

The Chicken Heterophil- A Short Review

Anand Laxmi N*

Principal Scientist, ICAR- Directorate of Poultry Research, India

***Corresponding author:** Anand Laxmi N, Principal Scientist, ICAR- Directorate of Poultry Research, India Tel: 08950735726; Email: antianand@gmail.com

Mini Review

Volume 4 Issue 1

Received Date: January 01, 2019

Published Date: January 18, 2019

DOI: 10.23880/oajvsr-16000169

Mini Review

In the immune system of chickens along with other white blood cells namely lymphocytes, monocytes, eosinophils and basophils, heterophils are also found. Heterophils are granulocytes. The cytoplasm contains eosinophilic granules. The nucleus of mature heterophils is lobed. They are responsible for bactericidal activity. Heterophils are also classified as immature, mature and toxic heterophils. Toxic heterophils exhibit toxic changes in response to the severity of the illness [1,2]. Acquired and innate immune systems are well developed in chickens. Heterophils are the components of these two systems, protect the birds from pathogens.

Heterophils were described as early as in 1846 by Wharton-Jones [3]. Heterophils in chicken are similar to neutrophil white blood cells of humans. They are involved in defense mechanisms against pathological or inflammatory conditions. Innate and acquired immune systems play an important role in protecting the organisms against diseases. At hatch large number of heterophils is released from spleen which decline 7d post hatch. During the first few days post hatch, the innate system is not well developed. Hence the function of heterophils is also not well developed [4-7] when compared with their counter older age chickens. Hence during early days of post hatch period chicken are more vulnerable to infections. The heterophil to lymphocyte ratios are taken for assessment of stress [8]. It is also stated that this ratio is affected by plasma corticosterone levels. Even fasting and social stress increases H/L ratio in chickens. Heterophils generally outnumber lymphocytes in chicks during neonatal stage. Their numbers increase during moderate stressful conditions and consequently the heterophil/lymphocyte ratio can be used to detect the presence of physiological stress. Study by Cotter, et al. [9]

suggested that estimation of single parameter such as H/L ratio cannot indicate stress. Other blood cells abnormalities also have to be taken in to account. During stressful conditions depending on physiological demand like during peak egg production phase and during molting it was observed that H/L ratio increased significantly and in addition also had effect on corticosteroid and thyroid hormones [10].

Heterophil function and cytokine gene expression studies have been used to show the resisting power of birds to salmonella infection [11,12]. In chickens, heterophils extrude granules and chromatin like structure forming extracellular traps, upon stimulation. The traps contain DNA, histone-DNA complex and elastase from heterophil cytoplasmic granules [13]. Microbial molecules stimulate degranulation. The granules are large rod shaped, medium-oval and small-core type are known [14].

Heterophils are the predominant granulocytes; they are recruited to the site of infection and are capable of killing pathogens. On exposure to pathogens, cytokines like interleukins IL-6, IL-8, IL-18 are released, which act against pathogens [15,16]. Increased production of cytokine RNA may result in population of heterophils which are primed and are efficient in responding to the pathogens [12]. Number of heterophils reaching the infection site depends on the local production of chemo attractants [17]. Early response of heterophils to pathogens is by activation and transport according to chemotactic nature.

Heterophils release granular substances, which may be proteins, peptides and other toxic substances upon encountering pathogens. The granules contain matrix-

metalloproteinases enzymes which aid in migration and in conjunction with granular substances help in killing of pathogens [18-20]. Their cytoplasmic granules contain several lysosomal and non-lysosomal enzymes including acid phosphatase, arylsulphatase, β -glucuronidase, phosphorylase, uridine diphosphate glucose-glucogen glycosyltransferase, neutral and acid α -glucosidases, acid trimetaphosphatase and lysozyme [21].

Another phenomenon known as oxidative burst is also associated with granulocytes. They lack Myeloperoxidase enzyme, due to which they are not able to produce enough amount of peroxide ion which is involved in killing of pathogens. Hence in chickens heterophils depend on non oxidative antimicrobial reactions by using defensins and cathelicidins [20,22]. Contradictory reports with respect to presence of MPO enzyme are also available [23].

It has also been reported that phagocytosing, degranulation and oxidative burst are also linked to the genetic nature of the chicken [24,25]. Different breeds have different heterophil response to *S. Enteritidis* infection. In their study it was shown that, Fayoumi breed has highest level of heterophil response when compared to white leg horn and broilers. Genetic studies revealed that polymorphisms in the genes were the reason for differential activity of heterophils [26]. This is with respect to differential ability of heterophils for production of cytokines against pathogen. Based on resistance to *S. enteritidis*, to heterophil expression, phenotypic selection of chickens has been conducted.

In neonatal chickens heterophils confer resistance to salmonella infections more than the monocyte cells [12]. Selection methods for production performance in chickens have compromised with immune functions. It was observed that heterophils of chickens infected with Staphylococcal tenosinovitis were more active when compared with the function of heterophils of healthy chickens.

When bacteria stimulate Toll like receptors on heterophils it stimulates its bactericidal functions [27]. TLR 2 and TLR 4 ligands activate heterophils for production of different cytokines and interferons, these are usually bacteria and viruses. Type 1 and P fimbriae, curli, aerobactin, lipopolysaccharide (LPS), K1 capsular antigen etc. are virulence factors associated with pathogenic *E. coli* [28]. Heterophils also possess Fc receptors and complement receptors. These receptors act through signalling pathways. Signalling pathways are mediated through G proteins, Ca and Protein Kinase C

dependant pathways. The scavenger receptors of heterophils, stimulated by ligands caused oxidative burst and not degranulation [29]. Dectin 1 and mannose receptors are also present [30]. The Toll like receptors (TLR) on heterophils has conserved signalling system that determines the inflammatory response. TLR pathways have been reported in heterophils. TLR activation also leads to the production of cytokines through activation of MAPK and nuclear factor κ B pathway [31,32].

Studies on infection of heterophils with New castle disease virus (NDV) *in vitro* showed that upon infection, they have marked reduction in phagocytosing bacteria and marked fragmentation of DNA [33]. This indicates that heterophils need not be always activated but their function can decrease depending on the severity of infection.

Heterophils contain large amount of Cathelicidin-2, localized in the large rod-shaped granules. It has both bactericidal and fungicidal activity. It is suggested that they contribute greatly to innate immunity. When broilers were challenged with *Salmonella enteritidis*, heterophils containing Cathelicidin-2 were found in the jejunum region [34]. The same peptide has been shown to act against *S. aureus* [35]. It kills the bacteria by passing through the bacterial membrane and binding to inner components causing damage to integrity of membrane.

A study conducted on two lines of broilers revealed that heterophils from one line were more responsive than the other line. This was attributed to the differential synthesis of chemokines which in turn governs protection against bacterial infections. There was increased activity of protein Tyrosine kinase and specific MAP kinase pathway [36]. Studies on regulation of different pathways and cellular modulation in future may help us to tackle and utilise the functions of heterophils for better survival of chickens.

References

1. Thrall M (2004) Veterinary haematology and clinical chemistry, 2nd (Edn.), Wiley-Blackwell, pp: 776.
2. Feldman B (2000) Schalm's Veterinary Haematology. In: Douglas JW and Wardrop KJ, (Eds.), 5th (Edn.), Williams and Wilkins, Philadelphia.
3. Wharton-Jones T (1846) The blood corpuscle considered in its different phases of development in the animal series. Memoir 1. Vertebrata. Philosophical Transcripts of the Royal Society, London 136: 63-87.

4. Genovese KJ, Moyes RB, Genovese LL, Lowry VK, Kogut MH (1998) Resistance to *Salmonella enteritidis* organ invasion in day-old turkeys and chickens by transformed T cell line produced lymphokines. *Avian Diseases* 42(3): 545-553.
5. Genovese LL, Lowry VK, Genovese KJ, Kogut MH (2000) Longevity of augmented phagocytic activity of heterophils in neonatal chickens following administration of *Salmonella enteritidis*-immune lymphokines to chickens. *Avian Pathol* 29: 117-122.
6. Wells LL, Lowry VK, DeLoach JR, Kogut, MH (1998) Age-dependent phagocytosis and bactericidal activities of the chicken heterophil. *Dev Comp Immunol* 22(1): 103-109.
7. Lowry VK, Genovese KJ, Bowden LL, Kogut, MH (1997) Ontogeny of the phagocytic and bactericidal activities of turkey heterophils and their potentiation by *Salmonella enteritidis*-immune lymphokines. *FEMS Immunol. Med Microbiol* 19(1): 95-100.
8. Davison TF, Rowell JG, Rea J (1983) Effects of dietary corticosterone on peripheral blood lymphocyte and granulocyte populations in immature domestic fowl. *Res Vet Sci* 34(2): 236-239.
9. Cotter PF (2014) An examination of the utility of heterophil-lymphocyte ratios in assessing stress of caged hens. *Poult Sci* 94(3): 512-517.
10. Davis GS, Anderson KE, Carroll AS (2000) The effects of long-term caging and molt of Single Comb White Leghorn hens on heterophil to lymphocyte ratios, corticosterone and thyroid hormones. *Poult Sci* 79(4): 514-518.
11. Swaggerty CL, Kogut MH, Ferro PJ, Rothwell L, Pevzner IY, et al. (2004) Differential cytokine mRNA expression in heterophils isolated from *Salmonella*-resistant and susceptible chickens. *Immunol* 113(3): 139-148.
12. Swaggerty CL, Ferro PJ, Pevzner IY, Kogut MH (2005) Heterophils are associated with resistance to systemic *Salmonella enteritidis* infections in genetically distinct chicken lines. *FEMS Immunol Med Microbiol* 43(2): 149-154.
13. Chuammitri P, Ostojić J, Andreasen CB, Redmond SB, Lamont SJ, et al. (2009) Chicken heterophil extracellular traps (HETs): Novel defense mechanism of chicken heterophils. *Vet Immunol Immunopathol* 129(1-2): 126-131.
14. Daimon T, Caxton-Martins A (1977) Electron microscopic and enzyme cytochemical studies on granules of mature chicken granular leucocytes. *J Anat* 123: 553-562.
15. Kogut MH, Rothwell L, Kaiser P (2003) Differential regulation of cytokine gene expression by avian heterophils during receptor-mediated phagocytosis of opsonized and non-opsonized *Salmonella enteritidis*. *J Interferon Cytokine Res* 23(6): 319-327.
16. Stabler JG, McCormick TW, Powell KC, Kogut MH (1994) Avian heterophils and monocytes: phagocytic and bactericidal activities against *Salmonella enteritidis*. *Vet Microbiol* 38(4): 293-305.
17. Kogut MH (2002) Dynamics of a protective avian inflammatory response: the role of an IL-8-like cytokine in the recruitment of heterophils to the site of organ invasion by *Salmonella enteritidis*. *Comp Immunol Microb Infect Dis* 25(3): 159-172.
18. Borregaard N, Sorensen OE, Theilgaard Monch K (2007) Neutrophil granules: a library of innate immunity proteins. *Trends Immunol* 28(8): 340-345.
19. He H, Lowry VK, Ferro PM, Kogut MH (2005) CpG-oligodeoxynucleotide-stimulated chicken heterophil degranulation is serum cofactor and cell surface receptor dependent. *Dev Comp Immunol* 29(3): 255-264.
20. Brockus CW, Jackwood MW, Harmon BG (1998) Characterization of beta-defensin prepropeptide mRNA from chicken and turkey bone marrow. *Anim Genet* 29(4): 283-289.
21. Maxwell MH, Robertson GW (1998) The avian heterophil leucocyte: a review. *World's Poultry Sci J* 54(2): 155-178.
22. Van Dijk A, Veldhuizen EJ, Haagsman HP (2008) Avian defensins. *Vet Immunol Immunopathol* 124(1-2): 1-18.
23. Lam KM (1997) Myeloperoxidase activity in chicken heterophils and adherent cells. *Vet Immun and Immunopathol* 57(3-4): 327-335.
24. Redmond SB, Chuammitri P, Andreasen CB, Palic D, Lamont SJ (2011) Genetic control of chicken

- heterophil function in advanced inter cross lines: associations with novel and with known *Salmonella* resistance loci and a likely mechanism for cell death in extracellular trap production. *Immunogenetics* 63(7): 449-458.
25. Chuammitri P, Redmond SB, Kimura K, Andreasen CB, Lamont, et al. (2011) Heterophil functional responses to dietary immunomodulators vary in genetically distinct chicken lines. *Vet Immunol Immunopathol* 142(3-4): 219-227.
 26. Redmond SB, Chuammitri P, Andreasen CB, Palić D, Lamont SJ (2009) Chicken heterophils from commercially selected and non-selected genetic lines express cytokines differently after *in vitro* exposure to *Salmonella enteritidis*. *Vet Immunol Immunopathol* 132(2-4): 129-134.
 27. Farnell MB, Crippen TL, He H, Swaggerty CL, Kogut MH (2003) Oxidative burst mediated by toll like receptors (TLR) and CD14 on avian heterophils stimulated with bacterial toll agonists. *Dev Comp Immunol* 27(5): 423-429.
 28. Mellata M, Dho Moulin M, Dozois CM, Curtiss R, Lehoux B, et al. (2003) Role of Avian Pathogenic *Escherichia coli* Virulence Factors in Bacterial Interaction with Chicken Heterophils and Macrophages. *Infect Immun* 71(1): 494-503.
 29. He H, MacKinnon KM, Genovese KJ, Nerren JR, Swaggerty CL, et al. (2009) Chicken scavenger receptors and their ligand-induced cellular immune responses. *Mol Immunol* 46(11-12): 2218-2225.
 30. Genovese KJ, He H, Swaggerty CL, Kogut MH (2013) The avian heterophil. *Dev Comp Immunol* 41(3): 334-340.
 31. Kogut MH, Iqbal M, He H, Philbin V, Kaiser P, et al. (2005) Expression and function of Toll-like receptors in chicken heterophils. *Dev Comp Immunol* 29(9): 791-807.
 32. Kogut M H, Swaggerty CL, He H, Pevzner I, Kaiser P (2006) Toll-like receptor agonists stimulate differential functional activation and cytokine and chemokine gene expression in heterophils isolated from chickens with differential innate responses. *Microbes Infect* 8(7): 1866-1874.
 33. Lam KM, Kabbur MB, Eiserich JP (1996) Newcastle disease virus-induced functional impairments and biochemical changes in chicken heterophils. *Vet Immunol Immunopathol* 53(3-5): 313-327.
 34. van Dijk A, Tersteeg Zijderveld MH, Tjeerdsma van Bokhoven JL, Jansman AJ, Veldhuizen EJ, et al. (2009) Chicken heterophils are recruited to the site of *Salmonella* infection and release antibacterial mature Cathelicidin-2 upon stimulation with LPS. *Mol Immunol* 46(7): 1517-1526.
 35. Schneider V, Coorens M, Tjeerdsma van Bokhoven J, Posthuma G, van Dijk A, et al. (2017) Imaging the Anti staphylococcal Activity of CATH-2: Mechanism of Attack and Regulation of Inflammatory Response. *M Sphere* 2(6): e00370-17.
 36. Swaggerty CL, He H, Genovese KJ, Pevzner IY, Kogut M H (2011) Protein tyrosine kinase and mitogen-activated protein kinase signalling pathways contribute to differences in heterophil-mediated innate immune responsiveness between two lines of broilers. *Avian Pathol* 40(3): 289-297.

