory molecule for angiogenesis process, and plays a direct role in the control of endothelial cell behaviors, such proliferation, migration, specialization and survival enhancing the wound healing [17,21].

In this context, what happen if a large amount of fibrin clot is placed in a surgical wound?

Clinical Cases

Clinical case 1: Residual root extraction

A 21 years old female was come to the Periodontics and Dental Implantology course, for evaluation by a residual root referent to right maxillary second premolar.

Presurgical Therapy: The surgical procedure was explained to the patient and the informed consent was obtained. Pharmacology preparation of the patient included ketorolac (90mg), dexamethasone (4mg) and antibiotic prophylaxis with amoxicillin (2g) one hour before the procedure.

PRF preparation: The protocol was performed as described by Dohan, et al. [15]. Briefly, 10 mL of blood was drawn in 15 mL test tube without an anticoagulant and centrifuged immediately using a tabletop centrifuge (GREETMED CENTRIFUGE mod. GT119-100T, China) at 3000 rpm for 10 minutes.

The resultant product consists of the following three

- a. Top most layers consist of cellular plasma.
- b. PRF clot in the middle.
- c. Red blood corpuscles at the bottom.

With the absence of an anticoagulant, blood begins to coagulate as soon as is exposed to glass surface of test tube. Therefore, for successful preparation of PRF, blood must been collected and immediately centrifugation leads the clotting cascade initiated. PRF was obtain in the form of gelatinous plasma. Surgical procedure: after proper isolation of the surgical field, the operative site was anaesthetized using 72 mg of lidocaine with epinephrine (1:80000). An on line incision was made along the alveolar ridge soft tissue to retail a mucoperiostial flap. The residual root was extracted and the alveolar socket was curettage to remove any granulose and infected tissue. Immediately to curettage, the PRF was inserted in the alveolar socket. An autologous barrier was produced from PRF and this was putted on alveolar bone closed the alveolar socket. Soft tissue was approached and sutured to enhance a first intention wound healing. A periapical xray was performed immediately ending the surgical procedure and at 30 days (Figure 1).

Postoperative healing: There was not post-operative complication and healing was satisfactory. Radiographic control suggested the formation of an osseous matrix in all alveolar socket at 30 days.



Figure 1: a) Periapical radiography of residual root. b) Surgical access to proceed the residual root extraction. c) PRF procedure where shown the three phases. d)

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layers:

PRF clot were isolated and it cut in two parts. e) One part it was placed into the alveolar socket. f) The other part was performed an autologous barrier. g) The autologous barrier was placed to close the alveolar socket with PRF. h) Periapical radiography thirty days after surgery showed an alveolar socket fill with bone matrix.

Clinical case 2: Wisdom molar extraction

A 27 years old female was come to the Periodontics and Dental Implantology course, for evaluation by pain referent to left mandibular third molar.

Presurgical therapy: The surgical procedure was explained to the patient and the informed consent obtained. Pharmacology preparation of the patient included ketorolac (90mg), dexamethasone (4mg) and antibiotic prophylaxis with amoxicillin (2g) one hour before the procedure.

Surgical procedure: After proper isolation of the surgical field, the operative site was anaesthetized using 72 mg of lidocaine with epinephrine (1:80000). An Avellanal incision was performing to expose the coronal part and alveolar bone following by an osteotomy to extract the third molar. The alveolar socket was curettage to removeany granular tissue. Following the extraction, a PRF clot was placed into the alveolar socket and soft tissue was approached and sutured to enhance a first intention wound healing.

Postoperative healing: There was not post-operative complication and healing was satisfactory. Radiographic control suggested the formation of an osseous matrix in all alveolar socket at 30 days (Figure 2).



b) Surgical access to proceed the wisdom tooth extraction. c) Wisdom tooth with fused roots. d) PRF was placed into the alveolar socket. e) Periapical radiography thirty days after surgery showed an alveolar socket fill with bone matrix.

Clinical case 3: Rapid osteointegration in dental implants

A 18 years old male was come to the Private Practice, for evaluation by periodontosis of right superior central incisor. Because the vestibular bone cortical presented a fissure and the presence of parafunctional habits two stages implant rehabilitation was elected as treatment.

Presurgical therapy: The surgical procedure was explained to the patient and the informed consent obtained. Pharmacology preparation of the patient included ketorolac (90mg), dexamethasone (4mg) and antibiotic prophylaxis with clindamicin (600 mg) one hour before the procedure (Figure 3).



Figure 3: a) Central incisor lesion with apical decay and receding gum. b) Surgical access to proceed the central incisor extraction. c) PRF procedure where performed and dental implant was treated with PRF supernatant. d) Bone drilling was performing and was placed the treated dental implant. e) PRF autologous barrier was done and was placed into the alveolar socket protecting the dental implant. f) Sixty-days after surgery was performed the rehabilitation with ceramic crown and periapical radiography was taken suggesting the osseointegration.

Surgical procedure: After proper isolation of the surgical field, the operative site was anaesthetized using 72 mg of lidocaine with epinephrine (1:80000). A clean extraction was performing using a periotome to conserve alveolar bone diameter. Sequence drill was done to reach a 3.5 mm diameter, to place a conic 4.0 x 11 mm internal hexagon

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dental implant (Conexao, Sao Paulo, Brazil). Before dental implant place a PRF was performing and the fibrin clot was extracted. Dental implant was placed into de fibrin clot to capture any rich biochemical and biological factors in the surface. After that, the implant was immediately placed into the bone.

Postoperative healing: There was not post-operative complication and healing was satisfactory, radiographic and antitorque control suggested the osseointegration at two months.

Prosthetic rehabilitation: Prosthetic rehabilitation was performed using a porcelain crown.

Clinical case 4: Filling large bone defect with implant placement "Obando's fille technique"

A 48 years old female was come to the Private Practice for large coronal destruction of first left mandibular molar.

Presurgical therapy: The surgical procedure was explained to the patient and the informed consent obtained. Pharmacology preparation of the patient included ketorolac (90mg), dexamethasone (4mg) and antibiotic prophylaxis with amoxicillin (2g) one hour before the procedure.

Surgical procedure: after proper isolation of the surgical field, the operative site was anaesthetized using 72 mg of lidocaine with epinephrine (1:80000). An incision was performing and the molar was extracted. The mesial and distal roots were fusionated resulting a large alveolar socket. The inferior nerve canal was a 2mm to the apical portion of alveolar socket. The alveolar socket was curettage to remove any granular tissue. A 60 mL amount of blood was extracted from the patient and the PRF was performed. Following the extraction, a PRF clot was placed around of Bicon implant and the implant was placed in to the alveolar socket with primary stability less than 20 N. Another PRF clot was placed above of dental implant to close the alveolar socket.

Postoperative healing: there was not post-operative complication and healing was satisfactory. Radiographic control suggested the formation of an osseous matrix in all alveolar socket and anosseointegratoin of implant was observed at fourth month.



Figure 4: a) Periapical radiography of first inferior molar showed large coronal destruction with dental pulp compromise. b) Surgical access to proceed the molar extraction, it was present a fused roots. c) PRF procedure where performed and fibrin clot was placed around of dental implant. d) Dental implant with fibrin clot were placed into the alveolar socket reaching a primary stability less than 20N, fibrin clot and autologous barrier was done and was placed into the alveolar socket protecting the dental implant. e) Periapical radiography was taken fourth months after surgery showed a bone healing suggesting the osseointegration. f) Immediately was performed the rehabilitation with ceramic crown.

Discussion

The PRF protocol is frequently used with very favourable results for growth of hard and soft in the specialist dental clinic tissues. In implantology, it is also frequently used because of their biological factors that accelerate the processes of osseointegration [22] and especially accelerate bone formation in extraction sockets for future implant placement [23]. The case Nº1 and Nº2 PRF show the benefits of rapid bone regeneration in bone sockets after teeth extraction. This observation is in accordance with Anwandter (2016) [24] who describe well ridge preservation after teeth extraction [24]. This event can be explained by the realising of growth factors mainly and citokines immersed in the fibrin clot that can control the inflammatory response and the regenerative properties of immune system to modulate of cellular migration and proliferation, accelerating the bone healing [25,26]. In the case of third molar, PRF causes a periodontal health distal to second molar avoiding posterior inflammation [27].

The case Nº 3 not rich plasma was placed into fibrin in the socket, but the biological components clot implant transferred, leaving this in direct contact for the absorption of such biological factors may consequently form bone quickly by the expression BMPs [28]. Thus and subjectively, it can explain the accelerated osseointegration obtained in two months by activation of BMPs by the presence of the extracted biological factors PRF [28,29], these results are in agreement with those observed by Oncu et al. [22]. In case No. 4, fill a large alveolus was perform with concomitant implant placement without sufficient primary stability for osseointegration. According to some reports literature [22], the PRF is used to increase the primary stability of the implant filling the bone bed first with PRF and then place the implant. In this case, the clot PRF was prepared aroundthe implant and placed in the socket with a primary stability of 30N (only was given a direction to the implant with respect to the future prosthesis) and then completely fills the rest of the alveolus with PRF. In our teaching practice as a private, we perform the Obando technique to increase primary stability and filler large bone defects.

All cases showed the rapid bone healing and bone regeneration of PRF protocol in dental surgery and dental implant. Preserving bone quantity and quality for future implant rehabilitation are the most concern topic in implant surgery planning [10,30-32]. Specialist employed commercial bone to fill bone socket after dental extraction with the consideration of healing time of this socket between 4 or 6 months [33-35]. However, employing fibrin clot, the bone quality and quantity are same or more than commercial substitutes in less of time [24,36]. The rapid osseointegrationis directly related with the dental implant surface treatment and the biological growing factor as BMPs [37,38]. Some studies showed a rapid osseointegration of dental implant when BMPs are using on implant surface before placed into the bone [37-39]. However, commercial BMPs are very expensive and the cares of these proteins are many to avoid its denaturation. PRF offers similarities results because presents many growing factors including pre-BMPs which promotes acceleration of bone healing in dental implant treatments shortening the osseointegration time [28,40]. In this fact, large quantity of PRF cause positives effects in wound healing leading predicable bone regeneration and rapid osseointegration in dental implant clinics.

References

- 1. Chan AK, Paredes N (2013) The coagulation system in humans. Methods Mol Biol 992: 3-12.
- Versteeg HH, Heemskerk JW, Levi M, Reitsma PH (2013) New fundamentals in hemostasis. Physiol Rev 93(1): 327-358.
- 3. Margetic S (2012) Inflammation and haemostasis. Biochemia medica 22(1): 49-62.
- 4. Favaloro EJ (2016) Towards personalised therapy for von Willebrand disease: a future role for recombinant products. Blood Transfus 14(2): 262-276.
- 5. Litvinov RI, Weisel JW (2016) What Is the Biological and Clinical Relevance of Fibrin? Semin Thromb Hemost 42(4): 333-343.
- 6. Draxler DF, Medcalf RL (2015) The fibrinolytic system-more than fibrinolysis? Transfusion medicine reviews 29(2): 102-109.
- Zuliani-Alvarez L, Midwood KS (2015) Fibrinogen-Related Proteins in Tissue Repair: How a Unique Domain with a Common Structure Controls Diverse Aspects of Wound Healing. Adv Wound Care 4(5): 273-285.
- 8. Arunachalam M, Pulikkotil SJ, Sonia N (2016) Platelet Rich Fibrin in Periodontal Regeneration. Open Dent J 10: 174-181.
- 9. Choukroun J, Diss A, Simonpieri A, Girard MO, Schoeffler C, et al. (2006) Platelet-rich fibrin (PRF): a second-generation platelet concentrate. Part IV: clinical effects on tissue healing. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 101(3): e56-60.
- Oncu E, Bayram B, Kantarci A, Gulsever S, Alaaddinoglu EE (2016) Positive effect of platelet rich fibrin on osseointegration. Med Oral Patol Oral Cir Bucal 21(5): e601-607.
- 11. Saluja H, Dehane V, Mahindra U (2011) Platelet-Rich fibrin: A second generation platelet concentrate and a new friend of oral and maxillofacial surgeons. Ann Maxillofac Surg 1(1): 53-57.

- 12. Toriumi DM, Chung VK, Cappelle QM (2016) Surgical Adhesives in Facial Plastic Surgery. Otolaryngol Clin North America 49(3): 585-599.
- 13. Hershcovitch MD, Hom DB (2012) Update in Wound Healing in Facial Plastic Surgery. Arch Fac Plas Surg 14(6): 387-393.
- 14. Dohan DM, Choukroun J, Diss A, Dohan SL, Dohan AJ, et al. (2006) Platelet-rich fibrin (PRF): a secondgeneration platelet concentrate. Part I: technological concepts and evolution. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 101(3): e37-44.
- 15. Dohan DM, Choukroun J, Diss A, Dohan SL, Dohan AJ, et al. (2006) Platelet-rich fibrin (PRF): a secondgeneration platelet concentrate. Part II: plateletrelated biologic features. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 101(3): e45-50.
- Ghanaati S, Booms P, Orlowska A, Kubesch A, Lorenz J, et al. (2014) Advanced platelet-rich fibrin: a new concept for cell-based tissue engineering by means of inflammatory cells. J Oral Implantol 40(6): 679-689.
- 17. Dohan DM, Choukroun J, Diss A, Dohan SL, Dohan AJ, et al. (2006) Platelet-rich fibrin (PRF): a secondgeneration platelet concentrate. Part III: leucocyte activation: a new feature for platelet concentrates? Oral Surg Oral Med Oral Pathol Oral Radiol Endod 101(3): e51-55.
- 18. Patel S, Maheshwari A, Chandra A (2016) Biomarkers for wound healing and their evaluation. J Wound Care 25(1): 46-55.
- 19. Shah P, Keppler L, Rutkowski J (2014) A Review of Platelet Derived Growth Factor Playing Pivotal Role in Bone Regeneration. J Oral Implantol 40(3): 330-340.
- 20. Gleissner CA (2012) Platelet-derived chemokines in atherogenesis: what's new? Curr Vasc Pharmacol 10(5): 563-569.
- Passaretti F, Tia M, D'Esposito V, De Pascale M, Del Corso M, et al. (2014) Growth-promoting action and growth factor release by different platelet derivatives. Platelets 25(4): 252-256.
- 22. Oncu E, Alaaddinoglu EE (2015) The effect of plateletrich fibrin on implant stability. Int J Oral Maxillofac Implants 30(3): 578-582.

- Kotsakis G, Prasad H, Rohrer M, Hinrichs J, Boufidou F, et al. (2016) Extraction socket management utilizing Platelet-Rich-Fibrin: A proof-of-principle study of the "Accelerated-early implant placement" concept. J Oral Implantol 42(2): 164-168.
- 24. Anwandter A, Bohmann S, Nally M, Castro AB, Quirynen M, et al. (2016) Dimensional changes of the post extraction alveolar ridge, preserved with Leukocyte- and Platelet Rich Fibrin: A clinical pilot study. J Dent 52: 23-29.
- 25. Dohan EDM, Del CM, Diss A, Mouhyi J, Charrier JB (2010) Three-dimensional architecture and cell composition of a Choukroun's platelet-rich fibrin clot and membrane. J Periodontol 81(4): 546-555.
- 26. Dohan EDM, Doglioli P, de Peppo GM, Del CM, Charrier JB (2010) Choukroun's platelet-rich fibrin (PRF) stimulates in vitro proliferation and differentiation of human oral bone mesenchymal stem cell in a dose-dependent way. Arch Oral Biol 55(3): 185-194.
- 27. Doiphode AM, Hegde P, Mahindra U, Santhosh KSM, Tenglikar PD, et al. (2016) Evaluation of the efficacy of platelet-rich plasma and platelet-rich fibrin in alveolar defects after removal of impacted bilateral mandibular third molars. J Int Soc Prev Community Dent 6(Suppl 1): S47-52.
- 28. Woo SM, Kim WJ, Lim HS, Choi NK, Kim SH, et al. (2016)Combination of Mineral Trioxide Aggregate and Platelet-rich Fibrin Promotes the Odontoblastic Differentiation and Mineralization of Human Dental Pulp Cells via BMP/Smad Signaling Pathway. J Endod 42(1): 82-88.
- 29. Kim HJ, Nam HW, Hur CY, Park M, Yang HS, et al. (2011) The effect of platelet rich plasma from bone marrow aspirate with added bone morphogenetic protein-2 on the Achilles tendon-bone junction in rabbits. Clin Orthop Surg 3(4): 325-331.
- 30. Knapen M, Gheldof D, Drion P, Layrolle P, Rompen E, et al. (2015) Effect of leukocyte- and platelet-rich fibrin (L-PRF) on bone regeneration: a study in rabbits. Clinical implant dentistry and related research 17 (S1): e143-152.
- 31. Davis VL, Abukabda AB, Radio NM, Witt-Enderby PA, Clafshenkel WP, et al. (2014) Platelet-rich preparations to improve healing. Part II: platelet activation and enrichment, leukocyte inclusion, and

Obando-Pereda GA. Autologus Barriers, Fillers and Growing Factors: Using the Secondary Pathway of Haemostasis in Dental Clinics-Report of Four Clinical Cases. J Dental Sci 2016, 1(2): 000107.

other selection criteria. Journal of oral Implantology 40(4): 511-521.

- 32. Shawky H, Seifeldin SA (2015) Does Platelet-Rich Fibrin Enhance Bone Quality and Quantity of Alveolar Cleft Reconstruction? The Cleft palate-craniofacial journal: official publication of the American Cleft Palate-Craniofacial Association.
- 33. Allegrini S Jr, Koening B Jr, Allegrini MR, Yoshimoto M, Gedrange T, et al. (2008)Alveolar ridge sockets preservation with bone grafting--review. Ann Acad Med Stetin 54(1): 70-81.
- 34. Cardaropoli D, Cardaropoli G (2008) Preservation of the postextraction alveolar ridge: a clinical and histologic study. Int J Periodontics Restorative Dent 28(5): 469-477.
- 35. Hong JY, Lee JS, Pang EK, Jung UW, Choi SH, et al. (2014) Impact of different synthetic bone fillers on healing of extraction sockets: an experimental study in dogs. Clinical oral implants research 25(2): e30-37.
- 36. Temmerman A, Vandessel J, Castro A, Jacobs R, Teughels W, et al. (2016) The use of Leucocyte and Platelet Rich Fibrin (L-PRF) in socket management and ridge preservation: A split-mouth, randomised, controlled clinical trial. J Clin Periodontol.

- 37. Teng FY, Chen WC, Wang YL, Hung CC, Tseng CC (2016) Effects of Osseointegration by Bone Morphogenetic Protein-2 on Titanium Implants In Vitro and In Vivo. Bioinorganic chemistry and applications 2016: 3837679.
- 38. Wang J, Zheng Y, Zhao J, Liu T, Gao L, et al. (2012) Low-dose rhBMP2/7 heterodimer to reconstruct peri-implant bone defects: a micro-CT evaluation. Journal of clinical periodontology 39(1): 98-105.
- 39. Rahim I, Salt S, Heliotis M (2015) Successful longterm mandibular reconstruction and rehabilitation using non-vascularised autologous bone graft and recombinant human BMP-7 with subsequent endosseous implant in a patient with bisphosphonate-related osteonecrosis of the jaw. The British journal of oral & maxillofacial surgery 53(9): 870-874.
- 40. Suarez LDAF, Monje A, Padial MM, Tang Z, Wang HL (2015) Biologic Agents for Periodontal Regeneration and Implant Site Development. BioMed research international 2015: 957518.

