

Role of Bacterial Integrons in Development of Multidrug-Resistant Phenotypes

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Abstract

Antibiotics have revolutionized medicine in the 20th century and have been extensively used in the treatment and control of many types of infections in a wide variety of plant and animal species. However, the widespread use of antibiotics has led to a massive explosion of antibiotic-resistant phenotypes in human and animal pathogens. Multidrug resistance has been on the rise with some strains becoming resistant to most of the available chemotherapeutic agents. Integrons are ancient, gene acquisition systems commonly found in several bacterial species that allow capture and expression of exogenous genes. Integrons have been recognized as the primary source of resistance genes, and are known to aid in the spread of antimicrobial resistance genes and the rapid evolution of resistance within microbial populations.

Keywords: Integrons; Gene cassettes; Multidrug resistance; Transposable elements

Introduction

Antibiotics enabled the development of modern medicine over the past century by saving numerous lives and are one of the primary candidates in chemotherapy. Although Alexander Fleming is credited with the discovery of penicillin, penicillin as a drug was developed later by the efforst of Norman Heatley, Ernst Chain and Howard Florey. The German biochemist Gerhard Domagk discovered and developed the first sulfonamide popularly known by its trade name of Prontosil [1]. However, the use of antibiotics to treat human infections started with sulfonamides and followed by the amino glycoside streptomvcin and streptothricin. The sequential development of antibacterial drugs is provided in Figure 1. Antibiotics have revolutionized medicine in the 20^{th} century and have been extensively used in the treatment and control of many types of infections in a wide variety of plant and animal species [2]. Various antibiotics like tetracyclines, imidazoles, lincosamides as well as monoclonal antibodies held commanding share in global market and are anticipated to grow at a lucrative growth rate over the forecast period both in rich and poor nations alike. The antibiotics market generated sales of US\$42 billion in 2009 that represented 46% of sales of antiinfective agents and 5% of the global pharmaceutical market. Figure 2 depicts the sales of diverse classes of antibiotics [3]. In 2010, around 150 antibiotic molecules were in preclinical developmental phase, but only 17 in Phase II and 7 in Phase III trials. A recent report suggests that the antibiotics market is likely to reach US\$57 billion by the year 2024.

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Development of Antibiotic Resistance

Initially the antibiotics were extremely successful in clearing pathogenic bacteria that led to a belief that diseases caused by microorganisms would be eventually eliminated. This prompted large-scale production and use of antibiotics in diverse fields like clinical and veterinary medicine, horticulture, agriculture and aquaculture [4-6]. However, the widespread use of antibiotics has led to a massive explosion of antibiotic-resistant phenotypes in human and animal pathogens [7,8]. According to WHO, antimalarial drug resistance is the ability of a strain to survive and/or multiply despite administration and absorption of a drug given in doses equal to or higher than those usually recommended, but within tolerance limit of the patient. Antibiotic resistance is believed to develop de novo from spontaneous mutations or due to development of antibiotic resistance genes [9]. Antibiotic resistance can be acquired through horizontal gene transfer (HGT) by sharing of antibiotic resistance genes via. plasmids, bacteriophages, genomic islands (GIs), integrative and conjugative elements (ICEs), insertion sequences (ISs), transposons and miniature inverted repeat transposable elements (MITEs) (Figure 3) [7, 10-12]. This transfer of antibacterial resistance can even occur between distantly related bacterial species. Recently, integrons, the natural genetic engineering platforms, are being researched for their role in spread of antibacterial resistance [13].



Integrons

Integrons are ancient, gene acquisition systems commonly found in bacterial genomes that allow capture and expression of exogenous genes [14]. Integrons occur in almost all environments, and have the capability to move between species and lineages over evolutionary time frames. These also have access to a vast pool of novel genes whose functions are in the process of being determined. Integrons have been associated with the evolution of bacterial genomes for millions of years and are present in the chromosomes of approximately 17% of the bacterial species whose genomes have been sequenced [15,16]. Integrons are found in a wide range of environments that includes soils, riverine sediment, marine sediment, deep-sea sediment, plant surfaces, aquatic biofilms, and hot springs [2,17-19].

Integrons consist of an integrase (intl) gene that encodes a tyrosine recombinase; an attachment site recognized by the integrin integrase for acquisition of gene cassettes (attl) and the promoter (Pc) which is necessary for efficient transcription and expression of gene cassettes present in the integron [20]. The integron also contains an array of gene cassettes that is highly variable in number and composition [21]. Integron integrases, a distinct group of tyrosine (Y)-recombinases closely related to XerCDrecombinases, carry out the recombination process between attl and the genecassette-borne attachment site (attC), or between two attC sites [22-24]. Have described more than 100 integrase genes in bacteria.

Gene cassettes are non-replicative mobile elements that comprises of an open reading frame (ORF) bounded by a cassette-associated recombination site, originally called a 59-base element but now referred to as *attC* [14, 25].

Integrons have been Divided into Two Maingroups:

(i) Mobile integrons- These are non-self-transposable elements located on mobile genetic elements such as transposons and plasmids, and have cassettes encoding for antibiotic resistance and different *attC* sites [25,26]. Mobile integrons are also sometimes also known as 'resistant integrons' or 'multidrug resistance integrons.

(ii) Superintegrons- These were known to have homogenous *attC* sites and can carry upto 200 cassettes. On the basis of the differences and divergence in the sequences of intl, integrons have been divided into 4 types. Class I integrons are most common, ubiquitous and found in approximately 9% of the sequenced bacterial genomes [27]. Class 1 integrons reside on mobile elements and capture gene cassettes from a vast pool of resistance genes and therefore have a pivotal role in the dissemination of antibiotic resistance [28,29]. This class has been reported in both Gram positive and Gram negative bacteria like Acinetobacter, Aerococcus, Aeromonas, Alcaligenes, Brevibacterium, Burkholderia, Campylobacter, Citrobacter, Corynebacterium, Enterobacter, Enterococcus, Escherichia, Klebsiella, Mycobacterium, Pseudomonas, Salmonella, Serratia, Stenotrophomonas, Shigella. Staphylococcus, Streptococcus and Vibrio. Class 2 integrons are found embedded in the Tn7 transposon family (Tn7 and its derivatives, such as Tn1825, Tn1826 and Tn4132), carrying both the promoter Pc and the recombination site attI2 [30]. This class has been reported in Escherichia coli, Shigellaflexneri, Pseudomonas aeruginosa, Acinetobacter baumannii. Proteus vulgaris. Proteus mirabilis and

Salmonella [2,31-33]. Class 3 integrons are structurally similar to class 2 integrons and have been reported in Klebsiella pneumoniae, Acinetobacter spp., Alcaligenes, Citrobacter freundii, Escherichia coli, Pseudomonas aeruginosa, Pseudomonas putida and Salmonella spp. after their initial discovery in Serratiamarcescensin 1993 [34,35].

Class 4 integrons, first reported in Vibrio sp. and later discovered in microbes like Vibrionaceae, Shewanella, Xanthomonas, Pseudomonad, and other proteobacteria, are believed to be in existence prior to the antibiotic era. Class 4 integrons harbor a large array of gene cassettes encoding adaptations with extension beyond antibiotic resistance and pathogenicity, and have been found to carry cassettes that impart resistance against chloramphenicol and fosfomycin [27].

Multidrug Resistance and Integrons

Ever since the discovery of penicillin in 1928, antibiotics have been extensively used in antibiotic therapy due to which many pathogenic strains have antibiotics become resistant to many and chemotherapeutic agents, the phenomenon termed as multidrug resistance [36]. Several gram-negative bacteria have become resistant to essentially all of the available agents [37]. Multidrug resistance has been on the rise with some strains becoming resistant to most of the available chemotherapeutic agents. For example, methicillin-resistant Staphylococcus aureus (MRSA) which is a major source of hospital-acquired infections is resistant not only to methicillin but also to a range of antibiotics like aminoglycosides, macrolides, tetracycline, chloramphenicol, and lincosamides [36,38]. Integrons are thought to play an important role in the development of multidrug-resistant (MDR) bacteria and spread of antibiotic resistance through horizontal transfer of integrons via mobile elements [39,40]. Integrons have been recognized as the primary source of resistance genes, and are known to aid in the spread of antimicrobial resistance genes and the rapid evolution of resistance within microbial populations [27]. Integrons can harbor more than 100 different antibiotic resistance gene cassettes, which encode adaptations that extend beyond antibiotic resistance and pathogenicity. Another interesting aspect is that the association of multi-drug resistance with integrons enhances the possibility of coselection and persistence of other resistance determinants under the selective pressure due to the use of antimicrobial agents [41]. Table 1 shows the association of diverse classes of integrons in multidrug resistance in bacteria.

Bacteria	Integron class	Reference
Acinetobacter baumannii	I, II	Martins et al. [42]
Escherichia coli	I, II, III	Kargar et al. [39]
Helicobacter pylori	II	Goudarzi et al. [43]
Klebsiellapneumoniae	I, II	Hou et al. [44]
Mycobacterium tuberculosis	Ι	Nazir et al. [45]
Providenciavermicola	Ι	Rajpara et al. [46]
Shigellaflexneri	I, II	Yang et al. [47]
Salmonella enterica	Ι	Ribeiro et al. [48]
Vibrio cholerae	Ι	Jain et al. [49]; Rajpara et al. [46]

Table 1: Role of integrons in development of antibiotic resistance in different bacteria

Conclusion

The use of antimicrobial agents as therapeutic agents in antibacterial therapy has led to rapid emergence of bacterial resistance, especially multiple antibiotic resistances. The role of integrons in the horizontal transfer of antibiotic resistance is increasingly being recognized since the last couple of decades. The future research should address issues like exploring the mechanisms underpinning the recombination process, formation of new gene cassettes and the dynamics of gene-cassette exchange in complex bacterial populations.

References

- 1. Lyddiard D, Jones GL, Greatrex BW (2016) Keeping it simple: lessons from the golden era of antibiotic discovery. FEMS Microbiol Lett 363(8): fnw084.
- 2. Bhargava A, Srivastava S (2017) Biotechnology: Recent Trends and Emerging Dimensions. CRC Press.
- 3. Hamad B (2010) The antibiotics market. Nature Rev Drug Dis 9(9): 675-676.
- 4. Kapil A (2005) The challenge to antibiotic resistance: need to contemplate. Ind J Med Res 121(2): 83-91.
- Khan SJ, Roser DJ, Davies CM, Peters GM, Stuetz RM, et al. (2008) Chemical contaminants in feedlot wastes: Concentrations, effects and attenuation. Environ Int 34(6): 839-859.

- 6. Meek RW, Vyas H, Piddock LJV (2015) Nonmedical uses of antibiotics: time to restrict their use? PLoS Biol 13(10): e1002266.
- 7. Levy SB, Marshall B (2004) Antibacterial resistance worldwide: causes, challenges and responses. Nature Med 10(12): S122-129.
- 8. Gilchrist MJ, Greko C, Wallinga DB, Beran GW, Riley DG, et al. (2007) The potential role of concentrated animal feeding operations in infectious disease epidemics and antibiotic resistance. Environ Health Perspect 115(2): 313-316.
- 9. Read AF, Woods RJ (2014) Antibiotic resistance management. Evol Med Public Health 2014(1): 147.
- 10. Gullberg E, Cao S, Berg OG, Ilback C, Sandegren L, et al. (2011) Selection of resistant bacteria at very low antibiotic concentrations. PLoS Pathog 7: e1002158.
- 11. Stokes HW, Gillings MR (2011) Gene flow, mobile genetic elements and the recruitment of antibiotic resistance genes into Gram-negative pathogens. FEMS Microbiol Rev 35(5): 790-819.
- 12. Ventola CL (2015) Theantibiotic resistance crisis: Part 1: causes and threats. Pharm Therap 40(4): 277-283.
- Mazel D (2004) Integrons and the origin of antibiotic resistance gene cassettes. ASM News 70(11): 520-525.
- 14. Gillings MR (2014) Integrons: past, present, and future. Microbiol Mol Biol Rev 78(2): 257-277.
- 15. Rowe-Magnus DA, Guerout AM, Mazel D (2002) Bacterial resistance evolution by recruitment of super-integron gene cassettes. Mol Microbiol 43(6): 1657-1669.
- 16. Cambray G, Guerout AM, Mazel D (2010) Integrons. Annu Rev Genet 44: 141-166.
- 17. Elsaied H, Stokes H, Nakamura T, Kitamura K, Fuse H, et al. (2007) Novel and diverse integron integrase genes and integron-like gene cassettes are prevalent in deep-sea hydrothermal vents. Environ Microbiol 9(9): 2298-2312.
- 18. Gillings MR, Holley MP, Stokes H, Holmes AJ (2005) Integrons in Xanthomonas: a source of species

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genome diversity. Proc Natl Acad Sci (USA) 102(12): 4419-4424.

- 19. Gillings MR, Holley MP, Stokes HW (2009) Evidence for dynamic exchange of qac gene cassettes between class 1 integrons and other integrons in freshwater biofilms. FEMS Microbiol Lett 296(2): 282-288.
- 20. Mazel D (2006) Integrons: agents of bacterial evolution. Nature Rev Microbiol 4(8): 608-620.
- 21. Engelstädter J, Harms K, Johnsen PJ (2016) The evolutionary dynamics of integrons in changing environments. ISME J 10(6): 1296-1307.
- 22. Collis CM, Hall RM (1992) Site-specific deletion and rearrangement of integron insert genes catalyzed by the integron DNA integrase. J Bacteriol 174(5): 1574-1585.
- Collis CM, Recchia GD, Kim MJ, Stokes HW, Hall RM (2001) Efficiency of recombination reactions catalyzed by class 1 integron integrase IntI1. J Bacteriol 183(8): 2535-2542.
- 24. Boucher Y, Labbate M, Koenig JE, Stokes HW (2007) Integrons: mobilizable platforms that promote genetic diversity in bacteria. Trends Microbiol 15(7): 301-309.
- 25. Stalder T, Barraud O, Casellas M, Dagot C, Ploy MC (2012) Integron involvement in environmental spread of antibiotic resistance. Frontiers Microbiol 3: 119.
- 26. Naas T, Mikami Y, Imai T, Poirel L, Nordmann P (2001) Characterization of In53, a class 1 plasmidand composite transposon-located integron of Escherichia coli which carries an unusual array of gene cassettes. J Bacteriol 183(1): 235-249.
- 27. Deng Y, Bao X, Ji L, Chen L, Liu J, et al. (2015) Resistance integrons: class 1, 2 and 3 integrons. Ann Clin Microbiol Antimicrob 14: 45.
- 28. Hall RM, Collis CM (1998) Antibiotic resistance in gram-negative bacteria: the role of gene cassettes and integrons. Drug Resistance Updates 1(2): 109-119.
- 29. Gillings MR, Paulsen IT, Tetu SG (2017) Genomics and the evolution of antibiotic resistance. Ann N Y Acad Sci 1388(1): 92-107.

- 30. Radstrom P, Skold O, Swedberg G, Flensburg J, Roy PH, et al. (1994) Transposon Tn5090 of plasmid R751, which carries an integron, is related to Tn7, Mu, and the retroelements. J Bacteriol 176(11): 3257-3268.
- 31. Hansson K, Sundstrom L, Pelletier A, Roy PH (2002) Intl2 integron integrase in Tn7. J Bacteriol 184(6): 1712-1721.
- 32. McIver CJ, White PA, Jones LA, Karagiannis T, Harkness J, et al. (2002) Epidemic strains of Shigella sonnei biotype g carrying integrons. J Clin Microbiol 40(4): 1538-1540.
- 33. Miko A, Pries K, Schroeter A, Helmuth R (2003) Multiple-drug resistance in d-Tartrate-positive Salmonella enterica serovar paratyphi B isolates from poultry is mediated by class 2 integrons inserted into the bacterial chromosome. Antimicrob Agents Chemother 47(11): 3640-3643.
- 34. Jones-Dias D, Manageiro V, Ferreira E, Barreiro P, Vieira L, et al. (2016) Architecture of class 1, 2, and 3 integrons from Gram negative bacteria recovered among fruits and vegetables. Front Microbiol 7: 1400.
- 35. Simo Tchuinte PL, Stalder T, Venditti S, Ngandjio A, Dagot C, et al. (2016) Characterisation of class 3 integrons with oxacillinase gene cassettes in hospital sewage and sludge samples from France and Luxembourg. Int J Antimicrob Agents 48(4): 431-434.
- 36. Nikaido H (2009) Multidrug resistance in bacteria. Annu Rev Biochem 78: 119-146.
- 37. Livermore DM (2004) The need for new antibiotics. Clin Microbiol Infect 10(4): 1-9.
- 38. Jo A, Ahn J (2016) Phenotypic and genotypic characterisation of multiple antibiotic-resistant Staphylococcus aureus exposed to subinhibitory levels of oxacillin and levofloxacin. BMC Microbiol 16: 170.
- 39. Kargar M, Mohammadalipour Z, Doosti A, Lorzadeh S, Japoni-Nejad A (2014) High Prevalence of Class 1 to 3 integrons among multidrug-resistant diarrheagenic Escherichia coli in Southwest of Iran. Osong Public Health Res Perspec 5(4): 193-198.
- 40. Hajiahmadi F, Ghale ES, Alikhani MY, Mordadi A, Arabestani MR (2017) Detection of Integrons and Staphylococcal cassette chromosome mec types in

clinical methicillin-resistant coagulase negative Staphylococci strains. Osong Pub Health Res Perspec 8(1): 47-53.

- 41. Deng Y, Wu Y, Jiang L, Tan A, Zhang R, et al. (2016) Multi-drug resistance mediated by Class 1 Integrons in Aeromonas isolated from farmed freshwater animals. Front Microbiol 7: 935.
- 42. Martins N, Picão RC, Adams-Sapper S, Riley LW, Moreira BM (2015) Association of class 1 and 2 integrons with multidrug-resistant Acinetobacter baumannii international clones and Acinetobacter nosocomialis isolates. Antimicrob Agents Chemother 59(1): 698-701.
- 43. Goudarzi M, Heidary M, Azad M, Fazeli M, Goudarzi H (2016) Evaluation of antimicrobial susceptibility and integron carriage in Helicobacter pylori isolates from patients. Gastroenterology and Hepatology Bed Bench 9(1): S47-S52.
- 44. Hou XH, Song XY, Ma XB, Zhang SY, Zhang JQ (2015) Molecular characterization of multidrug-resistant Klebsiella pneumoniae isolates. Braz J Microbiol 46(3): 759-768.
- 45. Nazir T, Abraham S, Islam A (2012) Emergence of potential superbug Mycobacterium tuberculosis,

lessons from New Delhi mutant-1 bacterial strains. Intern J Health Sci 6(1): 87-94.

- 46. Rajpara N, Kutar BMRNS, Sinha R, Nag D, Koley H, et al. (2015) Role of integrons, plasmids and SXT elements in multidrug resistance of Vibrio cholerae and Providencia vermicola obtained from a clinical isolate of diarrhea. Frontiers Microbiol 6: 57.
- 47. Yang C, Li P, Zhang X, Ma Q, Cui X, et al. (2016) Molecular characterization and analysis of high-level multidrug-resistance of Shigella flexneri serotype 4s strains from China. Scientific Rep 6: 29124.
- 48. Ribeiro VB, Lincopan N, Landgraf M, Franco BDGM, Destro MT (2011) Characterization of class 1 integrons and antibiotic resistance genes in multidrug-resistant Salmonella enterica isolates from foodstuff and related sources. Braz J Microbiol 42(2): 685-692.
- 49. Jain M, Kumar P, Goel AK, Kamboj DV, Singh L (2008) Class 1 integrons and SXT elements conferring multidrug resistance in Vibrio cholerae O1 strains associated with a recent large cholera outbreak in Orissa, Eastern India. Int J Antimicrob Agents 32(5): 459-460.