



Gas-Phase Isotopic Immunometry based on Reflectron-Assisted *in Situ* Detection of Products of Gas Chromatographic Separation of Antibodies and Antigens

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Abstract

The system for gas-phase immunometric measurements with isotopic resolution is proposed. This system based on the reflectron-based Finnigan MAT mass spectrometer can be used in combination with gas chromatograph or gas-liquid chromatographic device. The IMMUNOREFLECTRON-2016 project has been quickly closed for the organizational \ administrative reasons, and all the schemes are now confined in the room where this project has been started. Consequently, it is only possible to propose the variants of similar research and development task solutions in different conditions.

Keywords: Immunometry; Radioimmunology; Gas Chromatography; Gas-Liquid Chromatography; Immunochemistry

Introduction

It is well known that gas chromatography (including gas-liquid chromatography), and particularly, gas chromatography with mass-spectrometric detection, is the classical instrument for antigen / antibody measurements and antibody-assisted experimental immunological investigations. Alternative gas phase detection techniques, such as surface Plasmon resonance spectroscopy on the solid-state antibody-containing carrier or gas-phase electrophoretic molecular mobility analysis are not so useful, because they are very exotic (instrumentally) [1-10]. Despite this fact, the best analytical sensitivity for all chromatographic and precipitation techniques, including gas chromatography, can be realized only in mass-spectrometric detection with isotopic resolution. For example, for accurate or selective measurement of protective antigen in plasma isotope dilution mass spectrometry can be used with preliminary immunocapture or, in different case, precision analysis of some specific antigen isoforms using immunoprecipitation can be realized only in stable isotope labeling mass spectrometry [11,12]. Equivalent resolution is useful for antibody measurements and antibody-assisted investigations. For example, inductively coupled plasma mass

spectrometry technique can be used for the quantification of metal-antibody based drugs using stable isotope tracing [13]. Isotopic approaches for antigen or antibody detection are not only good techniques for immunology, but also they are big challenges for labeling mass spectrometry techniques, for example, the production and application of high quality stable isotope labeled monoclonal antibodies is a powerful tool for mass spectrometry analysis using labeled antibodies, which can be used in experiments, approved by ultrahigh-precision standards, such as SISCAPA (Stable-Isotope Standard and Capture by Anti-Peptide Antibody) [14,15]. Unfortunately, they are not useful in GC-MS techniques, because the LC-MS (particularly, HPLC-MS, UPLC-fluorescence-MS etc.) are more applicable in "liquid" biochemistry [16-18]. Many isotope effects, investigable in gas phase, in aerosol conditions or in gas-liquid / solid-gas interfaces are measured in soft matter or liquid conditions [19,20].

Despite this fact, gas chemistry is not inappropriate in immunochemistry and, consequently, such techniques cannot be interpreted as a "Cinderella of contemporary immunology". Many gas effects in immunology have been studied in the past century, starting from the observation of the effect of mustard gas on antibody formation (this

research trend has been stable before the second part of 1960th) and continued in the last decade of the last century by the gas-phase assisted antibody production investigations [21-25]. “Gas phase immunology” (*sensu lato*) is a potentially fruitful research area, which started from the immunology of archaeobacteria that produce methane gas (on the cellular level) and from the kinetic loss of antibody titer following labeling by tritium gas exposure (on the isotopic / biochemical level and in the “humoural immunity” measurements, assisted by isotopes), possesses a significant applicability potential in clinical medicine – from the colon pathology to the gas gangrene infection agent detection [26-29].

Interface effects in gas chemistry, which can be applied in clinical and / or biochemical immunology, are very interesting. For example, verification of a specific reaction between an airborne antigen and an immobilized antibody at a gas-solid interface is the Holy Grail of the real world immunology, epidemiology and infection agent dissemination (contamination, from the standpoint of infected surface) research [30]. Also it is the Holy Grail of the atmospheric allergology, because gas-exchange following bronchial challenge with antigen in patients with extrinsic-asthma, but methacholine (well-known as a non-selective muscarinic receptor agonist in the parasympathetic nervous system) and antigen challenge upon gas exchange in allergic subject are different by mechanisms [31,32]. Consequently, it is necessary to provide a multiplexed technique for estimation of mast cell activation assessed by antigen challenge in asthmatics. It is obvious that the most sensitive approach for enzymatic assays of plasma histamines is the double-isotope technique [33]. From the above considerations one should think about the isotopic measurement techniques for two, three or more antigens or antibodies (and, consequently, about techniques, which can be realized using two, three {or more} isotopes).

Many authors of old classical articles (since 1970th) use double-antibody triple isotope RIAs, double isotope methods for the determination of antibody affinities and antigen trapping mechanism investigations [34-39]. Double-isotope techniques are actual until now, because dual isotope 3D QIQA or “DICIQA” (Dual-Isotope CryoImaging Quantitative Autoradiography) technique are useful in affimetry and qualimetry of antibody–drug conjugate distributions and payload delivery through imaging, just like the simple dual-antibody imaging (not only in static, but also in kinetic and spatiotemporal dynamic regimes [40-43]). But it is the *radioisotope* technique pull (excluding some tomography techniques, but not all [44,45]). However, it is impossible to use such techniques in clinical conditions in XXI century. Moreover, in 1970th such techniques have been preliminary approbated in veterinary but not all of them have been introduced in practice. It was not only industrial inapplicability problem, but also it was the biggest physico-chemical problem of radioimmunology: besides the antigen competition (which can be analyzed also using stable isotope labeling) and the isotope selectivity problem for antigen response monitoring or analysis there is a problem of biological fractioning of isotopes and soft matter sorption of them. Similar effects can be observed in different branches of organic chemistry, colloid chemistry and polymer chemistry – for example, kinetic isotope effects in the decarboxylation of 5-nitro-3-carboxybenzisoxazole (which can be catalyzed by antibodies and other biomimetic reactions. All described processes can be realized using non-radioactive, stable isotopes. Consequently, there is a strong need in the appropriate methods for stable isotopic measurements in chemical immunology (but not radioimmunology \ RIA) and immunometry (a term from the old articles, for example institutionalized in the second part of 2010th by Cold Spring Harbor Lab [46-54]).

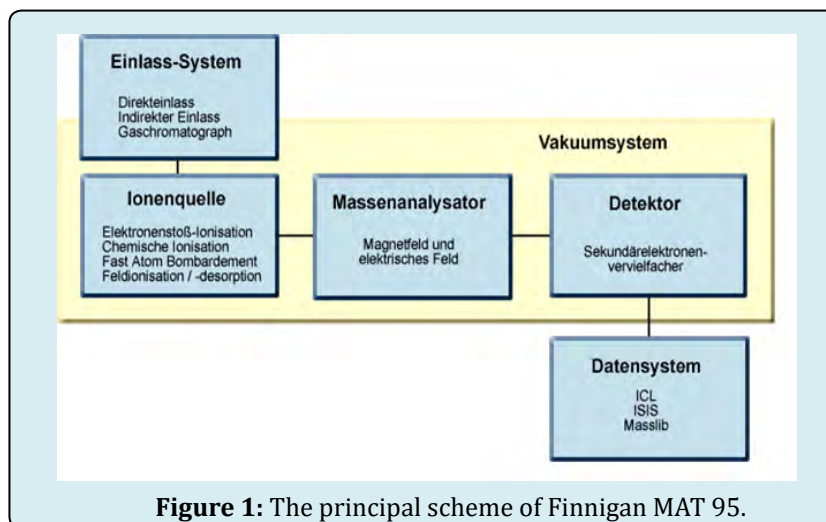


Figure 1: The principal scheme of Finnigan MAT 95.

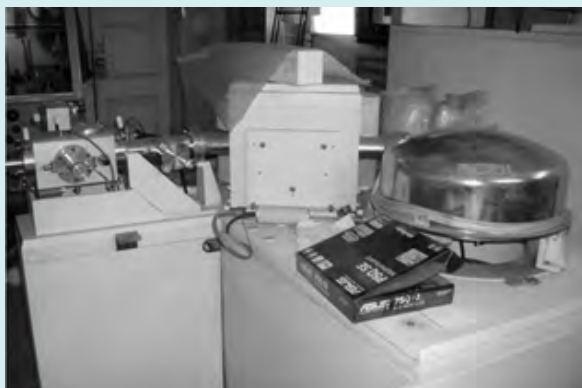
It is well known that the best technique for stable isotope analysis is isotopic mass-spectrometry. This instrument is optimal for gas phase and ultra-low concentrations and volumes of gas analytes (for example, for measurements of gas-filled microbubble-mediated delivery of antigen and the induction of immune responses gas vesicle nanoparticle utilization for antigen displaying and observations of the phagocytosis of gas-filled microbubbles by human and murine antigen-presenting cells [55-59]). The project of such instrumentation, applicable for immunometry, is presented in this article.

Technical Implementation

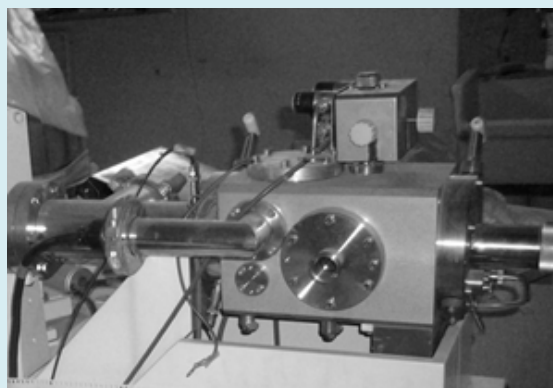
The double focusing sector type instrument with reversed Nier-Johnson geometry Finnigan MAT 90 / 95 has been used as a general part of our setup (general scheme of this instrument is given in Fig. 1). Such devices have been used for the carbon geochemistry aims earlier [60-62]. They can also be used as instruments for gas-phase analysis in combination (tandem) with gas chromatographic devices or as the gas isotope mass spectrometer setup itself [63,64]. Finnigan MAT HDO-Euiliberator for H₂O / gas phase equilibration in hydrogen and oxygen analysis has been developed in 1990th [65]. Many modifications / upgrade options since 1980th have been implemented in some configurations of this instrument family starting from the earliest MAT models. Many papers and technical documents have been published by Finnigan engineers / constructors and related firms (for example, the most cited papers and Figure 5). The MAT family exemplars have been

intensively used in Chinese works since 1980th [66-85]. It is possible to measure not only stable isotopes (including heavy ones), but also radioisotopes using MAT systems with special accessories [86,87]. It is a very good instrument until now (for example, Table 1). Such considerations can be interpreted as a prerequisite of applicability of this device in our aims (below).

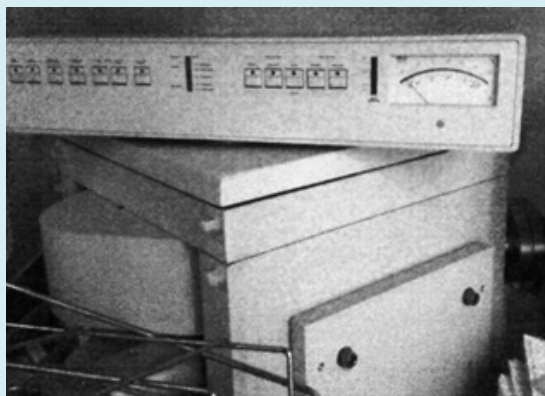
Our general aims were oriented on the data-dependent analysis and transformation of interference of peaks to informative signals using deconvolution and other mathematical algorithms [88-90]. Previously our instrument has been disassembled (Figure 2). The gas chromatography section (established by "Varian 3400" gas chromatograph, which has been supported since 1980th until 2000th was disconnected before inlet withdrawing from the technical hole (Figure 3). Old computer \ data station with old dot matrix printer unit on the special chassis (Figure 4) were changed, because our project was started from the novel data station projecting, which was started on the PXI NI platform with LabVIEW supporting. The target date for the starting of novel MAT modification for immunometry was scheduled on August, 2018, but, unfortunately, the prototype exemplar has been demolished and utilized by the institute stewards \ supply managers in October, 2016. Our project documentation is confined in the room, where this system was installed previously. But it is obvious, that such measurement systems are appropriate and in the nearest future can be implemented in other conditions using alternative technical prototypes [91,92].



A



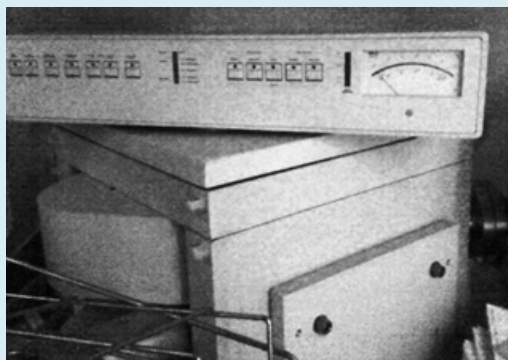
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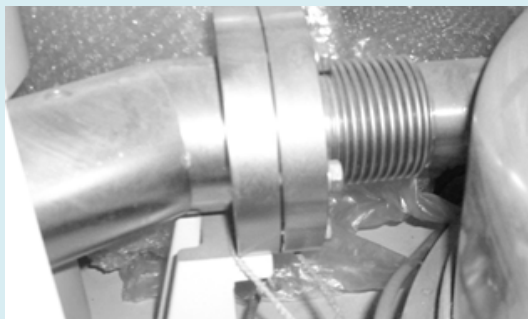
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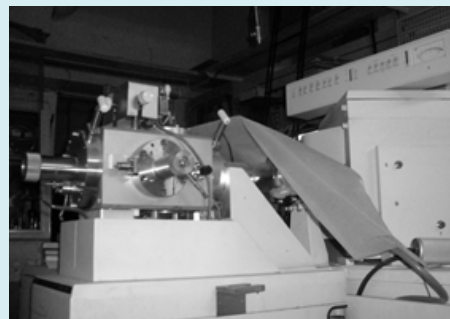
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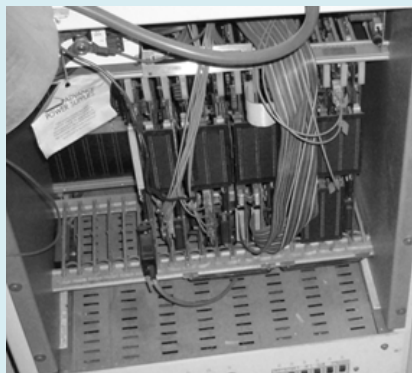
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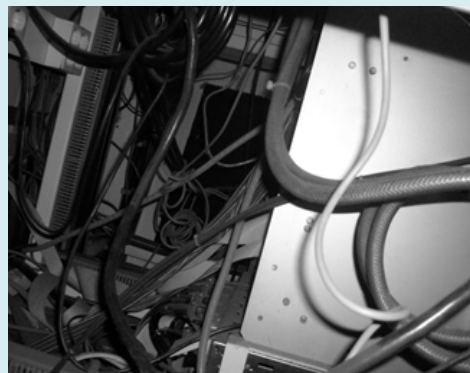
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Figure 2: Disassembled Finnigan MAT scheduled and planned for gas-phase isotopic immunometry aims.

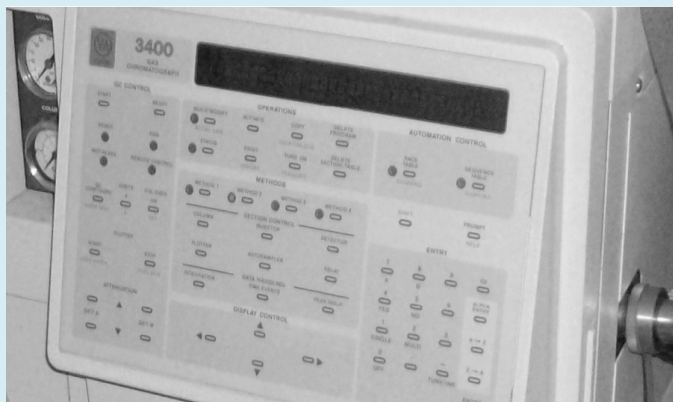


Figure 3: Gas chromatographic part of the above setup.



Figure 4: Disqualified printer and automation cable box.

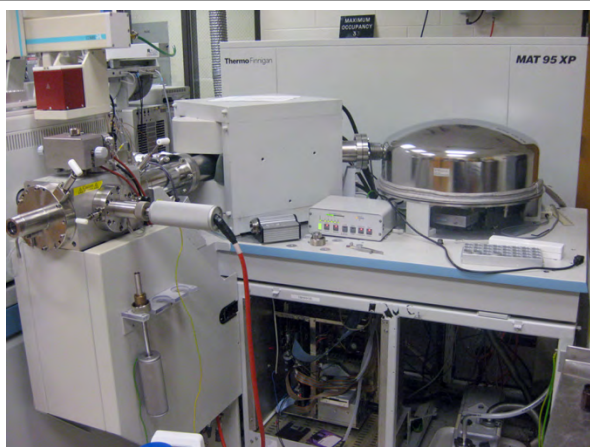


Figure 5: Instruction and documentation of the MAT system.

Photography of MAT system	University or Lab and QR-codes
	<p>Universität Oldenburg Ammerländer Heerstraße 114-118 26129 Oldenburg</p>  <p>Finnigan MAT 95</p> <ul style="list-style-type: none"> • double focussing sector type instrument with reversed Nier-Johnson geometry • EI, CI, FD, FI, FAB, ESI • High resolution and exact mass measurement • GC/MS
	<p>Max-Planck-Institut für Kohlenforschung Double focussing sector field MS. Equipped with ESI, FAB, EI, CI sources. Used for accurate mass measurements.</p> 
	<p>Die Universität Bayreuth Central analytical services Finnigan MAT95 Manufacturer: Thermo Fisher Scientific Analysator: Sector field (double focusing) Mass-Range: 45 – 1000 Da (typical) Resolution: 3000 Ionization-Source: EI (Electron Ionization) Sample-Inlet: GC (Hewlett Packard 5890 Series II)</p> 



Doppelfokussierendes Sektorfeldgerät mit umgekehrter Nier-Johnson-Geometrie (d.h. elektrostatischer Sektor hinter dem magnetischen Sektor)



State University of New York



MAT-95XP
DOUBLE FOCUSING

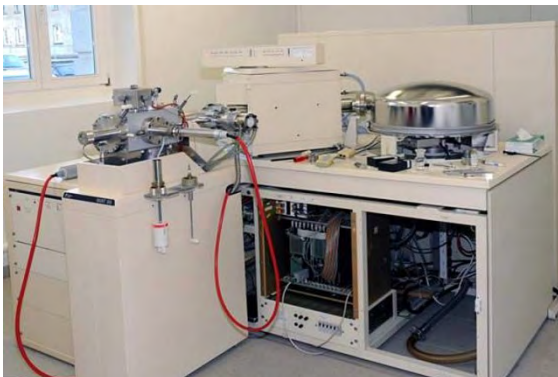


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University of Rostock Faculty of Agriculture and
Environmental Sciences



University of Gent
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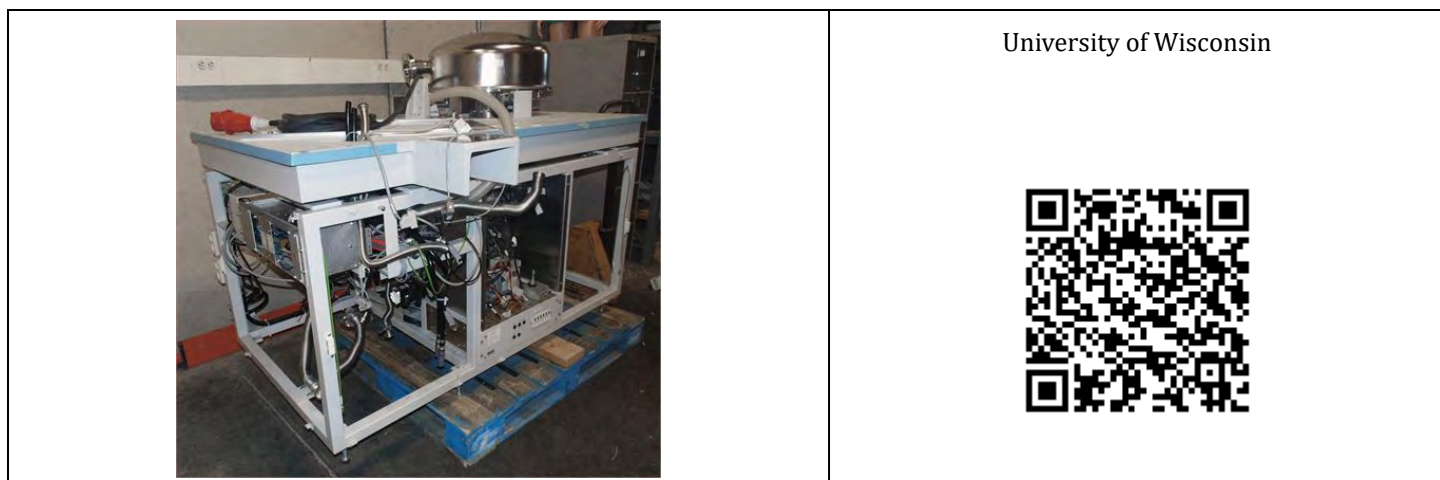


Table 1: Finnigan MAT localization examples in different labs.

Conclusion

Thus, in this brief review it has been shown that:

- Gas phase immunometry or gas phase immunochemistry is a promising research area with the maximum accuracy and informative results provided by gas chromatographic methods with MS detection.
- When using isotope mass-spectrometers, particularly, with Finnigan MAT application, it is possible to develop a new branch of analytical biochemistry – isotopic gas phase immunometry or gas phase isotopic immunochemistry.
- The ubiquity of Finnigan MAT-like systems in research laboratories all over the world makes it possible to continue the above studies and to develop the ideas described.

References

1. Medley CD, Kay J, Li Y, Gruenhagen J, Yehl P, et al. (2014) Quantification of residual solvents in antibody drug conjugates using gas chromatography. *Analytica Chimica Acta* 850: 92-96.
2. Abad A, Manclus JJ, March C, Montoya A (1993) Comparison of a monoclonal antibody-based enzyme-linked immunosorbent assay and gas chromatography for the determination of nicotine in cigarette smoke condensates. *Analytical Chemistry* 65(22): 3227-3231.
3. Nakagawa A, Tanishima Y, Hirota T, Takahagi H, Horiguchi M, et al. (1986) Determination of a carbacyclin derivative in plasma by gas-chromatography mass-spectrometry using cleanup method with immobilized antibody. *Bunseki Kagaku* 35(3): 298-302.
4. Furuya K, Urasawa S (1981) Gas-liquid chromatographic demonstration of the specificity of rabbit IgG antibody to the pesticide DDT and its metabolites. *Molecular Immunology* 18(2): 95-102.
5. Hubbard HL, Eller TD, Mais DE, Halushka PV, Baker R, et al. (1987) Extraction of thromboxane B2 from urine using an immobilized antibody column for subsequent analysis by gas chromatography-mass spectrometry. *Prostaglandins* 33(2): 149-160.
6. Lemm U, Tenczer J, Baudisch H, Krause W (1985) Antibody-mediated extraction of the main tetrahydrocannabinol metabolite, 11-nor- Δ^9 -tetrahydrocannabinol-9-carboxylic acid, from human urine and its identification by gas chromatography-mass spectrometry in the sub-nanogram range. *Journal of Chromatography* 342(2): 393-398.
7. Hansen K, Lau AM, Giles K, McDonnell JM, Struwe WB, et al. (2018) A Mass-Spectrometry-Based Modelling Workflow for Accurate Prediction of IgG Antibody Conformations in the Gas Phase. *Angewandte Chemie International Edition* 57(52): 17194-17199.
8. Bagheri H, Ghambarian M, Salemi A, Es-Haghi A (2009) Trace determination of free formaldehyde in DTP and DT vaccines and diphtheria-tetanus antigen by single drop microextraction and gas chromatography-mass spectrometry. *Journal of Pharmaceutical and Biomedical Analysis* 50(3): 287-292.
9. Bowen J, Noe LJ, Sullivan BP, Morris K, Martin V, et al. (2003) Gas-phase detection of trinitrotoluene utilizing a solid-phase antibody immobilized on a gold film by means of surface plasmon resonance spectroscopy. *Applied Spectroscopy* 57(8): 906-914.
10. Laschober C, Wruss J, Blaas D, Szymanski WW, Allmaier G (2008) Gas-phase electrophoretic molecular mobility

- analysis of size and stoichiometry of complexes of a common cold virus with antibody and soluble receptor molecules. *Analytical Chemistry* 80(6): 2261-2264.
11. Solano MI, Woolfitt AR, Boyer AE, Lins RC, Isbell K, et al. (2019) Accurate and selective quantification of anthrax protective antigen in plasma by immunocapture and isotope dilution mass spectrometry. *Analyst* 144(7): 2264-2274.
 12. Chen YT, Tuan LP, Chen HW, Wei IA, Chou MY, et al. (2014) Quantitative analysis of prostate specific antigen isoforms using immunoprecipitation and stable isotope labeling mass spectrometry. *Analytical Chemistry* 87(1): 545-553.
 13. Ciavardelli D, DAnniballe G, Nano G, Martin F, Federici G, et al. (2007) An inductively coupled plasma mass spectrometry method for the quantification of yttrium-antibody based drugs using stable isotope tracing *Rapid Commun Mass Spectrom* 21(14): 2343-2350.
 14. Amsler P, Wolf T, Lanshoeft C, Bettighofer A, Einfeld J, et al. (2017) Production and application of high quality stable isotope labeled monoclonal antibody for mass spectrometry analysis. *Journal of Labelled Compounds and Radiopharmaceuticals* 60(3): 160-167.
 15. Yang X, Naughton SX, Han Z, He M, Zheng YG, et al. (2018) Mass spectrometric quantitation of tubulin acetylation from pepsin-digested rat brain tissue using a novel stable-isotope standard and capture by anti-peptide antibody (SISCAPA) method. *Analytical Chemistry* 90(3): 2155-2163.
 16. Heudi O, Barteau S, Zimmer D, Schmidt J, Bill K, et al. (2008) Towards absolute quantification of therapeutic monoclonal antibody in serum by LC- MS/MS using isotope-labeled antibody standard and protein cleavage isotope dilution mass spectrometry. *Analytical Chemistry* 80(11): 4200-4207.
 17. Kang P, Tanya M, Wei C, Sheng Z, Eric PS, et al. (2016) Use of a stable isotope labeled reporter peptide and antioxidants for reliable quantification of methionine oxidation in a monoclonal antibody by liquid chromatography/mass spectrometry. *Rapid Communications in Mass Spectrometry* 30(14): 1734-1742.
 18. Martyn S, Iglesias N, Delporte C, Farrell A, Navas Iglesias N (2015) Comparative analysis of monoclonal antibody N-glycosylation using stable isotope labelling and UPLC-fluorescence-MS. *Analyst* 140(5): 1442-1447.
 19. Laissue JA, Stoner RD (1979) Deuterium isotope effects on lymphoid tissues and humoral antibody responses in mice. *Virchows Archiv A* 383(2): 149-166.
 20. Xing S, Cekan SZ (1983) Isotope effects of tritium in a ligand-antibody binding. *The International Journal of Applied Radiation and Isotopes* 34(7): 957-958.
 21. Hektoen L, Corper HJ (1921) The effect of mustard gas (dichlorethylsulphid) on antibody formation. *The Journal of Infectious Diseases* 28(3): 279-285.
 22. Schneider R (1964) Comparative investigations on the effectiveness of some cytostatics and X-ray irradiation. Part 2. Histological and cytological conditions of spleen, antibody determinations after brucella melitensis antigen doses, and bone marrow changes as a result of the action of n-mustard gas-ph osphamide ester, trisethyleneiminobenzoquinone X-radiation alone and in various combinations on. *Strahlentherapie (West Germany)*, pp: 124.
 23. Heidel J (1997) Monoclonal antibody production in gas-permeable tissue culture bags using serumfree media. *Center for Alternatives to Animal Testing: Alternatives in Monoclonal Antibody Production* 8: 18-20.
 24. Lipski LA, Witzleb MP, Reddington GM, Reddington JJ (1998) Evaluation of small to moderate scale in vitro monoclonal antibody production via the use of thei-MAb gas-permeable bag system. *Research in Immunology* 149(6): 547-552.
 25. Stang BV, Wood PA, Reddington JJ, Reddington GM, Heidel JR (1998) Monoclonal antibody production in gas-permeable flexible flasks, using serum-free media. *Journal of the American Association for Laboratory Animal Science* 37(6): 55-60.
 26. de Macario EC, Wolin MJ, Macario AJ (1981) Immunology of archaebacteria that produce methane gas. *Science* 214(4516): 74-75.
 27. Schlegel DE (1964) The Loss of Antibody Titer Following Labeling by Tritium Gas Exposure. *The Journal of Immunology* 93(4): 566-566.
 28. Clark WR, Bernard HR, Gray VC (1969) Gas gangrene: Diagnostic problems and the use of the fluorescent-antibody technique for the study of clostridium perfringens infections. *Archives of Surgery* 99(2): 239-244.
 29. Kagnoff MF (1989) Immunology and disease of the gastrointestinal tract. In: Sleisenger MH, Fordtran JS, (Eds.), *Gastrointestinal Disease: pathophysiology, diagnosis, management* 1: 1005-1019.

30. Iwanaga H, Tsuzuki H, Kamiyama Y, Ueda H (2009) Verification of a specific reaction between an airborne antigen and an immobilized antibody at a gas-solid interface. *Analytical Sciences* 25(9): 1101-1106.
31. Wagner PD, Ramsdell JW, Incaudo GA, Rubinfeld AR, Young IH (1978) Gas-exchange following bronchial challenge with antigen in patients with extrinsic-asthma. *American Review of Respiratory Disease* 117(4): 409-409.
32. Olgiati R, Birch S, Rao A, Wanner A (1981) Differential effects of methacholine and antigen challenge on gas exchange in allergic subjects. *Journal of Allergy and Clinical Immunology* 67(4): 325-329.
33. Brown MJ, Ind PW, Causon R, Lee TH (1982) A novel double-isotope technique for the enzymatic assay of plasma histamine: application to estimation of mast cell activation assessed by antigen challenge in asthmatics. *Journal of Allergy and Clinical Immunology* 69(1): 20-24.
34. Egan ML, Coligan JE, Lautenschleger JT, Todd CW (1972) The triple isotope double antibody assay: Application to the carcinoembryonic antigen. In *Proceedings of the Second Conference on Embryonic and Fetal Antigens 2*: 267-272.
35. Safian R, Smith P, Rosenberg E (1974) Quantitation of Carcinoembryonic Antigen Levels in Normal Individuals Using a Double Antibody Triple Isotope Radioimmunoassay. *Southern Medical Journal* 67(11): 1392.
36. Egan ML, Todd CW, Knight WS (1977) ⁵⁷Co: a volume marker for the triple-isotope, double-antibody radioimmune assay. *Immunochemistry* 14(8): 611-613.
37. Fritsche HA, Tashima CK, Romsdahl MM, Holoye PY, Geitner A (1978) A clinical evaluation of the triple-isotope double-antibody radioimmunoassay for carcinoembryonic antigen. *American Journal of Clinical Pathology* 69(2): 140-146.
38. Gaze S, West NJ, Steward MW (1973) The use of a double isotope method in the determination of antibody affinity. *Journal of Immunological Methods* 3(4): 357-363.
39. Van Rooijen N (1974) Mechanism of follicular antigen trapping: Evidence for a two cell mechanism using double isotope autoradiography. *Immunology* 27(4): 617-622.
40. Ilovich O, Qutaish M, Hesterman JY, Orcutt K, Hoppin J, et al. (2018) Dual-Isotope Cryoimaging Quantitative Autoradiography: Investigating Antibody-Drug Conjugate Distribution and Payload Delivery Through Imaging. *Journal of Nuclear Medicine* 59(9): 1461-1466.
41. Ilovich O (2016) A dual-isotope 3D cryo-imaging quantitative autoradiography (CIQA) method for simultaneous and quantitative assessment of both antibody and drug conjugate tumor distribution and kinetics. *Cancer Research* 76: 615.
42. Knight JC, Mosley MJ, Kersemans V, Dias GM, Allen PD, et al. (2019) Dual-isotope antibody imaging. *Nuclear Medicine and Biology* 70: 14-22.
43. Adams GP, DeNardo SJ, Deshpande SV (1990) Dual isotope pharmacokinetics of an anti-lymphoma antibody (LYM-1) with a new bifunctional chelating agent. *Journal of Nuclear Medicine* 31(823): 23.
44. Johnson LL (1992) Dual isotope thallium-201 and indium-111 antimyosin antibody tomographic imaging to identify viable myocardium at further ischemic risk after myocardial infarction. *Journal of Nuclear Biology and* 36(2): 91-96.
45. Hertel A, (1993) Characterization of brain-tumors using dual-isotope SPECT with thallium, somatostatin-analog, anti-EGF antibody. *Journal of Nuclear Medicine* 34(5): 205.
46. Hoffmann A (1976) Zur Anwendung radioaktiver Isotope bei Antigen-Antikörper-Reaktionen in der Veterinärmedizin (Doctoral dissertation).
47. Potters L, Huang D, Fearn P, Kattan MW (2003) The effect of isotope selection on the prostate-specific antigen response in patients treated with permanent prostate brachytherapy. *Brachytherapy* 2(1): 26-31.
48. Sommer F, Mühlhaus T, Hemme D, Veyel D, Schroda M (2014) Identification and validation of protein-protein interactions by combining co-immunoprecipitation, antigen competition, and stable isotope labeling. *Methods Mol Biol* 1188: 245-261.
49. Lewis C, Paneth P, OLeary MH, Hilvert D (1993) Carbon kinetic isotope effects on the spontaneous and antibody-catalyzed decarboxylation of 5-nitro-3-carboxybenzisoxazole. *Journal of the American Chemical Society* 115(4): 1410-1413.
50. Bonini PA, Banfi G, Pontillo M, Casari E, Murone M (1988) Free-thyroxine estimation by enhanced luminescent immunometry. *Clinical Chemistry* 34(6): 1207.
51. Dorizzi RM, Giavarina D, Schiavon R (1995) The transition from manual organization to random access automation of an immunometry section. *European*

- Journal of Laboratory Medicine 3: 257-261.
52. Kohl TO, Ascoli CA (2017) Immunometric Antibody Sandwich Enzyme-Linked Immunosorbent Assay. Cold Spring Harbor Protocols 2017(6).
 53. Kohl TO, Ascoli CA (2017) Indirect immunometric ELISA. Cold Spring Harbor Protocols 2017(5).
 54. Kohl TO, Ascoli CA (2017) Immunometric Double-Antibody Sandwich Enzyme-Linked Immunosorbent Assay. Cold Spring Harbor Protocols 2017(6).
 55. Bioley G, Lassus A, Bussat P, Terrettaz J, Tranquart F, et al. (2012) Gas-filled microbubble-mediated delivery of antigen and the induction of immune responses. *Biomaterials* 33(25): 5935-5946.
 56. Corthésy B, Bioley G (2017) Gas-filled microbubbles: Novel mucosal antigen-delivery system for induction of anti-pathogen's immune responses in the gut. *Gut Microbes* 8(5): 511-519.
 57. DasSarma S, DasSarma P (2015). Gas vesicle nanoparticles for antigen display. *Vaccines* 3(3): 686-702.
 58. Childs TS, Webley WC (2011) In vitro assessment of chlamydial antigen display, delivery and processing by halobacterial gas vesicles. In ASM General Meeting, New Orleans LA USA Session, pp: 197.
 59. Bioley G, Bussat P, Lassus A, Schneider M, Terrettaz J, et al. (2012) The phagocytosis of gas-filled microbubbles by human and murine antigen-presenting cells. *Biomaterials* 33(1): 333-342.
 60. Reineking A, Langel R, Schikowski J (1993) 15N, 13C-online measurements with an elemental analyser (Carlo Erba, NA 1500), a modified trapping box and a gas isotope mass spectrometer (Finnigan, MAT 251). *Isotopes in Environmental and Health Studies* 29(1-2): 169-174.
 61. Scrimgeour CM, Rennie MJ (1988) Automated measurement of the concentration and 13C enrichment of carbon dioxide in breath and blood samples using the Finnigan MAT breath gas analysis system. *Biomedical Environmental Mass Spectrometry* 15(7): 365-367.
 62. Matsumoto R (1985) Measurements of carbon and oxygen isotopes in carbonate samples with a Finnigan MAT delta E mass spectrometer. Report of Research Results on the 1984 Science Grant of the Ministry of Education Science and Culture, pp: 2-9.
 63. Hiltz JA, Power JJ (1986) Optimization of the Finnigan MAT 5100 Capillary Gas Chromatograph-Mass Spectrometer for the Analysis of Polychlorinated Biphenyls (No. DREA-TM-86/222). Defence Research Establishment Atlantic Dartmouth (Nova Scotia).
 64. Reineking A, Langel R, Schikowski J (1995) Elemental analyser (CARLO ERBA, NA 1500), a modified trapping box and a gas isotope mass spectrometer (Finnigan, MAT 251). *Isotopenpraxis Isotopes in Environmental and Health Studies* 29: 1-2.
 65. Avak H, Brand WA (1995) The Finnigan MAT HDO-Equilibrators: a fully automated H₂O/gas phase equilibration system for hydrogen and oxygen isotope analysis. *Finnigan MAT Application News* 11: 1-13.
 66. HP D (1983) 10n Trap Detector, from Finnigan MAT. *Chromatographia* 17(8).
 67. Loveridge WD (1986) Measurement of biases in the electron multiplier ion detection system of a Finnigan-MAT model 261 mass spectrometer. *International journal of mass spectrometry and ion processes* 74(2-3): 197-206.
 68. Ratnayake WM, Timmins A, Ohshima T, Ackman RG (1986). Mass spectra of fatty acid derivatives, of isopropylidenes of novel glyceryl ethers of cod muscle and of phenolic acetates obtained with the Finnigan Mat Ion Trap Detector. *Lipids* 21(8): 518-524.
 69. Henry Y (1988) Finnigan MAT-recent developments in mass-spectrometry. *Analysis* 16(6): 60-63.
 70. Reber SD, Cordes GT (1995) Modifications to the Finnigan MAT 271 mass spectrometer in the Inorganic Gas Analysis Lab (No. SAND-95-0961). Sandia National Labs, Albuquerque, NM (United States).
 71. Menéndez M, Iburguchi JIG, Cuesta LAO (1998) Análisis isotópico de alta precisión mediante técnicas de evaporación total. Posibilidades analíticas del Finnigan MAT 262 equipado con 8 cajas de Faraday. *Boletín de la Sociedad Española de Mineralogía* 21(1): 170-171.
 72. Manual FIM (1979) Finnigan MAT Corporation (Manual No. 20000-90193), San Jose.
 73. Manual FIM (1986) Finnigan MAT Corporation (Manual No. 20000-90193), San Jose.
 74. Brand WA, Ricci M, Habfast K (1988) Finnigan MAT Delta S/GC Application Data Sheet No. 1. Finnigan MAT, San Jose, CA, USA.
 75. Smith DH, McKown HS, Carter JA (1987) ISPO Task A-143: Final report of an evaluation of the Finnigan-MAT THQ mass spectrometer as an on-site inspection

- instrument (No. ORNL/TM-10627; ISPO-285). Oak Ridge National Lab, TN, USA.
76. Jackett S, Moini M (1994) Conversion of the Finnigan MAT TSQ-70 thermospray ionization interface to an electrospray ionization interface. *Review of Scientific Instruments* 65(3): 591-596.
 77. Tillert PR, Mutton IM, Lane SF (1996) Biofluid Assay for Peptide-related and Other Drugs. *Finnigan MAT, 355 River Oases Parkway* 24: 329.
 78. Chen Y (1986) FINNIGAN MAT launched high-performance desktop gas chromatography / mass spectrometry system. *Foreign scientific instruments* 3: 5.
 79. Zhou R (1986) Finnigan-MAT Magnetic Mass Spectrometer User Collaboration Group Holds the Fifth Edition Software Symposium on Data Systems. *Journal of Mass Spectrometry* 4: 1.
 80. Liu Z, Xiang T (1990) Chromatography of Finnigan Mat 4515 Chromatograph / Mass Spectrometer. *Scientific Instruments Abroad* 2: 42-43.
 81. Tongshou X (1995) Maintenance of Vacuum System in Finnigan Mat 4515 GC/MS [J]. *Analysis and Testing Technology and Instruments* 3.
 82. Weidong S, Zicheng P, Zhaorong W, Xun Z (1997) Fang Jiajun (Finnigan MAT China Inc, Beijing 100081, China) Received 1996-08-06; Application of Negative Thermal Ionization Mass Spectrometry in Os Isotope Determination. *Journal of Chinese Mass Spectrometry Society* 2.
 83. Xiaoliu H, Su Yun (1998) Multiple failures and maintenance of Finnigan Mat 4510 color / mass spectrometer. *Laboratory Research and Exploration* 17(3): 104-105.
 84. Chen Y, Jiang S, Ling H, Pan J, Lai M (2005) Calibration of Carbonate Oxygen Isotope Analysis Using Finnigan MAT-252 Gas Isotope Mass Spectrometer. *Journal of Chinese Mass Spectrometry Society* 26(2): 115-118.
 85. Satish Kumar M, Matsuda J, Yamazaki R, Takano A, Hideki W (2010) A new SF6 inlet system with a modified Faraday collector alignment of Finnigan MAT-251 mass spectrometer for sulfur isotope measurement. *Earth Science Research Report* 37: 41-49.
 86. Ingeneri KB, Riciputi LR, Hedberg PML (2002) Preliminary results of uranium and plutonium efficiency measurements using a high efficiency cavity ion source interfaced with a Finnigan MAT 262 mass spectrometer. *Proceedings of the Institute of Nuclear Materials Management*, pp: 24-27.
 87. Makishima A, Nakamura E, Akimoto S (1991) Investigation of the bias in a secondary electron multiplier of Finnigan-MAT 261 mass spectrometer for the quantitative analysis of rare-earth elements in rock samples. ISEI (Inst. Study Earth's Inter.), Okayama University.
 88. Lammert SA (1987) Data-Dependent Instrument Control on the Finnigan MAT TSQ70. *Technical Report Finnigan MAT 603*: 1-10.
 89. Tittes W, Jakubowski H, Stüwer D (1994) Reduction of some spectral interference in ICP-MS, finnigan MAT elemental mass spectrometry technical and application note 3. *Finnigan MAT GmbH, Bremen*.
 90. Greb U, Rottmann L (1994) Interferenzfreie Elementspuren und Ultraspurenanalyse mit einem hochauflösenden ICP MS. *Finnigan MAT Bericht* 7: 94.
 91. Muchmore CB, Chen JW (1985) Intermediates formed during supercritical desulfurization of coal. Seventh quarterly technical progress report, January 1-March 31, 1985. *Leco Total Sulfur Analyzer and Varian Model 3400 chromatograph* [No. DOE/PC/60797-7]. Southern Illinois Univ., Carbondale (USA). Dept. of Mechanical Engineering and Energy Processes.
 92. Jie W (2000) The Installing Adjusting of Gas Chromatograph Seviess 3400. *Fujian Analysis and Testing* 9(1): 1206-1209.

