



Marine Actinomycetes the Past, the Present and the Future

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Abstract

In the field of exploring new natural sources for biologically active products with economic importance, the marine environment draws particular attention due to the noticeable diversity and extreme conditions; it is well known that the marine environment is a valuable source of biological and therapeutic compounds with great value. The marine environment represents a novel source for the discovery of new secondary metabolites including antibiotic, antiviral, antitumor, antifouling agents, as well as enzymes. Marine actinomycetes are widely distributed through the marine environment from shallow to sea sediments. The secondary metabolites obtained from such marine actinomycetes have proved their value in different industries due to their unique properties and structures. This review focuses on the importance of marine actinomycetes as well as their secondary metabolites. The development of new technologies for marine actinomycetes bioprospection is very promising, leading to the discovery of high-quality value products with biotechnological and medical applications.

Keywords: Marine Actinomycetes; Bioactive Compounds; Biological Activities; Commercial Use

Introduction

Actinomycetes are groups of Gram-positive bacteria that are characterized by having high G+C content (>55%) in their DNA [1,2]. Actinomycetes exhibit high economical and biotechnological priceless importance. They play a major role in the production of many novel metabolites such as enzymes, anti-parasitic agents, herbicides, pesticides, immune-modifiers, antitumour agents, enzyme inhibitors, and vitamins as well as many other biologically active products [3-6]. Moreover, actinomycetes play a vital role in the degradation of various organic wastes due to the ability of actinomycetes to produce various enzymes such as ligninase, xylans, chitinase, and pectinase, etc [5]. The majority of actinomycetes have been isolated from different terrestrial sources, however, recently the scientists focus on the isolation of different marine actinomycetes as

well as studying the production of potential metabolites from these marine strains [7,8]. More than 70% of the earth's surface has been covered by oceans that represent a suitable environment for microbial diversity including actinomycetes. Marine organisms have shown their capacity to produce a wide variety of novel bioproducts with unique structures and functional features due to the occurrence of these organisms in extreme environmental conditions such as salinity, pressure, temperature, as well as many other conditions [9,10]. The marine environments are completely different from those of the terrestrial ones, thus marine actinomycetes are expected to provide a potential source of novel compounds that differs from those produced by the terrestrial actinomycetes [11,12]. Little is known about the versatility of the marine organisms' bioactive metabolites. This could be attributed to the difficulty of the isolation of such microorganisms. However, recently with the aid of the

new technologies, the existence of actinomycetes and their biosynthesis gene clusters as well as their occurrence in various marine ecosystems have been proved [13,14].

Marine Actinomycetes Bioactive Secondary Metabolites

Among 23,000 bioactive secondary metabolites produced by microorganisms, about 1,000 are produced by actinomycetes, which represent about 45% of all discovered bioactive microbial metabolites [15]. The genus *Streptomyces* members produce approximately 7,600 compounds [15]. Between 1969 and 1999, nearly 300 patents on marine bioactive products were issued [16,17].

Actinomycetes are well known for the production of bioactive compounds with high industrial importance [18,19]. Several actinomycetes play important roles in environmental protection, mineralization of organic matter, nitrogen fixation, and immobilization of mineral nutrients, etc [20]. Among these actinomycetes, several genera have

been reported from the marine environment. Many of these marine strains produce many metabolites with biological activities and which can be developed as therapeutic and pharmaceutical agents [21]. Among these biologically active compounds are peptides, polyketides, isoprenoids, sterols, and phenazines as well as others [22, 23].

Antimicrobial Agents

Secondary metabolites with potent antimicrobial properties including antibacterial and antifungal properties have been widely used against various infectious diseases (Table1 and Table2). Thus, antibiotic-producing actinomycetes have gained great importance in the pharmaceutical industry [15]. Early in 1940, 1942, and 1943, actinomycin and streptothricin, and streptomycin produced by actinomycetes were reported respectively as effective sources of antibiotics [24, 25]. Antibiotics obtained from actinomycetes have been used in various fields including agriculture, veterinary, and pharmaceutical industries, etc [26-29].

Compound	Source	References
Amphotericin B	<i>Streptomyces nodosus</i>	Hartsel and Bolard [30]
Antimycin	<i>Streptomyces</i> sp. SCSIO 1635	Su-Mei, et al. [31]
Bonactin	<i>Streptomyces</i> sp. BD21-2	Schumacher, et al. [32]
Daryamides	<i>Streptomyces</i> sp. CNQ-085	Asolkar, et al. [33]
Natamycin	<i>Streptomyces natalensis</i>	Pedersen [34]
Nystatin	<i>Streptomyces noursei</i> ATCC 11455	Zotchev, et al. [35]
Urauchimycins (member of antimycin class) Urauchimycins A and B-from marine sponge Ni-80 Urauchimycin C-from marine sediment	<i>Streptomyces</i> sp.	Sharma, et al. [36]

*Most of the antifungal agents derived from *Streptomyces* species are macrolide polyenes.

Table 1: Examples for antifungal metabolites produced by marine actinobacteria.

Compound	Source	References
Bonactin	<i>Streptomyces</i> sp. BD21-2	(Schumacher, et al. [32])
Chandrananimycins	<i>Actinomadura</i> sp.	Maskey, et al. [37]
Diazepinomicin (ECO-4601)	<i>Micromonospora</i> sp.	Charan, et al. [38]
Frigocyclinone	<i>Streptomyces griseus</i>	Bruntner, et al. [39]
Glaciapyrroles A, B and C	<i>Streptomyces</i> sp. NPS008187	Macherla, et al. [40]
Helquinoline	<i>Janibacter limosus</i>	Asolkar, et al. [33]
Lajollamycin	<i>Streptomyces nodosus</i>	Mann [41]
Marinomycins	<i>Marinispora</i>	Kwon, et al. [26]
Rifamycin	<i>Streptomyces arenicola</i>	Floss and Yu [42]
Tetracenomycin D	<i>Streptomyces corchorusii</i>	Adinarayana, et al. [43]

Table 2: Examples for antibacterial metabolites produced by marine actinobacteria.

Antiviral Agents: Some marine actinomycetes have shown their ability to produce some antiviral agents that show various applications in various fields such as biological control of human viral infections, also it can be used in chemotherapy of humans viral diseases. Additionally, these antiviral compounds can be applied in the treatment of sewage-polluted waters. Benzastatin C produced by *Streptomyces nitrosporeus* has been used as a potent antiviral agent [44]. Another study reported a marine actinomycetes strain named *Streptomyces kaviengensis*, produced a novel metabolite “antimycin A” that showed potent antiviral activity. Antimycin A derivative was very effective against the Western equine encephalitis virus where the IC_{50} value was less than 4 nM. It was also revealed that Antimycin A exerts its antifungal activity via disrupting the mitochondrial electron transport and pyrimidine biosynthesis [45]. *Streptomyces* sp. HK18 isolated from the soil of a Korean solar saltern also produced biologically active metabolites “xiamycins C-E” with antiviral properties. Among these, xiamycin D showed the maximum antiviral effect against porcine epidemic diarrhea virus (PEDV) replication with of EC_{50} value equals to 0.93 μ M [46].

Antitumor Compounds: Cancer is a serious health problem that requires a great attention. Besides the antimicrobial properties exhibited by marine actinomycetes they also show cytotoxicity against many tumor cells. Many compounds isolated from marine actinobacteria gained great importance as antitumor compounds. These actinomycetes derived antitumor drugs belong to various structural classes [47,48], these include:

- Antimetabolites: Carzinophilin and pentostatin;
- Aureolic acids: chromomycin A3 and mithramycin;
- Eneidyne: neocarzinostatin;
- Heterocyclic quinones: mitomycin C;
- Indolocarbazoles: rebeccamycin and staurosporine;
- Polyketides: anthracyclines, daunomycin, elloramycin, geldanamycin, oviedomycin, etc.

The Antitumor compounds obtained from marine actinomycetes function via various processes. These include mitochondria permeabilization, DNA cleavage process that can be mediated by topoisomerase I or II inhibition, inhibition of vital enzymes like proteases that are involved in signal transduction, and even by the inhibition of tumor-induced angiogenesis [48].

Moreover, the family Micromonosporaceae that belongs to marine actinomycetes produce potent bioactive compounds. These strains are reported to target proteasome and hence found success in pharmaceuticals [16]. *Streptomyces chartreusis* that is first isolated from by Leach, et al., produces chartreusin that exhibits antibacterial activity as well as potent antitumor activity against various human cell lines

[49]. Another marine actinomycete designated MAR4 that belongs to the family Streptomycetaceae were also reported to produce a host of meroterpenoids belonging to the class of napyradiomycin [50-52]. These napyradiomycins were first reported for their antimicrobial activity, however, they have also been reported to inhibit gastric (H^+ - K^+) ATPases and act as antagonists for estrogen receptors. These properties allow napyradiomycins to be effective in the treatment of cancer, however, more studies are needed to define their mechanisms of action in cancer cells.

Enzymes Production

Actinomycetes are well-known producers of enzymes. Marine actinomycetes are reported to produce many enzymes with industrial importance and that have more stability and unique substrate specificities. The availability of the natural product in marine environments may rely on the ratio of enzyme produced by marine microorganisms [53]. Among the enzymes produced by marine actinomycetes are Proteases and α -Amylases, cellulases, chitinases, xylanases, ribonucleases, etc. Proteases isolated from marine actinomycetes have been purified as well as characterized [54]. Proteases have great commercial importance that is utilized in various industries, such as detergents, brewery, cheese-making, meat tenderization, and baking, etc [55]. Also, alkaline proteases have been extensively applied in other industries including textile, leather, wastewater treatment, etc. On the other side, *Streptomyces* species are well known as potent produced of amylolytic enzymes [56]. Amylases are widely applied in fermentation, food, textile, and paper industries.

Cellulase producers have been reported in the actinomycetes. These cellulolytic enzymes are applied in several industries such as cellulosic biomass pretreatment to improve its nutritional quality, pretreatment of industrial wastes, color extraction from juices, and detergents for color brightening [57,58]. Chitinases have been also reported to be produced by actinobacteria [59]. Chitinase finds a great application as a potent antifungal agent due to its ability to degrade chitin [60]. Xylanase has been applied widely in the pulp and paper industry due to the ability of xylanases to disrupt the cell wall structure of xylan at elevated temperatures. Actinobacteria have shown their capacity to produce xylanases [61]. Ribonuclease which is also known as RNase plays an important role in many biological processes, including self-incompatibility in flowering plants and angiogenesis. Several prokaryotic toxin-antitoxin systems have been reported to have RNase activity. Thus, various enzymes are being produced by marine actinomycetes and which show great industrial importance. These enzymes produced are used as pharmaceuticals, fine chemicals and food industries [62, 63].

Enzyme Inhibitors: Enzyme inhibitors are molecules that can bind to enzymes and as a result decrease or inhibit their activities. Some drugs can act as enzyme inhibitors by blocking the enzyme's activity and which in turn can correct a metabolic imbalance or even kill a pathogen. Also, they can be applied to many pesticides. The discovery and improvement of enzyme inhibitors are areas of interest in the pharmacology and biochemistry fields [64-70]. Also, some marine actinomycetes showed their ability to produce enzyme inhibitors.

Future Prospects and Scope

Future efforts in this field should include deep comprehension of microbial physiology, systematics, and metabolism. Exploring the sequencing of various actinomycete genomes as well as studying the secondary metabolite pathways found in actinomycetes are required. More effort should be directed towards the development of more techniques that help easier and more efficient isolation. Various novel compounds should be exerted.

Conclusion

Exploring the biotechnological and therapeutical active metabolites from marine actinomycetes has gained attention recently. Nevertheless, more effort should be done to address the exact pharmaceutical potency of various marine actinomycetes. The marine ecosystem is not completely unexplored for its potential compounds despite their vast resources. Actinomycetes are well known as producers of many novel compounds with pharmaceutical and clinical importance. Traditional and innovative techniques and strategies have been developed to characterize the marine actinomycete diversity as well as studying the relationship between the marine environment and the secondary metabolite produced by marine microbes. Altogether, this will help increase our ability to understand their systematics, as well as clarify their evolution and ecology. Actinomycetes have shown their ability to produce various potent secondary metabolites that are required in several important industries such as pharmaceutical, cosmetic, medical, and food industries. More effort is needed to explore the potential of marine microorganisms especially marine actinomycetes as producers of novel drugs of natural sources. Many studies worldwide have lined up the points towards the research on marine actinomycetes for searching of drug lead or new drugs.

References

1. Chater KF (2006) *Streptomyces* inside-out: a new perspective on the bacteria that provide us with antibiotics. *Philosophical Transactions of the Royal Society B: Biological Sciences* 361(1469): 761-768.
2. Das S, Lyla P, Khan S (2008) Distribution and generic composition of culturable marine actinomycetes from the sediments of Indian continental slope of Bay of Bengal. *Chinese Journal of Oceanology and Limnology* 26(2): 166-177.
3. Butler MS (2004) The role of natural product chemistry in drug discovery. *Journal of natural products* 67(12): 2141-2153.
4. Atta H (2007) Production of vitamin B 12 by *Streptomyces fulvissimus*. *Egyptian Journal of Biomedical Sciences* 23(1): 166-184.
5. Arbat AB, Zodpe SN (2014) Biodiversity of Actinomycetes species isolated from saline belt of Akola district. *Indian Journal of Applied Research* 4(7): 450-452.
6. Rashad FM, Fathy H, El-Zayat A, Elghonaimy A (2015) Isolation and characterization of multifunctional *Streptomyces* species with antimicrobial, nematicidal and phytohormone activities from marine environments in Egypt. *Microbiological research* 175: 34-47.
7. Kurtböke DI (2012) Biodiscovery from rare actinomycetes: an eco-taxonomical perspective. *Applied microbiology and biotechnology* 93(5): 1843-1852.
8. Komaki H, Sakurai K, Hosoyama A, Kimura A, Igarashi Y, et al. (2018) Diversity of nonribosomal peptide synthetase and polyketide synthase gene clusters among taxonomically close *Streptomyces* strains. *Scientific reports* 8(1): 1-11.
9. Kathiresan K, Nabeel M, Manivannan S (2008) Bioprospecting of marine organisms for novel bioactive compounds. *Scientific Transaction Environmental Technovation* 1: 107-120.
10. Kathiresan K (2019) Salt-tolerant microbes in mangroves: ecological role and bioprospecting potential. *Research Developments in Saline Agriculture* pp: 237-255.
11. Zhou M (1998) Identification of marine actinomycetes S-216 strain and its biosynthetic conditions of antifungal antibiotic. *J Xiamen Univ Nat Sci* 37: 109-114.
12. Piel J (2009) Metabolites from symbiotic bacteria. *Natural product reports* 26(3): 338-362.
13. Donadio S, Monciardini P, Alduina R, Mazza P, Chiocchini C, et al. (2002) Microbial technologies for the discovery of novel bioactive metabolites. *Journal of Biotechnology* 99(3): 187-198.

14. Janssen P, Yates P, Grinton B, Taylor P, Sait M (2002) Improved culturability of soil bacteria and isolation in pure culture of novel members of the divisions Acidobacteria, Actinobacteria, Proteobacteria, and Verrucomicrobia. *Applied and environmental microbiology* 68(5): 2391-2396.
15. Berdy J (2005) Bioactive microbial metabolites. *The Journal of antibiotics* 58(1): 1-26.
16. Kathiresan K, Balagurunathan R, Selvam M (2005) Fungicidal activity of marine actinomycetes against phytopathogenic fungi. *Indian Journal of Biotechnology* 4(2): 271-276.
17. Sithranga N, Kathiresan K (2010) Anticancer drugs from marine flora: an overview. *Journal of oncology*.
18. Tamehiro N, Hosaka T, Xu J, Hu H, Otake N, et al. (2003) Innovative approach for improvement of an antibiotic-overproducing industrial strain of *Streptomyces albus*. *Applied and environmental microbiology* 69(11): 6412-6417.
19. Higginbotham SJ, Murphy CD (2010) Identification and characterisation of a *Streptomyces* sp. isolate exhibiting activity against methicillin-resistant *Staphylococcus aureus*. *Microbiological research* 165(1): 82-86.
20. Williams ST, Mordarski M, Goodfellow M (1988) Actinomycetes in biotechnology, Academic.
21. Kieser T, M. Bibb M, Buttner K, Chater D Hopwood (2000) Preparation and analysis of genomic and plasmid DNA. *Practical Streptomyces Genetics* 1: 161-210.
22. ul Hassan SS, Anjum K, Abbas S, Akhter N, Shagufta B, et al. (2017) Emerging biopharmaceuticals from marine actinobacteria. *Environmental toxicology and pharmacology* 49: 34-47.
23. Binayke A, Ghorbel S, Hmidet N, Raut A, Gunjal A, et al. (2018) Analysis of diversity of actinomycetes from arid and saline soils at Rajasthan, India. *Environmental Sustainability* 1(1): 61-70.
24. Waksman SA (1943) Production and activity of streptothricin. *Journal of bacteriology* 46(3): 299-310.
25. Waksman SA, Woodruff H (1940) Bacteriostatic and bactericidal substances produced by a soil Actinomycetes. *Proceedings of the society for Experimental Biology and Medicine* 45(2): 609-614.
26. Comroe Jr JH (1978) Pay dirt: the story of streptomycin: Part I. From Waksman to Waksman. *American Review of Respiratory Disease* 117(4): 773-781.
27. Niu XM, Li SH, Goerls H, Schollmeyer D, Hilliger M, et al. (2007) Abyssomicin E, a highly functionalized polycyclic metabolite from *Streptomyces* species. *Organic letters* 9(13): 2437-2440.
28. Anzai K, Ohno M, Nakashima T, Kuwahara N, Suzuki R, et al. (2008) Taxonomic distribution of *Streptomyces* species capable of producing bioactive compounds among strains preserved at NITE/NBRC. *Applied microbiology and biotechnology* 80(2): 287-295.
29. Hohmann C, Schneider K, Bruntner C, Irran E, Nicholson G, et al. (2009) Caboxamycin, a new antibiotic of the benzoxazole family produced by the deep-sea strain *Streptomyces* sp. NTK 937. *The Journal of antibiotics* 62(2): 99-104.
30. Hartsel S, Bolard J (1996) Amphotericin B: new life for an old drug. *Trends in pharmacological sciences* 17(12): 445-449.
31. Su-Mei L, Xin-Peng T, Si-Wen N, Wen-Jun Z, Cai Z, et al. (2011) Antimycins from marine *Streptomyces* sp. SCSIO 1635 from the South China Sea. *Natural Product Research and Development* 23(1): 10.
32. Schumacher RW, Talmage S, Miller S, Sarris K, Davidson B, et al. (2003) Isolation and structure determination of an antimicrobial ester from a marine sediment-derived bacterium. *Journal of natural products* 66(9): 1291-1293.
33. Asolkar R, Jensen P, Kauffman C, Fenical W (2006) Daryamides A– C, weakly cytotoxic polyketides from a marine-derived actinomycete of the genus *Streptomyces* strain CNQ-085. *Journal of natural products* 69(12): 1756-1759.
34. Pedersen JC (1992) Natamycin as a fungicide in agar media. *Applied and environmental microbiology* 58(3): 1064-1066.
35. Zotchev S, Haugan O, Sekurova H, Sletta T, Ellingsen T, et al. (2000) Identification of a gene cluster for antibacterial polyketide-derived antibiotic biosynthesis in the nystatin producer *Streptomyces noursei* ATCC 11455 The GenBank accession numbers for the sequences reported in this paper are AF071512 for ORF1, AF071513 for ORF2, AF071514 for ORF3, AF071515 for ORF4, AF071516 for ORF5, AF071517 for ORF6, AF071518 for ORF7, AF071519 for *gdhA*, AF071520 for ORF8, AF071521 for ORF9, AF071522 for ORF10 and AF071523 for ORF11. *Microbiology* 146(3): 611-619.
36. Sharma M, Dangi P, Choudhary M (2014) Actinomycetes: source, identification, and their applications. *Int J Curr*

- Microbiol App Sci 3(2): 801-832.
37. Maskey R, Li F, Qin S, Fiebig H, Laatsch H (2003) Chandranamycins AC: production of novel anticancer antibiotics from a marine Actinomadura sp. isolate M048 by variation of medium composition and growth conditions. The Journal of antibiotics 56(7): 622-629.
 38. Charan R, Schlingmann G, Janso J, Bernan V, Feng X, et al. (2004) Diazepinomicin, a new antimicrobial alkaloid from a marine *Micromonospora* sp. Journal of Natural Products 67(8): 1431-1433.
 39. Bruntner C, Binder T, Pathom-aree W, Goodfellow M, Bull A, et al. (2005) Frigocyclinone, a novel angucyclinone antibiotic produced by a *Streptomyces griseus* strain from Antarctica. The Journal of antibiotics 58(5): 346-349.
 40. Macherla VR, Liu J, Bellows C, Teisan S, Nicholson B, et al. (2005) Glaciapyrroles A, B, and C, pyrrolesquiterpenes from a *Streptomyces* sp. isolated from an Alaskan marine sediment. Journal of natural products 68(5): 780-783.
 41. Mann J (2001) Natural products as immunosuppressive agents. Natural product reports 18(4): 417-430.
 42. Floss HG, Yu TW (2005) Rifamycin mode of action, resistance, and biosynthesis. Chemical reviews 105(2): 621-632.
 43. Adinarayana G, Venkateshan MR, Bapiraju V, Sujatha P, Premkumar J, et al. (2006) Cytotoxic compounds from the marine actinobacterium *Streptomyces corchorusii* AUBN 1/7 1. Russian Journal of Bioorganic Chemistry 32(3): 295-300.
 44. Lee JG, Yoo ID, Kim WG (2007) Differential antiviral activity of benzastatin C and its dechlorinated derivative from *Streptomyces nitrosporeus*. Biological and Pharmaceutical Bulletin 30(4): 795-797.
 45. Raveh A, Delekta P, Dobry C, Peng W, Schultz P, et al. (2013) Discovery of potent broad spectrum antivirals derived from marine actinobacteria. PloS one 8(12): e82318.
 46. Kim SH, Ha TKQ, Oh W, Shin J, Oh DC (2016) Antiviral indolosesquiterpenoid xiamycins C-E from a halophilic actinomycete. Journal of natural products 79(1): 51-58.
 47. Newman DJ, Cragg GM (2007) Natural products as sources of new drugs over the last 25 years. Journal of natural products 70(3): 461-477.
 48. Olano C, Méndez C, Salas J (2009) Antitumor compounds from marine actinomycetes. Marine drugs 7(2): 210-248.
 49. Leach BE, Calhoun K, Johnson L, Teeters C, Jackson W (1953) Chartreusin, a new antibiotic produced by *Streptomyces chartreusis*, a new species. Journal of the American Chemical Society 75(16): 4011-4012.
 50. Soria-Mercado IE, Prieto-Davo A, Jensen P, Fenical W (2005) Antibiotic terpenoid chloro-dihydroquinones from a new marine actinomycete. Journal of natural products 68(6): 904-910.
 51. Gallagher K, Fenical W, Jensen P (2010) Hybrid isoprenoid secondary metabolite production in terrestrial and marine actinomycetes. Current opinion in biotechnology 21(6): 794-800.
 52. Cheng YB, Jensen P, Fenical W (2013) Cytotoxic and Antimicrobial Napyradiomycins from Two Marine-Derived *Streptomyces* Strains. European journal of organic chemistry 2013(18): 3751-3757.
 53. Ramesh S, Mathivanan N (2009) Screening of marine actinomycetes isolated from the Bay of Bengal, India for antimicrobial activity and industrial enzymes. World Journal of Microbiology and Biotechnology 25(12): 2103-2111.
 54. Dixit VS, Pant A (2000) Hydrocarbon degradation and protease production by *Nocardiopsis* sp. NCIM 5124. Letters in applied microbiology 30(1): 67-69.
 55. Kumar CG, Takagi H (1999) Microbial alkaline proteases: from a bioindustrial viewpoint. Biotechnology advances 17(7): 561-594.
 56. Vigal T, Gil L, Daza A, García-González M, Martín J (1991) Cloning, characterization and expression of an α -amylase gene from *Streptomyces griseus* IMRU3570. Molecular and General Genetics MGG 225(2): 278-288.
 57. Niehaus F, Bertoldo C, Kähler M, Antranikian G (1999) Extremophiles as a source of novel enzymes for industrial application. Applied microbiology and biotechnology 51(6): 711-729.
 58. Bhat MK (2000) Cellulases and related enzymes in biotechnology. Biotechnology advances 18(5): 355-383.
 59. Pisano MA, Sommer MJ, Taras L (1992) Bioactivity of chitinolytic actinomycetes of marine origin. Applied microbiology and biotechnology 36(4): 553-555.
 60. Kunz C, Ludwig A, Bertheau Y, Boller T (1992) Evaluation of the antifungal activity of the purified chitinase 1 from the filamentous fungus *Aphanocladium album*. FEMS microbiology letters 90(2): 105-109.

61. Bode W, Huber R (1993) Natural protein proteinase inhibitors and their interaction with proteinases. *EJB Reviews* pp: 43-61.
62. Oldfield C, Wood N, Gilbert S, Murray F, Faure F (1998) Desulphurisation of benzothiophene and dibenzothiophene by actinomycete organisms belonging to the genus *Rhodococcus*, and related taxa. *Antonie Van Leeuwenhoek* 74(1): 119-132.
63. Hough DW, Danson MJ (1999) Extremozymes. *Current opinion in chemical biology* 3(1): 39-46.
64. Garcia-Fernandez L, Reyes F, Sanchez-Puelles J (2002) The marine pharmacy: new antitumoral compounds from the sea. *Pharmaceutical News* 9(6): 495-502.
65. Imada C (2005) Enzyme inhibitors and other bioactive compounds from marine actinomycetes. *Antonie Van Leeuwenhoek* 87(1): 59-63.
66. Lam KS (2006) Discovery of novel metabolites from marine actinomycetes. *Current opinion in microbiology* 9(3): 245-251.
67. Asolkar RN, Schroeder D, Heckmann R, Lang S, Wagner-Doebler I, et al. (2004) Helquinoline, a new tetrahydroquinoline antibiotic from *Janibacter limosus* Hel 1+. *The Journal of Antibiotics* 57(1): 17-23.
68. Kwon HC, Kauffman C, Jensen P, Fenical W (2006) Marinomycins A- D, antitumor-antibiotics of a new structure class from a marine actinomycete of the recently discovered genus "Marinispora". *Journal of the American Chemical Society* 128(5): 1622-1632.
69. Elkhateeb WA, Elnahas MO, Daba GM (2021) Actinotherapy: Highlights on the Pharmaceutical Potentials of Actinomycetes. *Open access journal of microbiology & biotechnology* 6(2): 1-7.
70. Elkhateeb WA, Mohamed MA, Fayad W, Emam M, Nafady IM, et al. (2020) Molecular Identification, Metabolites profiling, Anti-breast cancer, Anti-colorectal cancer, and antioxidant potentials of *Streptomyces zaomyceticus* AA1 isolated from a remote bat cave in Egypt. *Research Journal of Pharmacy and Technology* 13(7): 3072-3080.

