

Differentiation of Myeloid Cell Precursors in Activated Gene-Engineered Dendritic Cells with Anti-Malignant Properties

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Research Article

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Abstract

Studies on the biology of dendritic cells (DCs) are mainly focused on their role as immune activators and modulators. In their appropriate cultivation and/or modifications, they have shown abilities for an enhanced expression of specific effective molecules. These cells have also been characterized as powerful antigen-presenting cells (APCs), including in composition of anti-malignant vaccines and gene-engineering products. By appropriate cultivation and/or modifications of DCs, they have shown abilities for enhanced expression of specific molecules. These properties have characterized them as promising candidates for construction of novel safe vaccines and gene-engineering products on their basis. In this aspect, in the last years the attention is directed to development of new safe therapeutic methods and techniques with DCs.

Keywords: Dendritic Cells (Dcs); Stem/Progenitor Cells; Myeloid Cell Differentiation; Recombinant Viral Vectors

Introduction

Dendritic cells (DCs) have been found to play a pivotal role in the processess of immune response initiation and modulation, mainly as powerful antigen-presenting cells (APCs) [1-17]. On the other hand, they have been found to participate in the maintenance of peripheral tolerance, and their capacity to induce anti-nuclear auto-immune response has been proven in experimental models [18].

Biological Properties of Dendritic Cells and Their Role in Generation of Adequate Immune Response

In the light of the unique properties of DCs, they have been proposed as powerful immunomodulation agents, including in the composition of novel vaccines and gene-engineering products for treatment of malignant disorders [19-37]. Complex mechanisms, which include molecular, genetic and cellular components, such as *Wnt-*, *BMP-* and *Notch/ Delta-*signaling pathways, have been found to underlie differentiation and functions of stem and progenitor cells [38-43]. By use of polymerase chain reaction in real time (RT-PCR), an ability for initiation of erythroid (β-globin) and/or myeloid (myeloperoxidase) gene expression programs by the same cell prior to exclusive commitment to the erythroid and/or, respectively, myeloid lineages for it, has been shown [44]. On the other hand, protein BCL-6 has also been detectable in inter- and intra-follicular CD4+ T-lymphocytes, but not in other follicular components, including B-lymphoid cells, plasma cells, monocytes/macrophages and DCs [45,46].

Normal Differentiation of Myeloid Cell Precursors

According many literature data, granulocytemacrophage colony-stimulating factor (GM-CSF) mobilizes CD34+ bone-marrow progenitor cells both in vitro and in vivo with an increased frequency and generation of DCs with anti-malignant properties [46-48,]. Similarly, the addition of GM-CSF plus tumor necrosis factor- α (TNF- α), induced development of DCs from purified CD34+ cells of bone marrow, cord blood and peripheral blood has been observed. The critical role of TNF- α for the differentiation of DCs has been supported by the demonstration that this cytokine induces the expression of molecule CD40 on CD34+ cells. Besides that, CD34+/CD40+ cells have been found to express only myeloid markers, significantly increase allo-antigen presenting function, compared with total CD34+ cells, and have also given rise to DCs' number. Capable of modulating

differentiation of DCs from these bipotent CD34+/CD40+ cells during the later stages of their cultivation, has also been shown cytokine interleukin-4 (IL-4) [16].

The possibility of generating or expanding tolerant DCs ex vivo has been found to open novel therapeutic perspectives. They're in vitro and/or ex vivo-maturation have been characterized as a critical step in the induction of T-cell responses and it has been proven to depend on the activation of transcription factors from the family of Nuclear FactorkappaB (NF- κ B) [12]. It has also been suggested that kinetic and the quality of DCs' activation is controlled by cytokine IL-10, which has been characterized as alternative promising pathway of their differentiation [30]. On the other hand, DCs, differentiated in the presence of vaso-active intestinal peptide (VIP), have shown impaired allogeneic haplotypespecific responses of donor CD4+ T-lymphocytes in mice, and have been found to induce generation of regulatory T-cells in the graft [49-52]. As a critical component for optimal function of DCs, has been characterized the TNF supperfamily member lymphotoxin- $\alpha\beta$ (LT $\alpha\beta$), independently of its described role in maintaining of the lymphoid tissue organization [53]. In the absence of $LT\alpha\beta$ on antigen-specific T-cells, DCs' dysfunction in vivo could be rescued via CD40 or $LT\alpha\beta$ receptor stimulation, respectively, which has suggested a possibility for eventual cooperation of these pathways. It has also been indicated that DCs, induced by ligand Flt3, are well positioned to regulate the qualitative nature of intestinal immune responsiveness, depending on the presence or absence of appropriate inflammatory signals [15]. In this way, a potential use of ligand Flt3 as a mucosal vaccine adjuvant in conjunction with the inflammatory mediator IL-1 has been suggested.

Abnormal Differentiation of Myeloid Cell Precursors

Myeloproliferative disorders have been defined as clone malignancies in hematopoietic stem cells (HSCs), characterized by their independency and/or hypersensitivity to numerous cytokines and/or growth factors [54]. In many patients with such diseases, a mutation in such gene has been established, and its presence in erythropoieticindependent erythroid colonies has demonstrated a link with the hypersensitivity to the respective growth factor or cytokine, which has been defined as a key biologic feature of these disorders. It has been concluded that active cellular cycles of bone-marrow cells, induced by appropriate cytokine stimulation, could be associated with an engraftment defect in the normal host, a respective consequent rearrangement of these pathways within the stem cells, as well as with an apparent lineage and differentiation status, which has been found to play an important role in the development of malignancies [55,56,]. In investigation on the role of

the cytoplasmic tyrosine-kinase JAK2 in patients with myelofibrosis, the mutation *Val617PHE JAK2* in this gene has been presented in erythropoietin-independent erythroid colonies, which has also been found in patients with other myeloproliferative diseases, and its presence in patients with these disorders has been found to lay the foundation of new approaches for classification, diagnosis, profilactic and treatment of malignant diseases [57].

Development Of Novel Therapeutic Strategies with Dendritic Cells

In the last years, the development of novel therapeutic strategies with DCs has become extensively investigated [58-62]. The antigen-presenting properties of DCs could be exploited as a new therapeutic tool in the therapy of malignancies for to enhance the immunity against respective malignancy-specific antigens. After such application of DCs, peptide-specific responses by cytotoxic T-lymphocytes (CTLs), improvement in performance status, decrease in malignancy markers levels, regression of malignancies, and, at the same time, no toxic side effects have been accounted [13]. Because isolated DCs, loaded with malignant antigens ex vivo and administered as effective cellular vaccines, have induced protective and therapeutic anti-malignant immunity in experimental animals with induced malignant disorders, adjuvant treatment of malignancies at high risk for recurrence after operation, as well as methods for targeting malignant antigens to DCs in vivo, have been explored [16]. On the other hand, appropriate modifications of DCs to express malignancy-specific antigens by in vitro and/or ex vivo-transfer of genes, coding respective antibodies, has been suggested. So genetically modified DCs have been widely tested in pre-clinical studies, including as anti-malignant agents. Besides that, genetically-modified DCs have been widely tested in many pre-clinical studies, including as anti-malignant agents. Therefore, exploitation of the antigen-presenting properties of DCs offers promise for the development of effective anti-malignant immunotherapy. Taken together, these data have revealed abilities for development of novel and safe therapeutic strategies with DCs.

Development of Elimination and Differentiation Therapy

Elimination and differentiation therapy of malignant stem cells has been shown to increase the efficacy against malignant diseases and disorders [63-66]. This therapeutic method could be applied in types of leukemia, which have shown a poor prognosis with conventional therapeutic treatment. As the most common example, the transplantation of allogenic HSCs with graft versus malignancy effect has been characterized.

Development of Novel Therapeutic Strategies with Hybrid Vaccine Constructs, Received by Fusion of Dendritic Cells with Malignant Cells

As alternative method for delivery into DCs, their fusion with malignant cells has been utilized, as well as the hybrid cell-based vaccines have shown high therapeutic activity, even in patients with malignant diseases and disorders. The immunization with such hybrid conjugates, derived by fusion between DCs and malignant cells, has significantly increased the production of Th1 cytokine-producing cells, the number of antigen-specific CD8+ T-cells, as well as the anti-malignant immunity. The observed anti-malignant immunity, induced by vaccination with DCs/malignant cells hybrid fusion products has reacted differently to injected malignant cells and autochthonous malignancies [60]. It has also been shown that immunization with such fusion cells induces rejection of metastases. Hybrid cells, obtained by fusion between DCs and malignant cells, have been found to express major histo-compatibility complex (MHC) molecules, both class I- and class II-restricted malignancy-associated epitopes and might, therefore, be useful for the induction of specific malignancy-reactive CD8+ and CD4+ T-lymphocytes both in vitro and in vivo, by human vaccination trials [53]. The observed greatly reduced number of established pulmonary metastases both with and without in vivo-administration of IL-2-adoptive transfer of T-lymphocytes, derived from B16/ DCs vaccine-primed lymph nodes into B16 tumor-bearing mice, has suggested a role of malignant cells/DCs hybrids as effective cellular vaccines for eliciting T-cell-mediated antimalignant immunity [52]. It has also been demonstrated an enhanced immunization with received by fusion of DCs with mouse 4T00 plasmacytoma cells FC/4T00 hybrid cells, plus anti-malignant immunity of IL-12 [58]. Findings, according which fusions of ovarian cancer cells to autogenic or allogeneic DCs induce cytolytic T-cell activity and lysis of autologous malignant cells, mediated by MHC molecules class I-restricted mechanism, have suggested that the fusions are probably functional, when are generated by autogenic and/or allogeneic transplantation of DCs [53]. It has also been demonstrated that sequential stimulation with DCs/ breast carcinoma cells fusion hybrids results in a marked expansion of activated malignancy-specific T-lymphocytes, which has suggested these fusion cells are probably effective APCs, which stimulate inhibitory T-cells that limit vaccine efficacy [17-42]. Similarly, hybrids, derived by fusion of spleen DCs from C57BL/6 mice with B16 melanoma cells, have expressed MHC-molecule B7, as well as B16 tumor marker M562, and have been characterized as an attractive strategy for immunotherapy of malignancies [2]. On the other hand, the results, according which the ex vivo-exposure of DCs in presence of cytokine transforming growth factorbeta (TGF- β) hasn't appeared to lessen the efficacy of DCs vaccines, have suggested that this cytokine, derived from

malignant cells, has probably reduced the their efficacy via in vivo-mechanism, and the neutralization of produced by the fusion cells TGF- β might enhance it [8-43]. An increase in the immunogenic potential of DCs/malignant cells fusion cellbased vaccines has been observed in heat-treated malignant cells [5].

Development of Novel Therapeutic Strategies with Dendritic Cells, Transduced by Recombinant Viral Vectors, Coding Malignant Antigens

DCs have shown a possibility to be genetically engineered to express constitutively respective genes of interest, coding immune-modulating cytokines, antibodies and/or antigens, derived from transformed cells or other pathogens 11 [19-48]. In laboratory conditions, human DCs, transduced with recombinant adenoviral vectors, have shown inhibition of a mixed leukocyte culture, reduced cell surface expression of co-stimulatory molecules CD80and CD86, as well as inability for production of the potent allo-stimulatory cytokine IL-12. In investigation on the in vivo-properties of the so modified DCs, skin transplantation of experimental mice with nonobese diabetes, combined with severe immunodeficiency (NOD/SCID), reconstituted via intra-peritoneal injection with allogeneic mononuclear cells (MNCs), mixed with autologous to the skin donor DCs, transduced with either recombinant adenoviral gene construct AdV/IL-10 or AdV/ MX-17, a reduced skin graft rejection, characterized by reduced mononuclear cell infiltration and less destruction of junctions between derma and epidermis, in comparison with the animals with inoculation of DCs [22]. Adenovirustransduced immature DCs have shown ability to differentiate in the presence of lipopolysaccharide (LPS) or a monocyte-/ macrophage-conditioned medium to express the surface markers of mature DCs, such as CD25, CD83, high levels of molecules CD86 and HLA-DR, as well as to secrete of IL-12. Their ability to induce T-lymphocytes' growth has also been enhanced. It has also been suggested that adenoviruses probably have mediated minor effects on the phenotype of DCs, which, however, could be seen only when a sufficient number of particles enter in each cell [48]. Recombinant adenoviral vectors have also been found to transduce effectively DCs and direct the generation of specific CTLs, which would be a potent strategy in the immunotherapy of Hodgkin's lymphoma [23]. According the results from other study, transduction of DCs with recombinant vectors with insertion of gene mTRP-2, (encoding tyrosinase-related protein-2, respectively), provides a potential therapeutic strategy for the management of melanoma, especially in the early stage of that disease [30]. So modified DCs have also shown high stimulatory activity in both allogeneic and autogenic mixed lymphocyte reaction. Similarly, mouse DCs, infected with recombinant fowlpox virus (rFWPV) vector,

have stimulated a powerful, MHC class I-restricted immune response against the recombinant antigen [1]. These data have also supported the efficiency of the recombinant viral vectors in studies on the biologic properties of DCs, including the expression of specific antigens for active immune therapy.

Development of Combined Therapeutic Strategies with Dendritic Cells

The fact that an increased Th1 cytokine production and stronger anti-malignant effect haven't been observed in mice, depleted of gamma-interferon (IFN- γ), has also supported the maintenance of DCs/malignant cells conjugates as potent anti-malignant vaccines, as well as the cytokine IL-18 [17-42]. These data could be additionally administrated by gene transfection of cells for enhancement of the immunity, which is probably mediated mainly by IFN- γ .

Development of Combined Therapeutic Strategies with Gene-Engineering of Dendritic Cells

For further increase of the potency of the vaccine, a combined variation of both technologies has been applied, in which *IL-18*-transfected DCs have been used to prepare DCs/ malignant cells conjugates. It has also been indicated that *GM*-*CSF* gene-modified DCs might lead to the generation of hybrid vaccines with potentially increased therapeutic efficacy [2]. Although the observed elicited anti-malignant effect with participation of both CD4+ and CD8+ T-lymphocytes by the hybrid vaccine *IL18DC-E.G7*, derived by fusion between geneengineered DCs, transduced with recombinant *adenoviral* vectors, carrying genes for β -galactosidase (*AdlacZ*) and/ or IL-18 (*AdIL18*), respectively, and *E.G7* malignant cells, derived by *EL4* cells, transfected with *cDNA*, carrying gene for chicken egg albumin, it has been largely blocked by anti-IFN- γ antibodies [29].

Development of Combined Therapeutic Strategies with Gene-Engineering of Malignant Cells

Results from experiments for immunization with fusion hybrids, derived by fusion of DCs with *IL-12* gene-transferred malignant cells, have shown an ability to elicit a previously enhanced anti-malignant effect in experimental therapeutic models. Such novel IL-12-producing fusion cell vaccine has been characterized as a promising intervention for future immune therapy of malignant diseases [15].

Development of Combined Therapeutic Strategies with Gene-Engineering of Both Malignant and Dendritic Cells

In immunization of mice with gene-engineered

DCRMAT/J558-IL-4 fusion hybrids, an elicited stronger *J558* tumor-specific CTLs immune response has been induced, in comparison of hybrid vaccine *DCRMAT/J558 in vivo* [39]. Similar results have been observed in immunization of C57BL/6 mice with gene-engineered *DC/J558-IL-4* hybrids, and gene-engineered fusion hybrid vaccine constructs have been characterized as an attractive strategy for immunotherapy of malignancies [66-68].

Conclusion

Dendritic cells (DCs) have been characterized as hopeful vehicles for appropriate modulation of the immune response, including in composition of vaccine constructs and gene-engineered products with anti-malignant activity. They have also shown abilities for enhanced expression of specific molecules in appropriate conditions of cultivation and/or by appropriate modifications. These properties characterize them as promising candidates for construction of novel and safe therapeutic products on their basis, by use of new technologies, as their fusion with malignant cells; transduction with recombinant viral vectors, as well as a combined variation, in which malignant cell, DC or both components of the hybrid fusion vaccine might be genetically transduced.

References

- Ahuja SS (2001) Genetic engineering of dendritic cells using retrovirus-based gene transfer techniques. Meth Mol Biol 156: 79-87.
- 2. Aicher A, Westermann J, Cayeux S, Willimsky G, Daemen K, et al. (1997) Successful retroviral mediated transduction of a reporter gene in human dendritic cells: feasibility of therapy with gene-modified antigen presenting cells. Exp Hematol 25(1): 39-44.
- 3. Arthur J, Butterfield L, Roth MD, Bui LA, Kierstscher SM, et al. (1997) Cancer Gene Ther 4: 1023-1028.
- 4. Avigan D (2004) Dendritic cell-tumor fusion vaccines for renal cell carcinoma. Clin Cancer Res 10(18 Pt 2): 6347S-6352S.
- Ballas CB, Zielske SP, Gerson SL (2002) Adult bone marrow stem cells for cell and gene therapies: implications for greater use. J Cell Biochem Suppl 38: 20-28.
- 6. Banchereau J, Briere F, Caux C, Davoust J, Lebesque S, et al. (2000) Ann Rev Immunol 18: 767-811.
- Baxter EJ, Scott LM, Campbell PJ, East C, Fourouclas N, et al. (2005) Acquired mutation of the tyrosine kinase JAK2 in human myeloproliferative disorders. Lancet 365(9464): 1054-1061.

- 8. Bonini C, Lee SP, Riddell SR, Greenberg PDJ (2001) Targeting Antigen in Mature Dendritic Cells for Simultaneous Stimulation of CD4+ and CD8+ T Cells. Immunol 166(8): 5250-5257.
- 9. Brown M, Zhang Y, Dermine S, de Wynter EA, Hart C, et al. (2000) Dendritic cells infected with recombinant fowlpox virus vectors are potent and long-acting stimulators of transgene-specific class I restricted T lymphocyte activity. Gene Ther 7(19): 1680-1689.
- 10. Bubenik J (2001) Genetically engineered dendritic cellbased cancer vaccines. Int J Oncol 18(3): 475-478.
- 11. Cao X, Zhang W, Wang J, Zhang M, Huang X, et al. (1999) Immunology 97(4): 616-625.
- 12. Caux C, Dezutter-Dambuyant S, Schmitt D, Banchereau J (1992) GM-CSF and TNF-alpha cooperate in the generation of dendritic Langerhans cells. Nature 360(6401): 258-261.
- Chaussabel D, Banchereau J (2005) Dendritic cells, therapeutic vectors of immunity and tolerance. Am J Transplant 5(2): 205-206.
- 14. Cheng L, Du C, Lavau C, Chen S, Tong J, et al. (1998) Sustained gene expression in retrovirally transduced, engrafting human hematopoietic stem cells and their lympho-myeloid progeny. Blood 92(1): 83-92.
- 15. Chorny A, Gonzalez-Rey E, Fernandez-Martin A, Ganea D, Delgado M (2006) Vasoactive intestinal peptide induces regulatory dendritic cells that prevent acute graftversus-host disease while maintaining the graft-versustumor response. Blood 107(9): 3787-3794.
- Clark EA, Grabstein K, Shu GL (1992) J Immunol 148(11): 3327-3335.
- 17. Coates PTH, Krishnan R, Kireta S, Johnston J, Russ GR (2001) Human myeloid dendritic cells transduced with an adenoviral interleukin-10 gene construct inhibit human skin graft rejection in humanized NOD-scid chimeric mice. Gene Ther 8(16): 1224-1233.
- 18. Crispin JC, Alcocer-Varela J (2007) The role myeloid dendritic cells play in the pathogenesis of systemic lupus erythematosus. Autoimmun Rev 6(7): 450-456.
- 19. Curti A, Fogli M, Ratta M, Biasco G, Tura S, et al. (2001) J Biol Reg Homeostat Agents 15: 49-52.
- 20. Dietz AB, Vuk-Pavlović S (1998) High efficiency adenovirus-mediated gene transfer to human dendritic cells. Blood 91(2): 392-398.

- Engelmayer J, Larsson M, Lee A, Lee M, Cox W, et al. (2001) Mature dendritic cells infected with canarypox virus elicit strong anti-human immunodeficiency virus CD8+ and CD4+ T-cell responses from chronically infected individuals. J Virol 75(5): 2142-2153.
- 22. Fong L, Engelman EG (2000) Dendritic cells in cancer immunotherapy. Ann Rev Immunol 18: 245-273.
- 23. Frasca L, Fedele G, Deaglio S, Capuano C, Palazzo R, et al. (2006) CD38 orchestrates migration, survival, and Th1 immune response of human mature dendritic cells. Blood 107(6): 2392-2399.
- 24. Gahn B, Siller-Lopez F, Pirooz AD, Yvon E, Gottschalk S, et al. (2001) Adenoviral gene transfer into dendritic cells efficiently amplifies the immune response to LMP2A antigen: a potential treatment strategy for Epstein-Barr virus--positive Hodgkin's lymphoma. Int J Cancer 93(5): 706-713.
- 25. Gong J, Chen D, Kashiwaba M, Kufe D (1997) Induction of antitumor activity by immunization with fusions of dendritic and carcinoma cells. Nat Med 3(5): 558-561.
- 26. Gong J, Chen L, Chen D, Kashiwaba M, Manome Y, et al. (1997) Induction of antigen-specific antitumor immunity with adenovirus-transduced dendritic cells. Gene Ther 4(10): 1023-1028.
- 27. Gong J, Koido S, Chen D, Tanaka Y, Huang L, et al. (2002) Immunization against murine multiple myeloma with fusions of dendritic and plasmacytoma cells is potentiated by interleukin 12. Blood 99(7): 2512-2517.
- 28. Gong J, Nikrui N, Chen D, Koido S, Wu Z, et at. (2000) Fusions of human ovarian carcinoma cells with autologous or allogeneic dendritic cells induce antitumor immunity. J Immunol 165(3): 1705-1711.
- 29. Gonzalez-Rey E, Chorny A, Fernandez-Martin A, Ganea D, Delgado M (2006) vasoactive intestinal peptide generates human tolerogenic dendritic cells that induce CD4 and CD8 regulatory T cells. Blood 107(9): 3632-3638.
- 30. Hiraoka K, Yamamoto S, Otsuru S, Nakai S, Tamai K, et al. (2004) Enhanced tumor-specific long-term immunity of hemagglutinating [correction of hemaggluttinating] virus of Japan-mediated dendritic cell-tumor fused cell vaccination by coadministration with CpG oligodeoxynucleotides. J Immunol 173(7): 4297-4307.
- 31. Inaba K, Inaba M, Romani N, Aya H, Deguchi M, et al. (1992) Generation of large numbers of dendritic cells from mouse bone marrow cultures supplemented with

granulocyte/macrophage colony-stimulating factor. J Exp Med 176(6): 1693-170228.

- 32. Ju DW, Tao Q, Lou G, Bai M, He L, et al. (2001) Interleukin 18 transfection enhances antitumor immunity induced by dendritic cell-tumor cell conjugates. Cancer Res 61(9): 3735-3740.
- 33. Kao JY, Gong Y, Chen CM, Zheng QD, Chen JJ (2003) Tumorderived TGF-beta reduces the efficacy of dendritic cell/ tumor fusion vaccine. J Immunol 170(7): 3806-3811.
- Kaplan J, Yu Q, Piraino S, Pennington SE, Shankara S, et al. (1999) Induction of antitumor immunity with dendritic cells transduced with adenovirus vector-encoding endogenous tumor-associated antigens. J Immunol 163(2): 699-707.
- 35. Kim HS, Zhang X, Choi YS (1994) Activation and proliferation of follicular dendritic cell-like cells by activated T lymphocytes. J Immunol 153(7): 2951-1961.
- 36. Koido S, Hara E, Homma S, Mitsunaga M, Takahara A, et al. (2007) Synergistic induction of antigen-specific CTL by fusions of TLR-stimulated dendritic cells and heat-stressed tumor cells. J Immunol 179(7): 4874-4883.
- Le Blannc K, Ringden O (2005) Immunobiology of human mesenchymal stem cells and future use in hematopoietic stem cell transplantation. Biol. Blood Marrow Transplant 11(5): 321-334.
- 38. Lessard J, Sauvageau G (2003) Bmi-1 determines the proliferative capacity of normal and leukaemic stem cells. Nature 423(6937): 255-260.
- 39. Liu Y (2003) Leuk Res 26(8): 757.
- 40. Lu L, et al. (1998) Transplantation 65: 757.
- 41. Massard C, Deutsch E, Soria JC (2006) Tumour stem celltargeted treatment: elimination or differentiation. Ann Oncol 17(11): 1620-1624.
- 42. Metharom P, Ellem KAO, Schmidt C, Wei MQ (2001) Hum Gene Ther 12: 2203-2213.
- 43. Moore F, Buonocore S, Aksoy E, Ouled-Haddou N, Goriely S, et al. (2007) An Alternative Pathway of NFκB Activation Results in Maturation and T Cell Priming Activity of Dendritic Cells Overexpressing a Mutated IκBα. J Immunol 178(3): 1301-1311.
- 44. Pardal R, Clarke MF, Morrison SJ (2003) Applying the principles of stem-cell biology to cancer. Nat Rev Cancer 3(12): 895-902.

- 45. Parkhurst MR, De Pan C, Riley JP, Rosenberg SA, Shu S (2003) Hybrids of dendritic cells and tumor cells generated by electrofusion simultaneously present immunodominant epitopes from multiple human tumor-associated antigens in the context of MHC class I and class II molecules. J Immunol 170(10): 5317-5325.
- 46. Passegue E, Jamieson CH, Ailles LE, Weissman H (2003) Normal and leukemic hematopoiesis: are leukemias a stem cell disorder or a reacquisition of stem cell characteristics? PNAS USA 100(1): 11842-11849.
- 47. Perona-Wright G, Anderton SM, Howie SEM, Gray D (2007) IL-10 permits transient activation of dendritic cells to tolerize T cells and protect from central nervous system autoimmune disease. Int Immunol 19(9): 1123-1134.
- 48. Reid CDL, Stackpole A, Meager A, Tikepae J (1992) J Immunol 149: 2681-2688.
- 49. Ribas A, Butterfield L, McBride W, Jilani SM, Bui LA, et al. (1997) Cancer Res 57: 2865-2869.
- 50. Sadanaga N, Nagashima H, Mashino K, Tahara K, Yamaguchi H, et al. (2001) Dendritic cell vaccination with MAGE peptide is a novel therapeutic approach for gastrointestinal carcinomas. Clin Cancer Res 7(8): 2277-2284.
- 51. Sell S (2004) Stem cell origin of cancer and differentiation therapy. Crit Rev Oncol Hematol 51(1): 1-28.
- 52. Siena S, Di Nicola M, Bregni M, Mortarini R, Anichini A, et al. (1995) Massive ex vivo generation of functional dendritic cells from mobilized CD34+ blood progenitors for anticancer therapy. Exp Hematol 23(14): 1463-1471.
- 53. Summers-deLuca LE, McCarthy DD, Cosovic B, Ward LA, Lo CC, et al. (2007) Expression of lymphotoxin-alphabeta on antigen-specific T cells is required for DC function. J Exp Med 204(5): 1071-1081.
- 54. Suzuki T, Fukuhara T, Tanaka M, Nakamura A, Akiyama K, et al. (2005) Vaccination of dendritic cells loaded with interleukin-12-secreting cancer cells augments in vivo antitumor immunity: characteristics of syngeneic and allogeneic antigen-presenting cell cancer hybrid cells. Clin Cancer Res 11(1): 58-66.
- 55. Terkikh AV, Bryant PJ, Schwartz PH (2006) Mammalian Stem Cells. Pediatric Res 59: 13-20.
- 56. Tillman BW, de Gruijl TD, Luykx-de Bakker SA, Scheper RJ, Pinedo HM, et al. (1999) Maturation of dendritic cells accompanies high-efficiency gene transfer by a CD40-

targeted adenoviral vector. J Immunol 162(11): 6378-6383.

- 57. Timmerman JM, Levy R (1999) dendritic cell vaccines for cancer immunotherapy. Annu Rev Med 50: 507-529.
- 58. Vasir B, Wu Z, Crawford K, Rosenblatt J, Zarwan C, et al. (2008) Fusions of Dendritic Cells with Breast Carcinoma Stimulate the Expansion of Regulatory T Cells while Concomitant Exposure to IL-12, CpG Oligodeoxynucleotides, and Anti-CD3/CD28 Promotes the Expansion of Activated Tumor Reactive Cells. J Immunol 181(1): 808-821.
- 59. Walden P (2000) Adv Exp Med Biol. 465: 347-354.
- 60. Wang J, Saffold S, Cao X, Krauss J, Chen W (1998) Eliciting T cell immunity against poorly immunogenic tumors by immunization with dendritic cell-tumor fusion vaccines. J Immunol161(10): 5516-5524.
- 61. Wen Ju D, Tao Q, Lou G, Bai M, He L, et al. (2001) Interleukin 18 Transfection Enhances Antitumor Immunity Induced by Dendritic Cell-Tumor Cell Conjugates. Cancer Res 61: 3735-3740.
- 62. Williamson E, Westrich GM, Viney JL (1999) Modulating

dendritic cells to optimize mucosal immunization protocols. J Immunol 163(7): 3668-3675.

- 63. Xia D, Li F, Xiang J (2004) Engineered fusion hybrid vaccine of IL-18 gene-modified tumor cells and dendritic cells induces enhanced antitumor immunity. Cancer Biother Radiopharm 19(3): 322-330.
- 64. Xia D, Chan T, Xiang J (2005) Dendritic cell/myeloma hybrid vaccine. Meth Mol Med 113 225-233.
- 65. Xia J, Tanaka Y, Koido S, Liu C, Mukherjee P, et al. (2003) Prevention of spontaneous breast carcinoma by prophylactic vaccination with dendritic/tumor fusion cells. J Immunol 170(4): 1980-1986.
- 66. Yongqing Z, Chan WT, Saxena A, Xiang J (2002) Leukemia Res 26(18): 757-763.
- 67. Zhang XY, La Russa VF, Reiser J (2004) Transduction of bone-marrow-derived mesenchymal stem cells by using lentivirus vectors pseudotyped with modified RD114 envelope glycoproteins. J Virol 78(3): 1219-1229.
- Zhong L, Granelli-Piperno A, Choi Y, Steinman RM (1999) Recombinant adenovirus is an efficient and nonperturbing genetic vector for human dendritic cells. Eur J Immunol 29(3): 964-972.

