



A Scooping Review on Anesthesia in Wild Animals

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Abstract

Different ancient cultures kept wild animals in captivity in collections, but there are essentially no records of the veterinary care that may have been provided to these animals. Live capture of wild animals by chemical immobilization was introduced in the 1950's. As a result, several novel capture techniques, darting tools and drug administration techniques have been created and applied to a variety of wildlife species. Even though captive wildlife in zoological collections is more protected they still pose challenges compared to domestic animal surgery. In captive, immobilization of wild animals is often required for health examination and for diagnosis and treatment of disease. Improvement of chemical capture is an important part of wildlife conservation and animal welfare to minimize distress for the animals and the risk of morbidity and mortality. Veterinarians warn that such field anesthesia is especially difficult because the pre-anesthetic evaluation of an animal's health is restricted to characteristics that are easily observed. Darting from a helicopter to the ground as a means of capture, followed by physical restraint and drug injection. Radio collaring is essential for understanding an animal's movement patterns, but capture and immobilization can alter an animal's activity pattern for days to weeks following capture. Many wild animals are particularly dangerous, and it is essential that the animal reaches deep level of sedation or anesthesia before any human gets close and that it remains in this level for the entire procedure. As well as for wild animals in particular, where residual sedation or re-narcotization can cause injury after release and where it is crucial to return to normal behavior as soon as possible, anesthesia recovery should be quick and complete. Veterinarians should be knowledgeable about the physiology, behaviour, and natural history of the target species in order to achieve a better chemical immobilization while capturing wild animals. All equipment must also be thoroughly checked before usage.

Keywords: Anesthesia; Capture; Chemical; Darting; Immobilization; Wildlife

Introduction

The veterinary care that may have been given to wild animals kept in captive in collections by many ancient

societies is almost undocumented [1]. Surgical treatments in captive wildlife species may help preserve endangered wildlife species as part of captive managed breeding programs [2]. Similar to veterinary care for humans and

domestic animals, the best care for wildlife should ideally be supported by evidence [3]. In the 1950s, live capture of wild animals by chemical immobilization was first used [4]. This led to the development of a number of new methods for drug administration, darting, and capturing that have been used on a range of animal species [5]. Despite being more protected, caged wildlife in zoological museums still presents difficulties when compared to operations on domestic animals [6]. Even within the same taxonomic group, body sizes vary greatly. MIS is likely to have benefits in captive or free-ranging wildlife, just as it does in domestic animals, despite the weakness of the current evidence base and the paucity of studies addressing safety, techniques, and specific applications applicable to wildlife veterinary patients [7]. However, the benefits of using MIS techniques in the medical treatment of wildlife animals in captivity and in the wild may be even greater. Additionally, it poses special challenges not found in surgery performed on domestic animals or on humans. Detection and post-operative care are difficult tasks [8].

Wild animals kept in captivity must frequently be immobilized in order to undergo health examinations, diagnose illnesses, and administer treatment. Free-ranging wild animals are frequently immobilized in isolated areas under the most trying conditions. Drugs should ideally have a broad safety margin because body weight and health status are typically unknown when wild animals are darted. A strong, concentrated medication that may be administered in tiny doses is required for remote darting. Because capture in the wild causes a great deal of stress and excitement, free-ranging animals frequently need larger dosages of drugs than caged wildlife. Rapid induction is essential to lower the possibility that the animal would sustain injuries on the ground or become obscured in dense. The use of medications whose effects can be reversed by an antagonist is advantageous in two emergency situations and speeds up the process. It is crucial for the animal to have rapid movement and self-defense skills in the wild. Assessing whether blood flow to essential organs is expected to be maintained and if oxygen supply is likely to satisfy tissue needs are important goals of anesthetic monitoring [9]. Under immobilization and anesthesia, mucous membrane color, refill time, and refill vigor are still useful as overall indicators of perfusion, but they might not accurately represent the blood flow to vital organs.

One of veterinary anesthesia's most difficult tasks is chemically immobilizing and capturing wild animals [10]. Remarkably, just a small portion of clinical data from domestic species can be directly used for this purpose [11]. For instance, the dose of an immobilizing chemical can only be calculated using the animal's estimated body weight because the animal's weight and physiological condition

cannot be ascertained until it has been immobilized. In any situation involving the immobilization of wild animals, careful preparation and planning are crucial. Every piece of equipment needs to be inspected before use, and a veterinarian needs to be knowledgeable with the physiology, behavior, and natural history of the target species [12]. Injuries, allergic reactions, and zoonotic infections are among the dangers associated with animals [13].

Since many wild animals are extremely hazardous, it is crucial that the animal achieve a profound level of sedation or anesthesia before any human approaches and that it stays there during the entire process [14]. For field primatologists, capture is one of the most important ethical issues [15]. An animal's movement patterns can be understood through radio collaring, although immobility and capture might change an animal's behavior for days or weeks after capture [16]. Delivering and maintaining anesthesia with current medications still carries a number of serious hazards for zoo patients [17]. Premedication to provide tranquilization or light sedation to allow intravascular catheter placement, followed by intravenous induction and maintenance with either inhalation or intravenous agents.

However, huge wild cats don't usually accept catheter implantation with only mild sedation, and they can't be easily or safely injected with pre-medicants. Therefore, initial immobilization to make the animal safe to approach and handle, followed by the administration of maintenance medications, if necessary, is the standard procedure for wild felid anesthesia in order to minimize patient discomfort and enhance personnel safety [18]. The optimal anesthetic drug should preserve steady cardiopulmonary function while enabling flexible anesthetic depth modification. Rapid and thorough recovery from anesthesia is necessary, especially for wild animals were returning to normal behavior as soon as possible is crucial since residual sedation or re-narcotization may cause harm after release [6]. In different countries there are a variety of findings regarding effect of chemical immobilization and capture of wild animals in the form of review paper and studies. However, in Ethiopia there is there is a shortage of reviews on this topic our continent that support this point.

Therefore, the objectives of this review are:

- To provide sufficient information regarding the effects of anesthesia, the main difference between wild and domesticated animals,
- To indicate wild animals' reaction to the medication and how difficult it is to anesthetize them.
- To describe the importance and benefits of wild animals.
- To provide veterinarians with a fundamental knowledge of the method for capturing wild animals.

Literature Review

Anesthesia was first successfully demonstrated at the Massachusetts General Hospital in Boston, in 1846 by William T Morton. Medications or combinations of medications known as general anesthetic drugs are thought to be essential therapeutic agents because they cause unconsciousness, reduce pain, and increase muscular reflectivity [19]. Proper usage of general anesthesia interventions in operative settings can significantly decrease the mortality and disability rate [20]. Because of their various qualities, a variety of anesthetic drugs are frequently employed in clinical contexts [21]. People have always been captivated by wild animals. In many societies around the world, a variety of wildlife species have religious and spiritual significance. Ancient human populations already relied heavily on wild animals for nourishment. A large portion of the population still consumes bush meat or wild animals. Allows for the better use of existing technology to reduce the danger of anesthesia, which is crucial because technological advancements in the delivery of anesthesia are rare.

Guidelines

Provide information to veterinarians and biologists on how to carry out anesthesia and surgery on wild animals in the field at a technical level that is acceptable [18]. Institutional Animal Care and Use Committees (IACUCs), or ethics committees can use these standards of care to assess the suitability of methods suggested in studies that are submitted to them [22]. Furthermore, when proposing to animal ethics committees, biologists and veterinarians can use these guidelines to determine the technical level at which their fieldwork would be conducted [18]. In order to account for evolving opinion techniques, procedures, monitors, and medications, we expect these standards of care to be modified and updated over time [23].

Considerations

The central nervous system is depressed during general anesthesia, which causes loss of feeling and consciousness [24]. Among the hallmarks of general anesthesia are 1) loss of consciousness 2) analgesia, 3) relaxation of the muscles, and 4) lack of reflex reactions. When working with free-ranging wildlife, where residual sedation or re-narcotization may cause harm or death after release, anesthesia must also be a quickly reversible process [18]. Almost always, a licensed veterinarian must obtain the medications required for immobilization, which are either prescription or prohibited narcotics. Biologists and field technicians provide the majority of field immobilizations, in contrast to domestic animal anesthetic. Only after receiving the appropriate training from veterinarians or other

professionals should non-veterinary staff administer and oversee anesthesia.

Capture Conditions

Wildlife species are affected differently by the different methods of chemical restraint and capture used [18]. This was already written forty years ago by Arthur M. Pearson of the Canadian Wildlife Service in Yukon. The anesthetist must be capable of safely administering the chosen medicine combination in a field situation. Compared to remote drug delivery to an unrestrained animal, physical restriction in a trap is frequently slower and less stressful for the animal [25]. A slow induction approach increases the risk of overexertion and myopathy [14], as well as the chance of escape or injury from environmental hazards.

Primates can be chemically restrained by remote drug delivery or chemically immobilized by injection upon capturing. The practice may be growing as a result of developments in telemetry and radio tracking [26], which necessitate the capture of monkeys in order to attach devices. While new neuro-modulatory drugs have altered the safety of veterinary anesthesia, the aim of anesthetic management in a zoological context is to increase patient anesthetic safety without compromising human safety [27]. Increasing the efficiency of wildlife capture and immobilization is crucial for both animal welfare and wildlife conservation [4].

Types of Restraint and the Target Species

The choice of medicine and administration technique can be influenced by factors such as size, habitat, and activity level. Planning captures around seasonal peaks in pregnancy, delivery, egg laying, and young rearing requires an understanding of life history and behavior. For every catch, the relative safety and practicality of chemical, mechanical, or manual constraint must be taken into account [18]. Animals can be physically or mechanically restrained based on their level of stress, the degree of invasiveness, the predicted pain, and the expected length of time.

Removals of Water and Food Prior to anesthesia, the majority of veterinary patients are fasted in order to avoid regurgitation or vomiting and the aspiration of stomach contents [28]. In the first several hours following capture, wild animals hardly ever eat or drink. The time needed to move an animal from the location of capture to the site of surgery is frequently enough for the animal to fast. Pre-anesthetic assessment of patients, the cause for anesthesia has an impact on the patient evaluation's goal. Eliminating animals with conditions 7 (injuries, apparent sickness, etc.) that make them unrepresentative of the general population and could consequently introduce bias into the study is a key

objective of preanesthetic evaluation for research studies. When treating wildlife patients, the veterinarian lacks does not have the luxury of medical history or many preanesthetic tests. However, some degree of evaluation should be done; even large animals such as whales can be inspected for visible abnormalities such as wounds, atypical behavior and poor body condition [29].

Nature of Wildlife

The creation and application of thorough procedures and methods for managing dangerous animal crises serve as proof of this broad legislative emphasis on accountability for safety as well as advancements in program implementation and health and safety planning [30]. Drug bioavailability may be particularly unpredictable, particularly when using oral premedication and distant drug application that relies on full intramuscular delivery [10], supplementing in the face of uncertainty may increase the danger of overdosing.

Anesthetic Protocols Anesthetic Drug Types

The most often used wildlife anesthetic drugs are described in this paper along with some basic information about them. Inhalation and injection are the two main types of anesthetic drugs. Intravenous (IV) and intramuscular (IM) routes are used to administer injectable agents. Certain medications, like ketamine, can be taken by either route, while others, like propofol, can only be administered intravenously. Agents to inhale are widely utilized in clinical settings; however, inhalant anesthetics have special benefits and drawbacks when administered in the field. The two most often used inhalant agents in field veterinary medicine are isoflurane and sevoflurane. In contrast to injectable anesthetics, inhalants are given and released through the respiratory system. This enables the patient's anesthesia depth to be quickly and precisely adjusted. The requirement to carry volatile fluids, the cost and size of the vaporizers and the logistical challenges of moving compressed gases are some drawbacks. Good somatic analgesia is provided by ketamine [31]. On its own, it can cause convulsions, heat, excessive salivation, catecholamine release, and increased muscular tone [32]. There is no documented antagonism with ketamine [33]. Although they are strong depressants of the central nervous system, alpha-2 adrenergic agonists, including xylazine, detomidine, medetomidine, dexmedetomidine, and romifidine, do not cause general anesthesia unless taken in combination with another medication [34]. This group of medications has analgesic, sedative, and muscle relaxant qualities. These medications contribute to dependable anesthesia when taken in conjunction with opioids or dissociative anesthetics.

It is possible to reverse the clinical effects of α -2-agonist medications with atipamezole [35]. Though they lack the

muscular relaxation that alpha-2 agonists deliver, they are comparatively fast-acting and offer drowsiness and analgesia. Anesthesia-related hazards a patient's physical state, the length of anesthesia, the degree of preoperative stabilization, the anesthetics used, the resources available, the working environment, and the anesthetist's and surgeon's skills all affect risk. The central nervous system is intoxicated by drugs under controlled, reversible anesthesia. Since anesthetics induce distinct substrates, most likely at different parts of the central nervous system, anesthesia itself is a uniquely complex state. In ways that are unique to various types of agents [36]. A toxic phase linked to milder anesthesia planes [37]. Complex relationships between the endocrine system, sympathetic nervous system, and muscle activity may also play a role in the development of the various capture myopathy syndromes. Some abnormal breathing patterns during anesthesia are directly related to medullary depression, apnea is prevalent after induction of anesthesia, and bradypnea may develop with excessive anesthetic depth. It is also linked to increased risks of anesthesia overdose, longer recovery from anesthesia, and potential coagulation problems as well as a higher risk of wound infection after surgery [38].

Acid-base disorders, electrolyte imbalances, and changes in glycemic states are prominent examples of potentially fatal homeostatic disturbances that also require close monitoring, especially in patients at-risk. Metabolic disorders may well reflect the combined effects of multiple system dysfunctions. The gastrointestinal system is affected by anesthetics in a number of ways, some of which may be especially important for the prognosis of the patient. The majority decrease gastrointestinal motility and can be linked to regurgitation, emesis, and gastroesophageal reflux [39]. Ruminants are especially vulnerable to ileus and regurgitation brought on by anesthesia [40], and the cardiovascular effects of gastric tympany can quickly become fatal.

Particular Challenges Associated with Wildlife Anesthesia

The practicalities of anesthetizing captive wildlife frequently include more risks for the patient and the personnel performing the surgery compared to anesthesia for humans and domestic animals. These dangers might be even more pronounced in animals that roam freely. The dangers associated with anesthesia are generally higher for larger animals [41]. Comparing equine practice to small animal practice, for instance, a greater variety and frequency of perioperative and postoperative problems are seen [42].

Immobilization of wild animals using chemicals, both invasive (like surgery) and noninvasive (like blood collection, collaring, and metabolism) operations frequently call for

field wildlife anesthetic. Loss of awareness, relaxation of the muscles, analgesia, and the inhibition of stress reactions are all included in anesthesia [43]. For standard operations like clinical examinations, sample collection, collaring, contraception, or medical treatment, they are frequently immobilized. Additionally, immobilization is frequently necessary during relocations for breeding efforts [44]. For temporary immobilization, ketamine plus xylazine or ketamine and medetomidine may be used as an alternative [45]. Wild African lions have been successfully immobilized for 45 minutes using the more recent ketamine-free combination of butorphanol, midazolam, and medetomidine (BMM) [46].

Conclusion and Recommendations

General anesthetic agents are drugs or drug combinations that cause loss of consciousness, pain relief, and muscular reflexivity. They are also considered as essential therapeutic agents. In surgical settings, proper use of general interventions can significantly reduce the death and suffering rates. Many anesthetic agents are commonly used in clinical settings due to their different properties. However, many wild animals are particularly dangerous, and risks include injuries, allergic reactions, and zoonotic infections. So, it is important that the animal enters a deep state of sedation or anesthesia before any human approaches and that it maintains this level for the entire treatment. As well as recovery from anesthesia should be rapid and complete particularly for wild animals where residual sedation or re-narcotization can result in injury following release, and where return to normal behavior as soon as possible is essential. The natural history, behavior, and physiology of the target species must also be understood by the veterinarian, and all equipment must be examined before usage. Including when choosing between chemical and physical methods of capture or restraint, consideration must be given to pain, animal stress, and both human and animal safety.

The following recommendations were forwarded based on the preceding conclusion:

- For a better chemical immobilization when capture of wild animals, veterinarians should be familiar with the natural history, behavior, and physiology of the target species, and all equipment must be checked before use.
- It is important to keep an eye on the body's physiological indicators while under chemical immobilization and the technician should be let rest to the animal for five minutes before approaching it.
- Supplementation with additional oxygen is strongly advisable to prevent hypoxemia in any case of chemicals immobilization.
- Antagonist naltrexone and atipamezole intramuscularly have to be used to reverse the anesthetic effect of BAM.

Contributions

All authors made a substantial, direct, and intellectual contribution to the work.

Conflict of Interest

The authors declare that they have no conflict of interest.

References

1. Poo S (2022) Bridging the research gap between live collections in zoos and preserved collections in natural history museums. *Bio Science* 72(5): 449-460.
2. Fountain K, Barbon A, Gibbon M J, Lloyd DH, Loeffler A, et al. (2022) *Staphylococcus aureus* lineages associated with a free-ranging population of the fruit bat *Pteropus livingstonii* retained over 25 years in captivity. *Scientific Reports* 12(1): 13457.
3. Greevy P, Thomson P, Dhand NK, Raubenheimer D, Masters S, et al. (2017) Vet Compass Australia: a national big data collection system for veterinary science. *Animals* 7(10): 74.
4. Board J (2016) Advances in animal welfare for free-living animals. *Journal of Wildlife Diseases* 52(2): 4-13.
5. Latham ADM, Davidson B, Warburton B, Yockney I, Hampton JO (2019) Efficacy and animal welfare impacts of novel capture methods for two species of invasive wild mammals in New Zealand. *Animals* 10(1): 44.
6. Chinnadurai SK, Mori B, Gai J (2022) Animals in zoos, aquaria, and free-ranging wildlife. In: Rollin BE, et al. (Eds.), *Ethics in veterinary practice: Balancing conflicting interests*. Wiley and Sons.
7. Pizzi R, Masters N, Barrows M, Cracknell J, Girling S, et al. (2014) Captive wildlife surgery complication rates in zoological collections in the United Kingdom over a 25 years period. *Analysis of applications and outcomes of minimally invasive surgical techniques in wildlife*, pp: 66.
8. Sanchez KL, Handayani AB, Nelson CL, Eng JW, Prameswari W, et al. (2016) Laparoscopic appendectomy to remove a metallic foreign body in a wild Bornean orangutan (*Pongo pygmaeus*) undergoing rehabilitation. *Journal of Medical Primatology* 45(6): 327-329.
9. West G, Heard D, Caulkett N (2014) Zoo animal and wildlife immobilization and anesthesia.
10. Caulkett NA, Arnemo JM (2015) Comparative anesthesia

and analgesia of zoo animals and wildlife. *Veterinary anesthesia and analgesia: The fifth edition of Lumb and Jones* pp: 764-776.

11. Schmidt HG, Mamede S (2015) How to improve the teaching of clinical reasoning: a narrative review and a proposal. *Medical Education* 49(10): 961-973.
12. Andrews RD, Baird RW, Calambokidis J, Goertz CE, Gulland FM, et al. (2019) Best practice guidelines for cetacean tagging. *Journal of Cetacean Research and Management* 20(1): 27-66.
13. Liguori G, Costagliola A, Lombardi R, Paciello O, Giordano A (2023) Human-animal interaction in animal-assisted interventions (AAI): Zoonosis risks, benefits, and future directions -A One Health Approach. *Animals* 13(10): 1592.
14. Arnemo JM, Ahlqvist P, Andersen R, Berntsen F, Ericsson G, et al. (2006) Risk of capture- related mortality in large free- ranging mammals: experiences from Scandinavia. *Wildlife Biology* 12(1): 109-113.
15. Fedigan LM (2010) Ethical issues faced by field primatologists: Asking the relevant questions. *American Journal of Primatology* 72(9): 754-771.
16. Morellet N, Verheyden H, Angibault JM, Cargnelutti B, Lourtet B, et al. (2009) The effect of capture on ranging behavior and activity of the European roe deer *Capreolus*. *Wildlife Biology* 15(3): 278-287.
17. Dougherty N (2020) Comparative assessment of physiological homeostasis in zoo mammals under general anaesthesia: a thesis presented in partial fulfilment of the requirements for the degree of Master of Veterinary Science in Wildlife Health at Massey University, Palmerston North, Manawatū, New Zealand.
18. Chinnadurai SK, Strahl D, Fiorello CV, Harms CA (2016) Best-practice guidelines for field-based surgery and anesthesia of free-ranging wildlife I. *Journal of Wildlife Diseases* 52: 14-27.
19. Sun LH, Fan YY, Wang X, Zheng HB (2020) Pharmacodynamic elucidation of glutamate and dopamine in ketamine-induced anaesthesia. *Chem Biol Interact* 327: 109-164.
20. Cherian M, Choo S, Wilson I, Noel L, Sheikh M, et al. (2010) Building and retaining the neglected anaesthesia health workforce: is it crucial for health systems strengthening through primary health care? *Bulletin of the World Health Organization* 88(8): 637-639.
21. Dawson J, Jones M (2013) The principles of anesthesia. *Surgery* 31: 67-71.
22. Sikes RS, Gannon WL (2011) Guidelines of the American Society of Mammalogists for the use of wild mammals in research. *Journal of Mammalogy* 92(1): 235-253.
23. Capiou S, Veenhof H, Koster RA, Bergqvist Y, Boettcher M, et al. (2019) Official international association for therapeutic drug monitoring and clinical toxicology guideline: development and validation of dried blood spot- based methods for therapeutic drug monitoring. *Therapeutic Drug Monitoring* 41(4): 409-430.
24. Gardenhire DS (2015) Drugs affecting the central nervous system. *Rau's Respiratory Care Pharmacology-E-Book*, pp: 330.
25. Cattet M, Boulanger J, Stenhouse G, Powell RA, Reynolds MJ (2008) An evaluation of long-term capture effects in ursids: implications for wildlife welfare and research. *Journal of Mammalogy* 89(4): 973-990.
26. Setchell JM, Curtis DJ (2001) Field and laboratory methods in primatology: a practical guide. Cambridge University Pres, UK.
27. Taylor P (2014) Veterinary anaesthesia and analgesia: from chloroform to designer drugs. *Veterinary Record* 174 (13): 318-321.
28. Injection AA (2023) Food for Thought-Pre-anaesthetic. International Air Transport Association, Dangerous goods regulations. In Dangerous goods regulations. International Air Transport Association.
29. Moore M, Walsh M, Bailey J, Brunson D, Gulland F, et al. (2010) Sedation at sea of entangled North Atlantic right whales (*Eubalaena glacialis*) to enhance disentanglement. *PLoS One* 5(3): 95-97.
30. Hammond EE (2019) Veterinary Occupational Health and Safety in the Zoo and Wildlife Setting. In *Fowler's Zoo and Wild Animal Medicine Current Therapy* 9: 53-58.
31. Abdollahpour A, Saffarieh E, Zoroufchi BH (2020) A review on the recent application of ketamine in management of anesthesia, pain, and health care. *Journal of Family Medicine and Primary Care* 9(3): 1317.
32. George SP (2015) A comparative study of hemodynamic effects of induction doses of propofol thiopentone and propofol-ketamine combinations. *J Evid Based Med* 4(57): 1-8.
33. Zeiler F (2015) Early use of the NMDA receptor antagonist ketamine in refractory and super refractory

- status epilepticus. *Critical Care Research and Practice* 2: 1-5.
34. Lehner A (2022) Guanabenz in the horse—A preliminary report on clinical effects and comparison to clonidine and other alpha-2 adrenergic agonists. *Pferdeheilkunde* 38(6): 554-565.
35. Entire MS (2020) The effects of procedure duration and atipamezole administration on hyperkalemia in tigers (*Panthera tigris*) and lions (*Panthera leo*) anesthetized with α -2 agonists. *Journal of Zoo and Wildlife Medicine* 51(3): 490-496.
36. Perouansky M, Pearce RA, Hemmings HC (2010) Inhaled anesthetics: mechanisms of action. *Miller's Anesthesia* 19: 487-508.
37. Meyer RE (2015) Euthanasia and humane killing. *Veterinary Anesthesia and Analgesia*. In: 5th (Edn.), of Lumb and Jones, John Wiley and Sons pp: 130-143.
38. Grimm KA, Lamont LA, Tranquilli WJ, Greene SA, Robertson SA (2015) *Veterinary Anesthesia and Analgesia*. In: 5th (Edn.) of Lumb and Jones.
39. Adams JG, Figueiredo JP, Graves TK (2015) Physiology, pathophysiology, and anesthetic management of patients with gastrointestinal and endocrine disease. *Veterinary Anesthesia and Analgesia*. In: 5th (Edn.), of Lumb and Jones, John Wiley and Sons, pp: 639-677.
40. Donnell WN, Kerr CL (2015) Physiology, pathophysiology, and anesthetic management of patients with respiratory disease. *Veterinary Anesthesia and Analgesia*, pp: 511-555.
41. Hall LW, Clarke KW (1983) *Veterinary Anaesthesia*. In: 8th (Edn.), Bailliere Tindall, St. Anne's Road.
42. Brodbelt DC, Flaherty D, Pettifer GR (2015) *Anesthetic risk and informed consent. Veterinary Anesthesia and Analgesia: Fifth Edition of Lumb and Jones*. John Wiley and Sons pp: 11-22.
43. Muir WW, Hubbell JA (2008) *Equine anesthesia: monitoring and emergency therapy*. Elsevier Health Sciences. *Monitoring the depth of anaesthesia*. *Sensors* 10(12): 10896-10935.
44. Kaczmarek M, Bruczynska M, Wojciechowska M, Klich D, Glowacz K, et al. (2016) Rules of capture and transport of wisents from Poland to other European countries. *European Bison Conservation Newsletter* 9: 71-86.
45. Semjonov A (2020) Evaluation of a fixed-dose combination of butorphanol-azaperone-medetomidine (BAM) for chemical immobilisation of African lion, blesbok, and cheetah.
46. Wenger S, Buss P, Joubert J, Steenkamp J, Shikwambana P, et al. (2010) Evaluation of butorphanol, medetomidine and midazolam as a reversible narcotic combination in free-ranging African lions (*Panthera leo*). *Veterinary Anaesthesia and Analgesia* 37(6): 491-500.